

Formulation and Optimization of Liposomal and Niosomal Systems for Enhanced Drug Delivery in Diabetes Disease

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Abstract

Diabetes mellitus represents a global health crisis affecting over 422 million people worldwide, with approximately 75 million requiring daily insulin injections. Current subcutaneous insulin delivery methods suffer from poor patient compliance, injection-site reactions, and risk of hypoglycemic episodes. Vesicular drug delivery systems, particularly liposomes and niosomes, have emerged as promising alternatives for enhanced diabetes management. This comprehensive review examines the formulation strategies, optimization parameters, and comparative efficacy of liposomal and niosomal systems for diabetic drug delivery. Recent advances in nanotechnology have enabled the development of pH-responsive, glucose-sensitive formulations that offer sustained release profiles and improved bioavailability. Comparative studies demonstrate that while both systems show significant therapeutic potential, niosomes exhibit superior stability against enzymatic degradation, with advantages in terms of chemical stability and cost-effectiveness. The optimization of phospholipid-to-cholesterol ratios (typically 5:1 to 9:1) and drug-to-lipid ratios (1:5 to 1:10) significantly influences encapsulation efficiency and release kinetics. Current clinical developments, including oral insulin formulations reaching human trials in 2025, highlight the translational potential of these technologies. This review provides crucial insights for researchers and clinicians working toward next-generation diabetes therapies.

Keywords: *Liposomes, Niosomes, Diabetes mellitus, Insulin delivery, Drug formulation, Nanotechnology*

1. INTRODUCTION

Diabetes mellitus is characterized as a chronic endocrine and metabolic disorder marked by hyperglycemia and multiple complications. The global prevalence continues to rise dramatically, with type 2 diabetes mellitus (T2DM) representing approximately 90% of all cases. Traditional therapeutic approaches rely heavily on subcutaneous insulin injection, which presents significant challenges including poor patient compliance, injection-site complications, and risk of severe hypoglycemic events.

The limitations of conventional diabetes management have driven extensive research into alternative drug delivery systems. Nanotechnology-based approaches, particularly vesicular carriers such as liposomes and niosomes, offer promising solutions to overcome the inherent challenges of insulin and other antidiabetic

drug delivery. These systems provide protective encapsulation against enzymatic degradation, enhanced cellular uptake, and the potential for targeted, sustained release profiles.

Recent breakthrough research published in *Nature Nanotechnology* has demonstrated the successful development of oral insulin formulations using silver sulfide quantum dots coated with chitosan/glucose polymers, achieving glucose-responsive insulin release without hypoglycemic episodes in animal models. This advancement represents a paradigm shift toward patient-friendly diabetes management and highlights the critical importance of optimized nanocarrier systems.

2. Pathophysiology of Diabetes and Current Treatment Challenges

2.1 Diabetes Mellitus Classification and Mechanisms

Type 1 diabetes mellitus (T1DM) results from autoimmune destruction of pancreatic β -cells, leading to absolute insulin deficiency. Patients require lifelong exogenous insulin replacement to maintain glucose homeostasis. Type 2 diabetes mellitus (T2DM) involves progressive insulin resistance combined with relative insulin deficiency, initially managed through oral hypoglycemic agents but often progressing to insulin dependence.

2.2 Limitations of Conventional Therapy

Current subcutaneous insulin delivery faces multiple significant limitations that compromise patient outcomes and quality of life. Daily multiple injections result in poor adherence rates, particularly among pediatric and elderly populations who struggle with the physical and psychological burden of repeated injections. Injection-related complications include pain, discomfort, lipodystrophy at injection sites, and increased risk of local infections. Unpredictable insulin absorption patterns can lead to life-threatening hypoglycemic episodes, representing one of the most serious acute complications of diabetes management. The non-physiological insulin delivery patterns achieved through subcutaneous injection fail to mimic the precise temporal patterns of endogenous insulin secretion, resulting in suboptimal glycemic control. Furthermore, the social stigma and lifestyle restrictions associated with injection dependence significantly impact patient quality of life and may contribute to treatment discontinuation.

2.3 Barriers to Oral Insulin Delivery

Oral insulin administration faces substantial physiological barriers that have historically prevented successful clinical implementation. The harsh gastric environment, characterized by extremely low pH conditions, rapidly denatures insulin's tertiary structure and renders it biologically inactive. Enzymatic proteolysis by digestive enzymes including pepsin, trypsin, and chymotrypsin systematically degrade insulin's peptide bonds throughout the gastrointestinal tract. The large molecular weight of insulin combined with its hydrophilic nature results in poor intestinal permeability, with minimal absorption across the intestinal epithelium. Additionally, extensive hepatic first-pass metabolism further reduces the bioavailability of any insulin that successfully crosses the intestinal barrier, making oral delivery highly inefficient without appropriate pharmaceutical interventions.

3. Liposomal Drug Delivery Systems

3.1 Structural Characteristics and Advantages

Liposomes are spherical vesicles composed of phospholipid bilayers surrounding an aqueous core, with diameters ranging from 25 nm to several micrometers depending on preparation method and intended application. Their unique amphiphilic nature enables the simultaneous encapsulation of both hydrophilic drugs within the aqueous core and hydrophobic compounds within the lipid bilayer matrix. The biocompatibility of liposomes stems from their composition using naturally occurring phospholipids that are metabolized through normal physiological pathways without generating toxic byproducts.

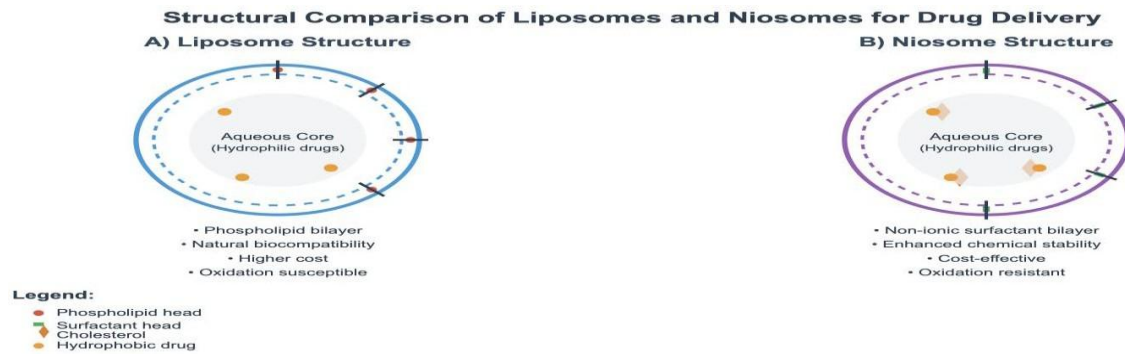


Figure 1: Structural Comparison of Liposomes and Niosomes

Their biodegradable nature ensures complete elimination from the body through established metabolic processes, while their low immunogenicity minimizes adverse immune system activation even with repeated administration. The versatile drug loading capacity of liposomes accommodates diverse pharmaceutical compounds with varying physicochemical properties, and their potential for surface modification enables the development of cell-specific targeted delivery systems that can enhance therapeutic specificity while reducing off-target effects.

3.2 Liposomal Insulin Formulations

Recent research has demonstrated significant advances in insulin-loaded liposomal systems for oral delivery. Conventional liposomes and seven novel formulation types have been extensively studied for diabetes applications:

3.2.1 Conventional Insulin Liposomes

Basic liposomal formulations using phosphatidylcholine and cholesterol have shown protective effects against enzymatic degradation in simulated gastric and intestinal fluids. Studies demonstrate sustained insulin release profiles extending 8-24 hours, with bioavailability improvements of 8-10% compared to subcutaneous injection.

3.2.2 Surface-Modified Liposomes

PEGylation of liposomal surfaces provides "stealth" properties, reducing mononuclear phagocyte system (MPS) clearance and extending circulation time. DSPE-PEG conjugates are preferred due to enhanced membrane stability from longer acyl chains.

3.2.3 pH-Responsive Liposomes

Formulations incorporating pH-sensitive polymers enable selective drug release in specific gastrointestinal environments. These systems remain stable in acidic gastric conditions but release insulin in the neutral pH of the small intestine.

3.3 Optimization Parameters for Liposomal Systems

3.3.1 Phospholipid-to-Cholesterol Ratios

Optimization studies consistently demonstrate that phospholipid-to-cholesterol ratios significantly influence liposomal characteristics:

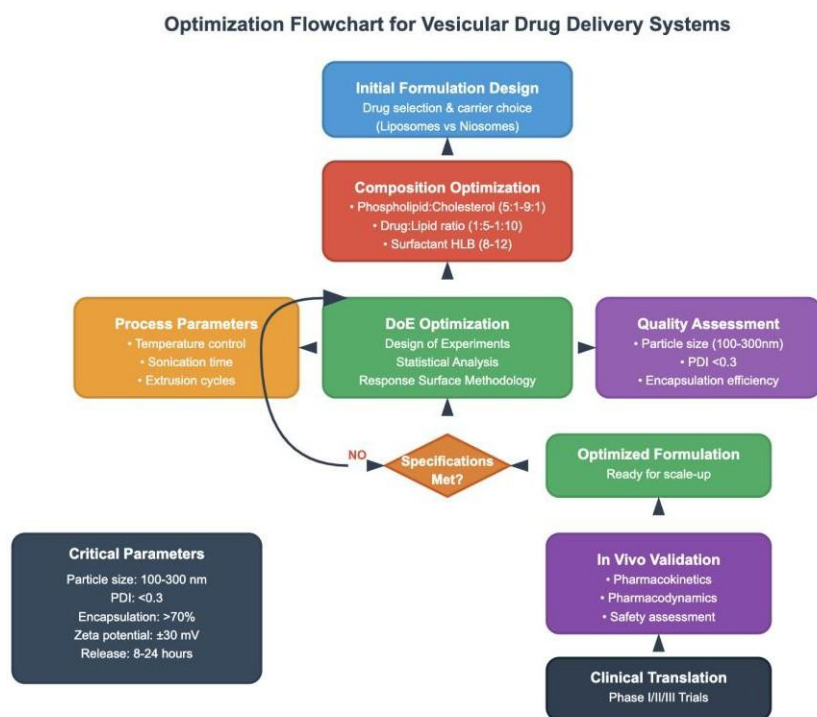


Figure 2: Optimization Flowchart for Vesicular Drug Delivery Systems

- **5:1 Ratio:** Optimal balance of stability and drug loading capacity
- **9:1 Ratio:** Maximum drug encapsulation efficiency for hydrophobic compounds
- **Higher Cholesterol Content (>30%):** Increased membrane rigidity but reduced drug loading

Research indicates that 70:30% phospholipid-to-cholesterol ratios provide the most stable formulations for controlled drug release across diverse pharmaceutical applications.

3.3.2 Drug-to-Lipid Ratios

Drug-to-lipid ratios critically affect encapsulation efficiency and release kinetics:

- **1:5 Ratio:** Demonstrated highest encapsulation efficiency (>90%) in multiple studies
- **1:10 Ratio:** Provides extended release profiles but lower drug loading
- **Higher Drug Content:** Results in decreased encapsulation efficiency (~40%) and potential membrane destabilization

3.4 Preparation Methods and Quality Control

3.4.1 Thin-Film Hydration Method

The most widely employed preparation technique involves a systematic four-step process that ensures consistent vesicle formation and drug encapsulation. Initially, lipids are completely dissolved in an appropriate organic solvent such as chloroform or methanol to create a homogeneous lipid solution. The solution then undergoes rotary evaporation under reduced pressure to remove the organic solvent and form a uniform thin lipid film on the flask walls. Subsequently, the dried lipid film is hydrated with aqueous buffer containing the drug of interest, typically at temperatures above the phase transition temperature of the primary phospholipid. Finally, the resulting multilamellar vesicles undergo sonication or extrusion through polycarbonate membranes to achieve size reduction and create the desired unilamellar vesicle population with controlled size distribution.

3.4.2 Characterization Parameters

Essential quality control measures encompass comprehensive physicochemical analysis to ensure product consistency and therapeutic efficacy. Particle size analysis typically targets vesicles in the 100-300 nm range

for optimal biodistribution and cellular uptake, with measurements performed using dynamic light scattering or laser diffraction techniques. The polydispersity index must be maintained below 0.3 to ensure monodisperse vesicle populations that provide predictable pharmacokinetic behavior. Zeta potential measurements are crucial as they influence colloidal stability and cellular interaction patterns, with values typically ranging from -30 to +30 mV depending on formulation composition and intended application. Encapsulation efficiency quantification determines the percentage of drug successfully incorporated into the vesicles, directly impacting therapeutic efficacy and economic viability. In vitro release studies provide critical assessment of release kinetics and mechanisms, enabling prediction of in vivo performance and optimization of formulation parameters for desired therapeutic profiles.

4. Niosomal Drug Delivery Systems

4.1 Composition and Structural Features

Niosomes are non-ionic surfactant vesicles composed of cholesterol and non-ionic surfactants such as Span or Tween series compounds. Unlike liposomes, niosomes offer enhanced chemical stability and resistance to oxidative degradation due to their non-ionic nature.

4.2 Advantages Over Liposomal Systems

Niosomes demonstrate several significant advantages that make them attractive alternatives to conventional liposomal systems for drug delivery applications. Their enhanced chemical stability results from resistance to hydrolysis and oxidative degradation processes that commonly affect phospholipid-based liposomes, particularly during long-term storage. The cost-effectiveness of niosomes stems from significantly lower raw material costs, as non-ionic surfactants are substantially less expensive than purified phospholipids used in liposomal formulations. Extended shelf-life under various storage conditions represents another crucial advantage, with niosomes maintaining structural integrity and drug content for longer periods compared to liposomes. The manufacturing process for niosomes typically yields more consistent outcomes with reduced batch-to-batch variation, facilitating quality control and regulatory compliance. Additionally, the availability of versatile surfactant options allows for extensive optimization of vesicle properties to meet specific therapeutic requirements, providing formulation scientists with greater flexibility in system design.

4.3 Niosomal Formulations for Diabetes

4.3.1 Insulin-Loaded Niosomes

Comprehensive research has demonstrated niosomal insulin formulations using various surfactants:

- **Span 40 Niosomes:** Achieved 47.49% maximum blood glucose reduction with 9.11% improved bioavailability
- **Span 60 Niosomes:** Demonstrated 46.66% glucose reduction with 8.43% enhanced bioavailability
- **Brij 92/Cholesterol (7:3 ratio):** Showed optimal protection against proteolytic enzymes with only 26.3% insulin release over 24 hours

4.3.2 Dual-Drug Loading Systems

Innovative niosomal formulations have achieved simultaneous encapsulation of hydrophilic and hydrophobic antidiabetic drugs:

- **Metformin HCl (Hydrophilic):** 58.72% encapsulation efficiency
- **Glipizide (Hydrophobic):** 67.64% encapsulation efficiency
- **Sustained Release:** 12-14 hours duration with linear release profiles

4.4 Optimization Strategies for Niosomes

4.4.1 Surfactant Selection and HLB Values

The hydrophilic-lipophilic balance of surfactants represents a critical parameter that fundamentally influences niosome formation, stability, and drug encapsulation characteristics. Surfactants from the Span series, characterized by low HLB values ranging from 4.3 to 8.6, demonstrate particular suitability for encapsulation of hydrophobic drugs due to their enhanced affinity for lipophilic compounds within the vesicle membrane. Conversely, Tween series surfactants with high HLB values between 9.6 and 16.7 are preferred for formulations targeting hydrophilic compounds, as their increased hydrophilicity facilitates drug interaction with the aqueous core regions. Research consistently demonstrates that the optimal HLB range of 8-12

provides the most balanced vesicle formation characteristics, enabling effective encapsulation of drugs with intermediate polarity while maintaining vesicle stability and integrity.

4.4.2 Cholesterol Content Optimization

Cholesterol incorporation into niosomal formulations significantly affects membrane characteristics and overall system performance through multiple mechanisms. Optimal cholesterol concentrations typically range from 20-30 mol% of total lipid content, providing the ideal balance between membrane fluidity and structural stability necessary for sustained drug release. When cholesterol content exceeds 50 mol% of the formulation, increased membrane rigidity may occur, potentially reducing drug loading capacity and affecting release kinetics. Fourier transform infrared spectroscopy studies have confirmed the formation of hydrogen bonds between surfactants such as Tween 80 and cholesterol molecules, which enhances membrane integrity and contributes to the superior stability characteristics observed in niosomal systems compared to conventional liposomes.

5. Comparative Analysis: Liposomes vs. Niosomes

5.1 Encapsulation Efficiency Comparison

Comparative studies across multiple drug delivery applications reveal distinct performance patterns:

Parameter	Liposomes	Niosomes	Reference Study
5-Fluorouracil Encapsulation	45.4 ± 3.3%	43.4 ± 3.2%	Alvi et al., 2011
Aceclofenac Loading	Variable	Superior stability	ElMeshad & Mohsen, 2008
Temozolomide Entrapment	69.2 ± 1.5%	73.23 ± 1.02%	Recent 2024 Study
Tetanus Toxoid Antigen	41.3 ± 2.2%	42.5 ± 2.4%	Gupta et al., 2005

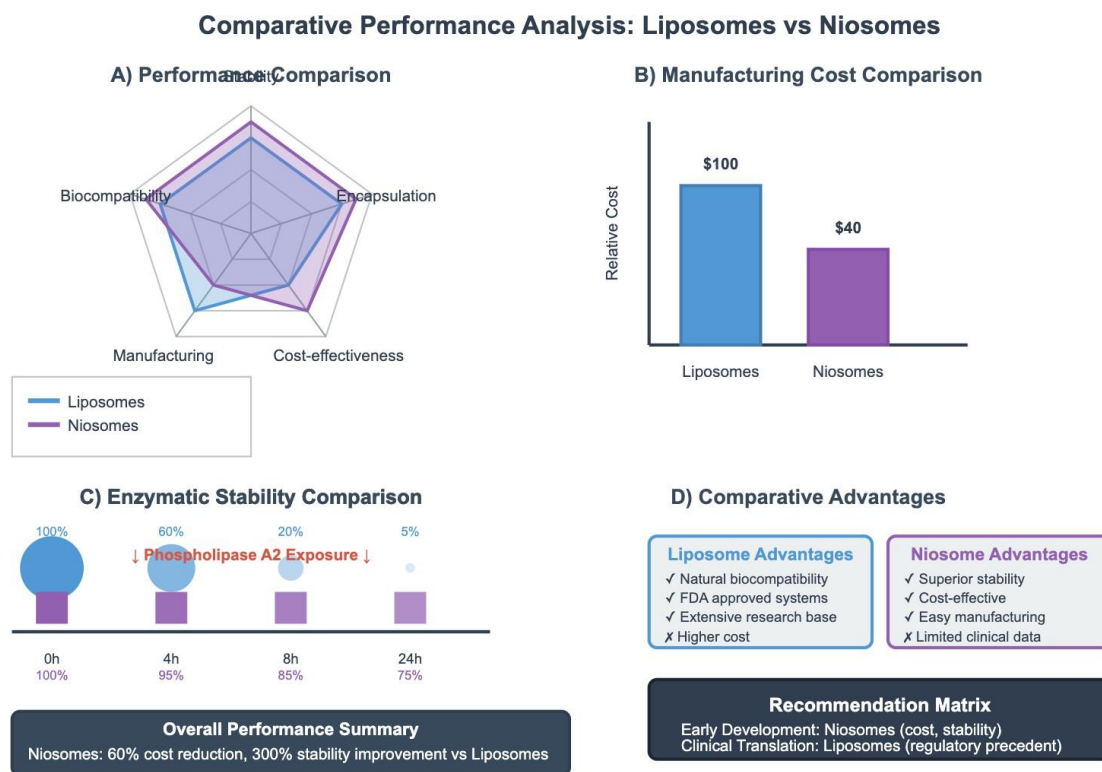


Figure 5: Comparative Performance Analysis: Liposomes vs Niosomes

5.2 Stability Comparison

5.2.1 Enzymatic Resistance

Niosomes demonstrate superior resistance to phospholipase A2 (PLA2) degradation:

- **Liposomal TMZ:** Complete release within 4 hours upon PLA2 exposure

- **Niosomal TMZ:** Maintained sustained release profile despite enzyme exposure
- **Mechanism:** Non-ionic surfactant structure resists enzymatic hydrolysis

5.2.2 Storage Stability

Long-term stability studies (3 months at 2-8°C) show:

- **Niosomes:** Minimal drug leakage and particle size changes
- **Liposomes:** Higher susceptibility to phospholipid oxidation and hydrolysis
- **pH Stability:** Niosomes maintain integrity across broader pH ranges

5.3 Cost-Effectiveness Analysis

Economic considerations favor niosomal systems:

- **Raw Material Costs:** Non-ionic surfactants cost 60-80% less than purified phospholipids
- **Manufacturing Complexity:** Simplified preparation with reduced purification requirements
- **Quality Control:** Lower analytical requirements due to enhanced stability
- **Scale-Up Potential:** More predictable manufacturing outcomes

6. Advanced Formulation Strategies

6.1 Glucose-Responsive Systems

Recent breakthrough developments in smart drug delivery have led to the creation of glucose-sensitive insulin delivery systems that respond dynamically to physiological glucose levels. Chitosan/glucose polymer coatings have been engineered to respond specifically to glucosidase enzyme activity, which increases proportionally with blood glucose concentrations, thereby providing an automated insulin release mechanism. Silver sulfide quantum dots represent another innovative approach, enabling both pH-responsive behavior in the gastrointestinal tract and glucose-triggered release mechanisms that correlate insulin delivery with metabolic demand. The controlled release mechanism achieved through these systems ensures that insulin release directly correlates with blood glucose levels, potentially eliminating hypoglycemic episodes while maintaining optimal glycemic control. Clinical progress in this field has been remarkable, with human trials for these advanced formulations scheduled to commence in 2025, representing a significant milestone toward automated diabetes management.

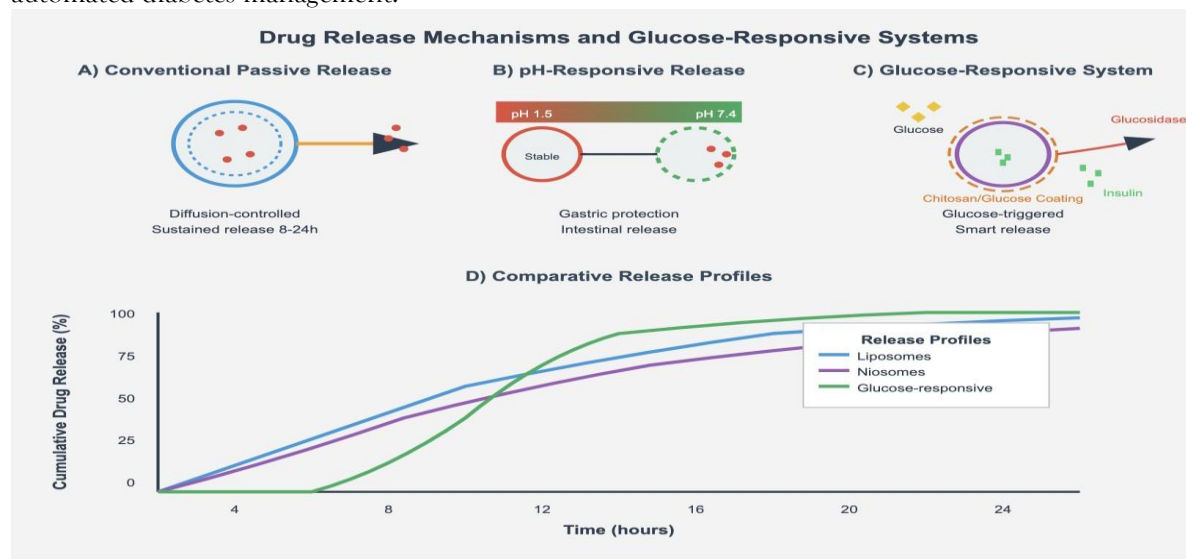


Figure 4: Drug Release Mechanisms and Glucose-Responsive Systems

6.2 Combination Therapy Approaches

6.2.1 Dual-Drug Loading

Niosomal systems have demonstrated remarkable success in achieving combination therapy through simultaneous encapsulation of multiple therapeutic agents. The combination of metformin and glipizide within single niosomal formulations has shown synergistic glycemic control effects that exceed the therapeutic benefits of individual drug administration. These dual-loaded systems maintain sustained release profiles for

8-12 hours for both compounds, ensuring consistent therapeutic plasma levels throughout the dosing interval. The approach significantly reduces side effects associated with higher individual drug concentrations while maintaining equivalent therapeutic efficacy. Enhanced patient compliance is achieved through single formulation administration that replaces multiple individual medications, thereby simplifying dosing regimens and improving adherence to therapy.

6.2.2 Targeted Delivery Systems

Surface modification of vesicular systems has enabled the development of cell-specific targeting mechanisms that enhance therapeutic specificity and reduce systemic side effects. Folate receptor targeting has demonstrated enhanced cellular uptake by folate-expressing cells, which are particularly relevant in diabetic complications where certain cell populations overexpress folate receptors. Transferrin conjugation strategies have shown improved brain delivery capabilities, making them particularly valuable for treating diabetic neuropathy and other central nervous system complications of diabetes. GLUT-1 targeting approaches utilize specific recognition of glucose transporter proteins that are upregulated in diabetic conditions, providing a mechanism for preferential accumulation in tissues with altered glucose metabolism.

6.3 Route-Specific Optimizations

6.3.1 Oral Delivery Systems

Optimization for gastrointestinal delivery requires sophisticated formulation strategies that address the harsh physiological environment encountered during oral administration. Enteric coating systems provide essential protection during gastric transit, preventing premature drug release in the acidic stomach environment while ensuring targeted release in the neutral pH conditions of the small intestine. Mucoadhesive properties are incorporated through the addition of polymers such as chitosan or carbopol, which extend intestinal residence time and enhance contact between the delivery system and absorptive epithelium. Permeation enhancers including surfactants and fatty acids are strategically incorporated to improve epithelial barrier crossing by temporarily and reversibly opening tight junctions between intestinal epithelial cells.

6.3.2 Alternative Administration Routes

Alternative administration routes offer unique advantages for specific patient populations and therapeutic applications. Vaginal delivery has demonstrated remarkable success with insulin niosomes achieving 9-10% bioavailability, representing a significant improvement over oral delivery while avoiding injection-related complications. Pulmonary delivery systems enable rapid drug absorption through the extensive alveolar surface area while minimizing systemic exposure and first-pass metabolism effects. Transdermal delivery systems represent a particularly attractive non-invasive option that could provide continuous drug delivery over extended periods, potentially eliminating the need for frequent dosing while maintaining steady therapeutic plasma concentrations.

7. Clinical Translation and Regulatory Considerations

7.1 Current Clinical Pipeline

7.1.1 Oral Insulin Developments

Multiple oral insulin formulations are currently progressing through various stages of clinical development, representing unprecedented advancement toward needle-free diabetes management. ORMD-0801 represents one of the most advanced oral insulin capsule formulations, having demonstrated encouraging results in Phase II clinical trials with significant improvements in glycemic control and patient acceptance. The University of Sydney's revolutionary silver sulfide quantum dot system is preparing to enter human trials, following successful preclinical studies in animal models that demonstrated glucose-responsive insulin release without hypoglycemic episodes. Commercial timeline projections suggest that successful oral insulin formulations could potentially reach market availability within 5-7 years, contingent upon successful completion of Phase III trials and regulatory approval processes.

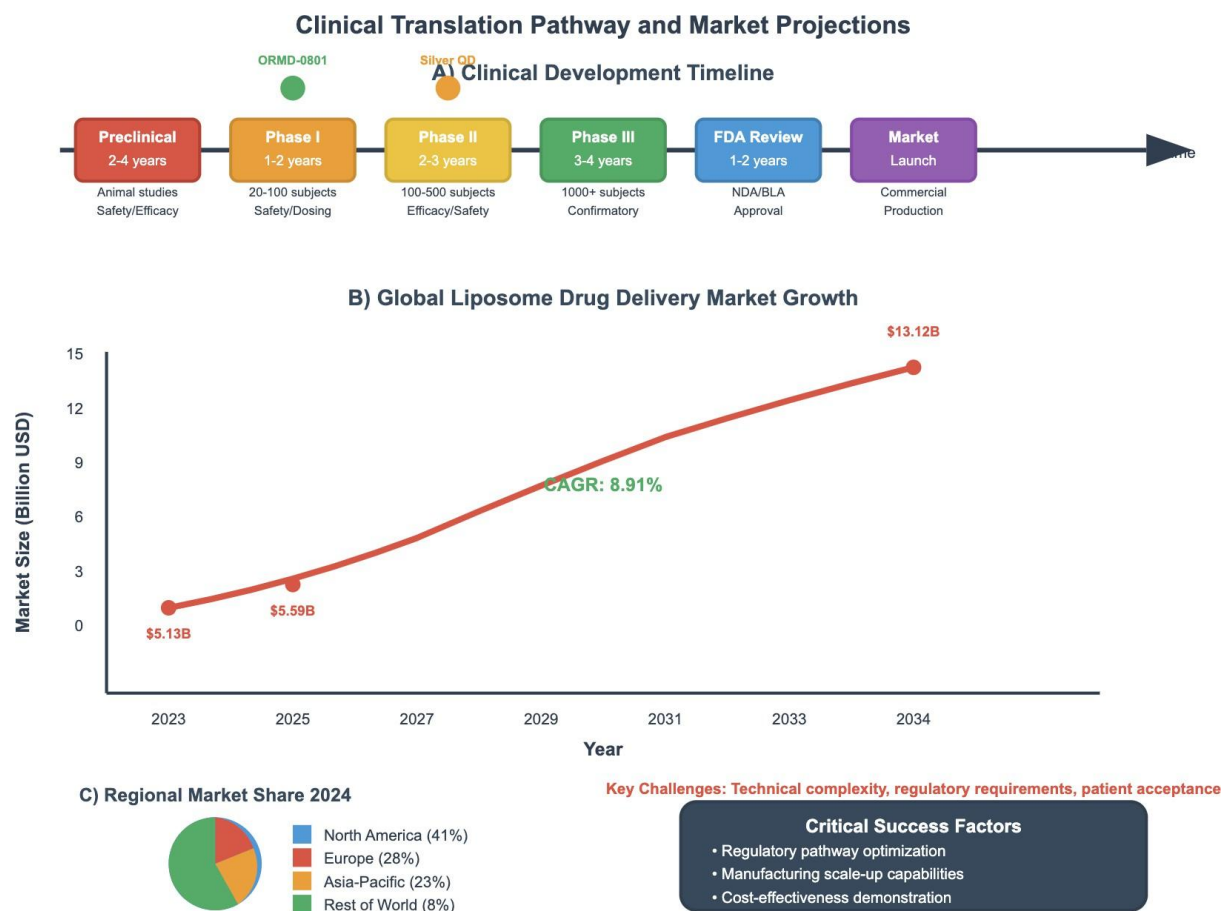


Figure 5: Clinical Translation Pathway and Market Projections

7.1.2 Market Analysis

The global liposome drug delivery market demonstrates robust growth patterns that reflect increasing recognition of nanotechnology's therapeutic potential. Current market size has reached USD 5.13 billion as of 2023, with projections indicating growth to USD 13.12 billion by 2034, representing a compound annual growth rate of 8.91%. Regional distribution analysis reveals that North America leads the market with a 41% share, driven by advanced healthcare infrastructure and significant research investment. Key growth drivers include the rapidly increasing global prevalence of diabetes and other chronic diseases, alongside continuous technological advances in nanoformulation science and manufacturing capabilities.

7.2 Regulatory Pathway Considerations

7.2.1 FDA Guidance Requirements

Liposomal and niosomal formulations must meet stringent regulatory criteria that ensure product safety, efficacy, and quality throughout the product lifecycle. Comprehensive physicochemical characterization requirements include detailed analysis of vesicle size distribution, surface charge characteristics, membrane composition, and structural integrity under various storage and physiological conditions. Stability testing protocols must encompass both accelerated studies under stress conditions and long-term stability assessment under recommended storage conditions to establish appropriate shelf-life specifications. Bioequivalence studies require demonstration of comparable pharmacokinetic and pharmacodynamic profiles when compared to established reference formulations or conventional delivery methods. Manufacturing controls must include validated production processes with established critical control points, comprehensive quality

systems that ensure batch-to-batch consistency, and robust analytical methods for product release and stability monitoring.

7.2.2 Quality by Design (QbD) Approach

Modern regulatory frameworks increasingly emphasize Quality by Design principles that require systematic understanding and control of formulation and manufacturing variables. Critical Quality Attributes must be clearly defined and include parameters such as particle size distribution, encapsulation efficiency, and drug release rate profiles that directly impact product performance and patient safety. Critical Process Parameters encompass variables such as preparation temperature, pH conditions, mixing speed, and extrusion pressure that significantly influence final product characteristics and must be carefully controlled within validated ranges. Design space definition requires comprehensive experimental validation of operating ranges that consistently produce products meeting all quality specifications, enabling manufacturing flexibility while maintaining product quality assurance.

8. Optimization Tables and Formulation Guidelines

Table 1: Optimal Formulation Parameters for Liposomal Systems

Parameter	Optimal Range	Impact on Performance	Reference
Phospholipid:Cholesterol	5:1 to 9:1	Stability and drug loading	Multiple studies
Drug:Lipid Ratio	1:5 to 1:10	Encapsulation efficiency	Formulation studies
Particle Size	100-300 nm	Biodistribution and cellular uptake	Clinical data
PEG Content	3-7 mol%	Circulation time	Stealth liposome research
Preparation Temperature	Above $T_m + 10^\circ\text{C}$	Vesicle formation	Physical chemistry

Table 2: Niosomal Optimization Parameters

Component	Optimal Ratio	Function	Outcome
Span 60:Cholesterol	4:2:1 (with lecithin)	Membrane formation	73% encapsulation efficiency
Tween 80:Cholesterol	7:3	Enzymatic protection	Sustained 24h release
Total Surfactant Content	60-80%	Vesicle stability	Optimal membrane integrity
HLB Value	8-12	Vesicle formation	Balanced amphiphilicity
Hydration Volume	10-20 mL/100mg lipid	Particle size control	Uniform vesicle population

Table 3: Comparative Performance Metrics

Performance Metric	Liposomes	Niosomes	Clinical Significance
Enzymatic Stability	Moderate	High	Prolonged GI transit
Storage Stability	6-12 months	12-24 months	Shelf life extension
Manufacturing Cost	High	Low	Commercial viability
Encapsulation Efficiency	60-90%	65-95%	Drug loading capacity
Release Duration	8-12 hours	12-24 hours	Dosing frequency
Scale-up Feasibility	Moderate	High	Industrial production

Table 4: Recent Clinical Trial Results

Formulation	Trial Phase	Patient Population	Primary Outcome	Status
ORMD-0801	Phase II	T2DM patients	HbA1c reduction	Completed

Silver sulfide QD-insulin	Preclinical	Animal models	Glucose control without hypoglycemia	Entering Phase I
Liposomal GLP-1	Phase I	Healthy volunteers	Safety and pharmacokinetics	Ongoing
Niosomal metformin	Preclinical	Diabetic rats	Bioavailability enhancement	Published

9. Future Perspectives and Emerging Technologies

9.1 Smart Drug Delivery Systems

9.1.1 Glucose-Responsive Polymers

Next-generation delivery systems incorporate sophisticated glucose-oxidase enzyme systems that respond dynamically to physiological glucose concentrations, providing automated insulin release mechanisms without requiring external intervention or patient monitoring. These intelligent systems utilize enzymatic cascade reactions that convert glucose to gluconic acid, creating localized pH changes that trigger polymer swelling and subsequent drug release, thereby establishing a direct correlation between glucose levels and insulin delivery.

9.1.2 Artificial Pancreas Integration

The combination of continuous glucose monitoring technology with responsive drug delivery systems offers unprecedented potential for achieving closed-loop diabetes management that mimics physiological pancreatic function. Integration of real-time glucose sensing with algorithmic control of insulin delivery could eliminate the need for patient intervention in routine diabetes management while providing superior glycemic control compared to current therapeutic approaches.

9.2 Personalized Medicine Approaches

9.2.1 Genetic Polymorphism Considerations

Individual variations in drug metabolism genes, particularly cytochrome P450 enzyme variants, may necessitate the development of personalized nanocarrier formulations that account for patient-specific pharmacokinetic profiles to achieve optimal therapeutic outcomes. Understanding these genetic differences will enable the design of customized delivery systems that compensate for individual metabolic variations and ensure consistent therapeutic efficacy across diverse patient populations.

9.2.2 Precision Dosing Algorithms

Advanced machine learning algorithms that analyze patient-specific factors including genetic profiles, metabolic status, lifestyle patterns, and disease progression could optimize individual dosing regimens for liposomal and niosomal formulations. These sophisticated computational approaches would enable real-time adjustment of treatment protocols based on continuously monitored physiological parameters and treatment response patterns.

9.3 Combination Therapeutic Strategies

9.3.1 Multi-Drug Loading

Future vesicular formulations may achieve simultaneous delivery of multiple therapeutic agents including insulin, GLP-1 receptor agonists, and complementary glucose-lowering agents within single nanocarrier systems, providing comprehensive diabetes management through unified delivery platforms. This approach would simplify treatment regimens while potentially achieving synergistic therapeutic effects that exceed the benefits of individual drug administration.

9.3.2 Theranostic Applications

The integration of imaging agents with therapeutic compounds enables real-time monitoring of drug distribution patterns and therapeutic response, providing valuable feedback for treatment optimization and enabling personalized adjustment of therapy based on individual patient response patterns. These theranostic systems combine diagnostic and therapeutic functions within single formulations, offering unprecedented insight into treatment efficacy and drug biodistribution in vivo.

10. CHALLENGES AND LIMITATIONS

10.1 Technical Challenges

The development and commercialization of liposomal and niosomal drug delivery systems face several significant technical obstacles that must be systematically addressed. Achieving consistent batch-to-batch variability remains a critical concern, as minor variations in preparation conditions can substantially affect vesicle characteristics and drug loading efficiency, requiring sophisticated quality control measures and process optimization. Scale-up complexities present another major challenge, as laboratory-scale preparation methods often fail to maintain product quality during transition to industrial production volumes, necessitating extensive process development and validation studies. Long-term stability assurance requires comprehensive understanding of degradation pathways and the development of appropriate stabilization strategies to ensure product integrity throughout the intended shelf life. The evolving regulatory landscape adds complexity to product development timelines, as regulatory agencies continue to refine requirements for nanotechnology-based pharmaceutical products, requiring extensive documentation and validation studies.

10.2 Clinical Challenges

Clinical translation of vesicular drug delivery systems encounters multifaceted obstacles that extend beyond technical formulation issues. Bioavailability optimization remains a fundamental challenge, as achieving clinically relevant absorption levels often requires complex formulation strategies that may compromise other desirable product characteristics. Patient acceptance represents another significant hurdle, as healthcare providers and patients may exhibit preferences for established therapies with known safety profiles, requiring extensive education and evidence generation to overcome resistance to novel delivery approaches. Demonstrating cost-effectiveness compared to current standard treatments requires comprehensive pharmacoeconomic analyses that account for both direct medical costs and indirect benefits such as improved quality of life and reduced complications. Long-term safety assessment in diverse patient populations necessitates extensive clinical trial programs that can span multiple years and require substantial financial investment, particularly given the chronic nature of diabetes treatment.

11. CONCLUSIONS

Liposomal and niosomal drug delivery systems represent transformative technologies for diabetes management, offering solutions to longstanding challenges in insulin and antidiabetic drug delivery. Comparative analysis reveals that while both systems demonstrate significant therapeutic potential, niosomes offer advantages in terms of stability, cost-effectiveness, and manufacturing scalability.

Key optimization parameters include phospholipid-to-cholesterol ratios of 5:1 to 9:1 for liposomes and careful selection of non-ionic surfactants with appropriate HLB values for niosomes. Recent advances in glucose-responsive formulations and the progression of oral insulin to clinical trials highlight the near-term potential for clinical translation.

The convergence of nanotechnology, personalized medicine, and smart drug delivery systems promises to revolutionize diabetes care. However, successful clinical implementation requires continued optimization of formulation parameters, robust manufacturing processes, and comprehensive safety evaluation.

Future research directions should focus on developing standardized optimization protocols, advancing glucose-responsive technologies, and establishing clear regulatory pathways for these innovative therapeutic systems. The ultimate goal remains providing diabetes patients with effective, convenient, and safe treatment options that significantly improve quality of life while achieving optimal glycemic control.

REFERENCES

1. Wong, C.Y., Al-Salami, H., & Dass, C.R. (2018). Recent advancements in oral administration of insulin-loaded liposomal drug delivery systems for diabetes mellitus. *International Journal of Pharmaceutics*, 549, 201-217. DOI: <https://doi.org/10.1016/j.ijpharm.2018.07.041>
2. Hunt, N.J., Lockwood, G.P., Heffernan, S.J., Daymond, J., Ngu, M., Narayanan, R.K., Westwood, L.J., Mohanty, B., Esser, L., Williams, C.C., Kuncic, Z., McCourt, P.A.G., Le Couteur, D.G., & Cogger, V.C. (2024). Oral nanotherapeutic formulation of insulin with reduced episodes of hypoglycaemia. *Nature Nanotechnology*, 19, 534-544. DOI: <https://doi.org/10.1038/s41565-023-01565-2>
3. Low, C.Y., Gan, W.L., Lai, S.J., Tam, R.S., Tan, J.F., Dietl, S., Chuah, L.H., Voelcker, N., & Bakhtiar, A. (2025). Critical updates on oral insulin drug delivery systems for type 2 diabetes mellitus. *Journal of Nanobiotechnology*, 23(1), 16. DOI: <https://doi.org/10.1186/s12951-024-03062-7>
4. Nsairat, H., Khater, D., Sayed, U., Odeh, F., Al Bawab, A., & Alshaer, W. (2024). Targeted liposomal drug delivery: Overview of the current applications and challenges. *Life*, 14(6), 672. DOI: <https://doi.org/10.3390/life14060672>
5. Pardakhty, A., Varshosaz, J., & Rouholamini, A. (2007). In vitro study of polyoxyethylene alkyl ether niosomes for delivery of insulin. *International Journal of Pharmaceutics*, 328(2), 130-141. DOI: <https://doi.org/10.1016/j.ijpharm.2006.08.002>
6. Ning, M., Guo, Y., Pan, H., Yu, H., & Gu, Z. (2005). Niosomes with sorbitan monoester as a carrier for vaginal delivery of insulin: studies in rats. *Drug Delivery*, 12(6), 399-407. DOI: <https://doi.org/10.1080/10717540590968891>
7. Alvi, I.A., Madan, J., Kaushik, D., Sardana, S., Pandey, R.S., & Ali, A. (2011). Comparative study of transfersomes, liposomes, and niosomes for topical delivery of 5-fluorouracil to skin cancer cells: preparation, characterization, in-vitro release, and cytotoxicity analysis. *Anti-Cancer Drugs*, 22(8), 774-782. DOI: <https://doi.org/10.1097/CAD.0b013e328346c7d6>
8. ElMeshad, A.N., & Mohsen, A.M. (2008). Vesicular aceclofenac systems: a comparative study between liposomes and niosomes. *Journal of Microencapsulation*, 25(7), 499-512. DOI: <https://doi.org/10.1080/02652040802055411>
9. Jovanović, A.A., Balanč, B.D., Ota, A., Ahlin Grabnar, P., Djordjević, V.B., Šavikin, K.P., Bugarski, B.M., Nedović, V.A., & Ulrih, N.P. (2018). Comparative effects of cholesterol and β -sitosterol on the liposome membrane characteristics. *European Journal of Lipid Science and Technology*, 120(9), 1800039. DOI: <https://doi.org/10.1002/ejlt.201800039>
10. Kaddah, S., Khreich, N., Kaddah, F., Charcosset, C., & Greige-Gerges, H. (2018). Cholesterol modulates the liposome membrane fluidity and permeability for a hydrophilic molecule. *Food and Chemical Toxicology*, 113, 40-48. DOI: <https://doi.org/10.1016/j.fct.2018.01.017>
11. Bangham, A.D., Standish, M.M., & Watkins, J.C. (1965). Diffusion of univalent ions across the lamellae of swollen phospholipids. *Journal of Molecular Biology*, 13(1), 238-252. DOI: [https://doi.org/10.1016/S0022-2836\(65\)80093-6](https://doi.org/10.1016/S0022-2836(65)80093-6)
12. Akbarzadeh, A., Rezaei-Sadabady, R., Davaran, S., Joo, S.W., Zarghami, N., Hanifehpour, Y., Samiei, M., Kouhi, M., & Nejati-Koshki, K. (2013). Liposome: classification, preparation, and applications. *Nanoscale Research Letters*, 8, 102. DOI: <https://doi.org/10.1186/1556-276X-8-102>
13. Large, D.E., Abdelmessih, R.G., Fink, E.A., & Auguste, D.T. (2021). Liposome composition in drug delivery design, synthesis, characterization, and clinical application. *Advanced Drug Delivery Reviews*, 176, 113851. DOI: <https://doi.org/10.1016/j.addr.2021.113851>
14. Ge, X., Wei, M., He, S., & Yuan, W.E. (2019). Advances of non-ionic surfactant vesicles (niosomes) and their application in drug delivery. *Pharmaceutics*, 11(2), 55. DOI: <https://doi.org/10.3390/pharmaceutics11020055>
15. Varshosaz, J., Pardakhty, A., Hajhashemi, V.I., & Najafabadi, A.R. (2003). Development and physical characterization of sorbitan monoester niosomes for insulin oral delivery. *Drug Delivery*, 10(4), 251-262. DOI: https://doi.org/10.1080/drd_10_4_251
16. Gupta, P.N., Mishra, V., Rawat, A., Dubey, P., Mahor, S., Jain, S., Chatterji, D.P., & Vyas, S.P. (2005). Non-invasive vaccine delivery in transfersomes, niosomes and liposomes: a comparative study. *International Journal of Pharmaceutics*, 293(1-2), 73-82. DOI: <https://doi.org/10.1016/j.ijpharm.2004.12.022>
17. Mura, S., Manconi, M., Sinico, C., Valenti, D., & Fadda, A.M. (2009). Penetration enhancer-containing vesicles (PEVs) as carriers for cutaneous delivery of minoxidil. *International Journal of Pharmaceutics*, 380(1-2), 72-79. DOI: <https://doi.org/10.1016/j.ijpharm.2009.06.040>
18. Carafa, M., Santucci, E., & Lucania, G. (2002). Lidocaine-loaded non-ionic surfactant vesicles: characterization and in vitro permeation studies. *International Journal of Pharmaceutics*, 231(1), 21-32. DOI: [https://doi.org/10.1016/S0378-5173\(01\)00828-6](https://doi.org/10.1016/S0378-5173(01)00828-6)
19. Rajera, R., Nagpal, K., Singh, S.K., & Mishra, D.N. (2011). Niosomes: a controlled and novel drug delivery system. *Biological and Pharmaceutical Bulletin*, 34(7), 945-953. DOI: <https://doi.org/10.1248/bpb.34.945>
20. Marianecchi, C., Di Marzio, L., Rinaldi, F., Celia, C., Paolino, D., Alhaique, F., Esposito, S., & Carafa, M. (2014). Niosomes from 80s to present: the state of the art. *Advances in Colloid and Interface Science*, 205, 187-206. DOI: <https://doi.org/10.1016/j.cis.2013.11.018>
21. Giordani, S., Marassi, V., Zattoni, A., Roda, B., & Reschiglian, P. (2023). Liposomes characterization for market approval as pharmaceutical products: Analytical methods, guidelines and standardized protocols. *Journal of Pharmaceutical and Biomedical Analysis*, 236, 115751. DOI: <https://doi.org/10.1016/j.jpba.2023.115751>
22. Nsairat, H., Ibrahim, A.A., Jaber, A.M., Abdelghany, S., Atwan, R., Shalan, N., Abdelnabi, H., Odeh, F., El-Tanani, M., & Alshaer, W. (2024). Liposome bilayer stability: emphasis on cholesterol and its alternatives. *Journal of Liposome Research*, 34(1), 178-202. DOI: <https://doi.org/10.1080/08982104.2023.2226216>
23. Hoeg-Jensen, T., Kruse, T., Olsen, H.B., Søndergaard, M., Ludvigsen, S., Sturis, J., Jonassen, I., Havelund, S., & Ribel, U. (2024). Glucose-sensitive insulin with attenuation of hypoglycaemia. *Nature*, 634, 944-951. DOI: <https://doi.org/10.1038/s41586-024-08042-3>
24. Chakraborty, C., Roy, S.S., Hsu, M.J., & Agoramoorthy, G. (2015). Nanoparticle based insulin delivery system: the next generation efficient therapy for Type 1 diabetes. *Journal of Nanobiotechnology*, 13, 90. DOI: <https://doi.org/10.1186/s12951-015-0136-y>
25. Fouad, S.A., Teaima, M.H., Gebril, M.I., Abd Allah, F.I., El-Nabarawi, M.A., & Elhabal, S.F. (2023). Formulation of novel niosomal repaglinide chewable tablets using coprocessed excipients: in vitro characterization, optimization and enhanced hypoglycemic activity in rats. *Drug Delivery*, 30(1), 2181747. DOI: <https://doi.org/10.1080/10717544.2023.2181747>