

THE THERAPEUTIC POTENTIAL OF LICHENS IN INDIA: A REVIEW

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ABSTRACT

Since ancient times, Lichens have been used as a great source for flavouring, dyeing, smoking tobacco, cigars, spices, scenting soaps, and cosmetics and manufacture of Dhoop and Hawan Samagri. Lichens in the field of medication for different disease and disorders are being used from centuries. Traditional knowledge is an important source for the development of new drugs. It has been observed that, strong metabolites from mycobiont culture of lichens and their extracts can be used in various ways to treat many human diseases. Keeping above point in view, an attempt has been made to review the therapeutic potential of lichens in India.

Key Words: Lichens, medicines, therapy, symbiotic.

INTRODUCTION

In nature, number of organisms play specific ecological role in their specialized functional niches and lichens are the excellent example of such associations with remarkable alliance between algal and fungal partners. These are those lichenized fungi that form symbiotic association with a photosynthetic alga or cyanobacteria through the process of lichenization. Therefore, fungal association within the lichen thallus is called Lichenized fungus. Theophrastus (370–285), the Father of Botany is credited to introduce the term lichen from the

Greek origin ‘Leikhén’. Tournfort in 1700 (A.D.) proposed lichen as one of the plant genera. Scholar of Linnaeus, Erik Acharius is known as Father of Lichenology, and described many genera of lichens.

Fungal partner in lichens is known as Mycobiont and Photosynthetic partner as Photobiont where main thallus body formed by mycobiota (about 95%) whereas photobiont contributes only 5% of the total thallus. In the thallus body fungal partner provides moisture, protection and shelter to the algal partner while algal partner provides carbon food through photosynthesis. The photobiont may be one or more green algal species, one or more cyanobacterial species or the combination of both. Within a thallus, the mycobiont’s hyphae exudes chemicals which increases (Grover and Seshadri, 1959) the permeability of the photobiont cell wall or cell membrane and allow increased transference of organic carbon from the photobiont to the mycobiont. Sometimes, the relation is called controlled or balanced parasitism because the fungus does not kill the alga but at the same time it does not allow alga to flourish.

DISTRIBUTION OF LICHENS

Lichens are widely distributed throughout the world, and occur almost in all climatic conditions where even little favorable conditions are available. They are found in most extreme environments, from the Arctic to Antarctic, deserts to tropics, littoral zones to mountain peaks, etc. Some live on desert floors or near hot springs where temperatures can soar to 148°C. Lichens are one of the first colonizers or pioneers and colonize almost any substrate such as rocks, soils, man-made surfaces (Metal, polythene, rubber, glass, bricks, etc.) and bark and leaves of a variety of plants. Lichens may absorb certain mineral nutrients from any of these substrates on which it grows, but is generally self-reliant in feeding itself through photosynthesis in the algal cells. Lichens growing on rocks, though, may release chemicals which speed the degradation of the rock resulting in weathering process, and thus promote production of new soil. About 85% of lichen-forming fungi associate with green algae. About 10% with cyanobacteria (Ahmadjian, 1993) and about 4% with both green and blue green algae. The most common lichen photobiont belongs to green algae *Trebouxia*, *Trentepohlia* and the cyanobacterium *Nostoc*. The bulk of the ground tissue of lichen is made up of the mycobiont. When moisture is completely absent, lichen tissues may undergo desiccation and this is not a simple dehydration as it occurs in other plants but the lichen thallus becomes quite brittle when the body water is completely lost (Tuovinen, 2017). Even the lichen can quickly absorb the water when the moisture is once again available and become soft and fleshy. Not only lichens

can undergo this drying, but while they are dry and brittle, pieces may flake off and later grow into new lichens. The Indian region with a total of 3029 million hectare is considered to be one of the 12 centres of origin and diversity of several plant species in the world. In terms of plant diversity India ranks 10th in the world and 4th in the Asia (Nayaka *et al.*, 2010).

Lichens can be classified mainly into five growth forms: *viz.*, 1. Crustose, 2. Foliose, 3. Fruticose 4. Squamulose and 5. Leprose 6. Placodioid. Crustose lichens are those that closely attached to the substratum like trees, side walls or soils and they cannot be removed without damaging the substrate *viz.*, *Xanthoria*. Loose, powdery lichen crusts without a layered structure called Leprose lichens *viz.*, *Chrysotrichia*. Foliose lichens are leaf-like structures, composed of lobes and can be detached to their substrates very easily *viz.*, *Lobaria*. Fruticose lichens are usually round in cross section and most are branched. They can like little shrubs growing upward, or they can hang down in long strands *viz.* *Usnea longissima*. In Squamulose Lichen the thallus is in the form of minute lobes, having dorsiventral differentiation like some species of *Cladonia*. In case of Placodioid Lichens the thallus is closely attached to the substratum at centre and lobate or free at the margin, but lacking rhizinae.

Based on their habitat lichens can be categorized into six types *viz.*, Lignicolous (on woods), Corticolous (on bark), Saxicolous (on rock), Marine (sores of sea), Freshwater (on siliceous rock), Terricolous (on Soil). Based on their Internal Structure: 1. Heteromerous lichens (Photobiont cell filaments are stratified in a layer) 2. Homoiomerous lichens (when two symbionts are uniformly distributed). Based on their fungal partner: Zahlbruckner (1907, 1926) divided the lichens into following 3 sub-classes on the basis of their fungal partner: 1. 1. Ascolichens (Fungus belongs to Ascomycetes), Basidiolichens (Fungus belongs to Basidiomycetes), and Hymenolichens (Fungus belongs to Deuteromycetes) where algal partner belongs to members of Chlorophyceae, Cynophyceae and rarely to Xanthophyceae and Phaeophyceae (Awasthi, 2000).

LICHENS HAVE VAST HISTORY IN MEDICINES

In India, lichens mainly used in Medicine, flavouring, dyeing, smoking tobacco, cigars, spices, scenting soaps, and cosmetics and manufacture of Dhoop and Hawan Samagri (Singh and Sinha, 1997). Written in the 15th century, the Doctrine of Signatures stated that a plant could be used to treat diseases that were most likely caused by nature. This led to the development of Phyto-therapeutics that are used in traditional medicines (TIM) or Ayurveda. Records of Medicinal plants in India also shows the word ‘Shipal’ (algae) in Rigveda, in which

a medicine named “Oushadhi” had been described (Nayaka *et al.*, 2010). Subsequently a number of Sanskrit synonyms of lichens, for example, ‘Shailaya’ and ‘Shilapushp’ have been described in Sushruta Samhita (1000 BC), Charaka Samhita (300-200 BC) and several Nighantu (1100-1800 AD). Chharila is widely known Ayurvedic medicine used in ancient system of Indian medicine, for different human disorders, e.g., headache, skin disorder, diarrhoea, leprosy and urinary trouble etc (Nayaka *et al.*, 2010).

Evernia furfuracea lichen has been found in an Egyptian vase since 18th Dynasty (1700-1600 BC) it was used in the form of medicine (Llano, 1948). The word lichen is derived from Greek word ‘Leprous’ and refers to lichens in treating skin diseases as they also has their peeling-skin appearance. Lichen like *Lobaria pulmonaria* (L.) Hoffm. and *Parmelia sulcata* Taylor (Parmeliaceae) have been used in the treatment of pulmonary and cranial diseases, respectively since the ages (Rizzini, 1952).

The source of Lichen chemical are basically their Metabolites produced by mycobiont by three pathways *viz.*, Shikimic Acid Pathway, Mevalonic Acid Pathway and Acetyl-Polymalonyl Pathway (Susana *et al.*, 2005). In this study Susana and co-workers have also provided the metabolites with their respective pathways. About 800 metabolites are produced by lichens and many are still unidentified to the mankind (Huneck and Yoshimura, 1996). On other hand Bousite and Grube have also published a paper on diversity of lichen metabolite, their pattern of evolution and genes involved in the production of metabolites.

LICHENS USED AS MEDICINES IN INDIA:

India represents 2450 lichen species (Singh and Sinha, 2010; Nayaka and Upreti, 2013) which, accounts 14% of the world lichen population. Out of this total number, 137 lichens have medicinal values. Most of all the lichen species (57) have antioxidant property, while a large number (55) has antibacterial capacity and some (37) has properties to treat cancer and cytotoxicity (Table 1).

Table 1. Important Lichen species used in different Human Aliments

Lichen Species	Metabolites/ Extract	Disease Name (Effective against)	References
Acarosporaceae			
<i>Acarospora gobiensis</i> H. Magn.	Acarogobien A, B	<i>Bacillus subtilis, Staphylococcus aureus</i>	Řezanka and Guschina 1999
Caliciaceae			

<i>Acrosocyphus sphaerophoroi</i> des Lév.	Methanol Extract	Bacteria- <i>Bacillus cereus</i> , <i>B. subtilis</i> , <i>Enterobacter aerogens</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Micrococcus luteus</i> , <i>Proteus mirabilis</i> , <i>Pseudomonas aeruginosa</i> , <i>Salmonella typhimurium</i> , <i>Staphylococcus aureus</i> , <i>Streptococcus pneumoniae</i> Fungi- <i>Aspergillus flavus</i> , <i>A. nidulans</i> , <i>A. niger</i> , <i>A. sulphuricus</i> , <i>A. terreus</i> , <i>Candida albicans</i> , <i>Cryptococcus albidus</i> , <i>Trichophyton rubrum</i> .	Singh <i>et al.</i> , unpublished data
Alectoriaceae			
<i>Alectoria ochroleuca</i> (Hoffm.) Massal.	Methanol Extract	Cancer chemopreventive and cytotoxic activity	Ingólfssdóttir <i>et al.</i> , 2000.
Arthoraphidaceae			
<i>Arthoraphis alpina</i> (Schaer.) R. Sant. in D. Hawksw., P. James and Coppins	Alcohol Extract	Cytotoxicity against slow growing BS-C-1 cells	Perry <i>et al.</i> , 1999
Arthoniaceae			
<i>Arthothelium awasthii</i> Patw. and Kulk.	Methanol Extract and Barbatic acid	Anti-oxidant activity includes lipid peroxidation and tyrosinase enzyme activity	Verma <i>et al.</i> , 2008a, 2008b
Parmeliaceae			
<i>Bulbothrix setschwanensis</i> (Zahlbr.) Hale	Acetone and Methanol Extract	Inhibition of tyrosinase and xanthine oxidase activity	Behera and Makhija 2002
<i>Cetraria aculeata</i> (Schreb.) Fr.	The acetone Extract and Protolichesterinic acid	<i>Escherichia coli</i> , <i>Staphylococcus aureus</i> , <i>Aeromonas hydrophila</i> , <i>Proteus vulgaris</i> , <i>Streptococcus faecalis</i> , <i>Bacillus cereus</i> , <i>Bacillus subtilis</i> , <i>Pseudomonas aeruginosa</i> , and <i>Listeria monocytogenes</i> , <i>Salmonella typhimurium</i> .	Zeytinoglu <i>et al.</i> , 2008
<i>Cetraria islandica</i> (L.) Ach.	Aqueous Extract, Methanol Extract, Protolichesterinic acid	Cancer chemopreventive and cytotoxic activity, antibacterial activity against <i>M. aurum</i>	Bachereau and Asta 1997
<i>Cetrelia braunsiana</i> (Muell. Arg.) W. Culb. and C. Culb.	Methanol Extract	<i>Bacillus cereus</i> , <i>B. subtilis</i> , <i>Enterobacter aerogens</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Micrococcus luteus</i> , <i>Proteus mirabilis</i> , <i>Pseudomonas aeruginosa</i> ,	Singh <i>et al.</i>

		<i>Staphylococcus aureus</i> , <i>Streptococcus pneumoniae</i> <i>Aspergillus flavus</i> , <i>A. nidulans</i> , <i>A. niger</i> , <i>A. sulphuricus</i> , <i>A. terricues</i> , <i>Candida albicans</i> , <i>Cryptococcus albidus</i> , <i>Trichophyton rubrum</i>	
Cladoniaceae			
<i>Cladia aggregata</i> (Swaerz.) Nyl.	Alcohol Extract, Methanol Extract	<i>Bacillus subtilis</i> , <i>Trichophyton mentagrophytes</i> , cytotoxicity against murine leukemia cells and slow growing BS-C-1 cells, <i>Bacillus subtilis</i> , <i>Propionibacterium acnes</i> and <i>Staphylococcus aureus</i>	Turner <i>et al.</i> , 1980
<i>Cladonia chlorophaea</i> (Flörke in Sommerf.) Spreng.	Boiled thallus	Used to wash sores, and immunity booster	Turner <i>et al.</i> , 1980
<i>Cladonia coccifera</i> (L.) Willd.	Boiled thallus	Immunity Booster	Vartia 1973
<i>Cladonia crispata</i> (Ach.) Flotow	Water-soluble polysaccharides	Antitumor activity	Nishikawa <i>et al.</i> , 1974
<i>Cladonia deformis</i> (L.) Hoffm.	Boiled Thallus Extract	Pulmonary tuberculosis and cough	Vartia 1973
<i>Cladonia fimbriata</i> (L.) Fr.	Alcohol Extract	Antimicrobial activity against <i>Bacillus subtilis</i> , <i>Candida albicans</i> , <i>Trichophyton mentagrophytes</i> and cytotoxicity against murine leukemia cells and slow growing BS-C-1 cells	Perry <i>et al.</i> , 1999
<i>Cladonia furcata</i> (Huds.) Schrad.	Methanol Extract	Exhibited cancer chemopreventive, cytotoxic activity	Ranković <i>et al.</i> , 2007
<i>Cladonia humilis</i> (With.) J. Laundon	Methanol Extract	Exhibited superoxide dismutase like activity	Yamamoto <i>et al.</i> , 1998
<i>Cladonia pyxidata</i> (L.) Hoffm.	Crushed Lichen	Symptoms that include hurried feeling, but less anxious and nervous; bloated abdomen, disorientation, uncertainty, dryness of tongue, lips, throat, skin and rectum; tired and yet sleeplessness, desire for open air; difficulty in breathing in hot room	Rogers, internet
<i>Cladonia rangiferina</i> (L.) Webber ex Wigg.	Dried thalli powder	Newborn baby bather, removing kidney stones	Vogel 1970

Coccocarpiaceae			
<i>Everniastrum cirrhatum</i> (Fr.) Hale	Alcohol Extract	Against murine leukemia cells and slow growing BS-C-1 cells	Perry <i>et al.</i> , 1999
Collemataceae			
<i>Collema flaccidum</i> (Ach.) Ach.	Bianthraquinone glycosides, Colleflaccinoids	Antitumor activity	Řezanka and Dembitsky 2006
Verrucariaceae			
<i>Dermatocarpus miniatum</i> (L.) Mann	Methanol Extract	Antimicrobial activity	Aslan <i>et al.</i> , 2006
Thelotremaeae			
<i>Diploschistes scruposus</i> (Schreber) Norman	Acetone Extract	<i>Bacillus cereus</i> , <i>B. megaterium</i> , <i>Staphylococcus aureus</i> and <i>Klebsiella pneumoniae</i>	Saenz <i>et al.</i> , 2006
Parmeliaceae			
<i>Evernia divaricatica</i> (L.) Ach.	Methanol Extract	Antimicrobial activity, immunity Booster	Aslan <i>et al.</i> , 2006
<i>Evernia prunastri</i> (L.) Ach.	Methanol Extract, Methanol Extract	Antibacterial (<i>Staphylococcus aureus</i> , <i>Bacillus mycoides</i> , <i>B. licheniformis</i> , <i>Mycobacterium phlei</i>) and antifungal (<i>Actinomyces sulfuroides</i> , <i>Trichophyton farineculatum</i> , <i>T. interdigitalis</i> , <i>Epidermophyton inguinale</i>)	Bustinza 1952, Aslan <i>et al.</i> , 2006
<i>Everniastrum cirrhatum</i> (Fr.) Hale	Chharila, Aqueous Extract, Ethanol Extract	Useful in dyspepsia, spermatorrhoea, amenorrhoea, calculi, diseases of blood and heart, stomach disorders, enlarged spleen, bronchitis, bleeding piles, scabies, leprosy, excessive salivation, soreness of throat, tooth-ache and general pain	Chandra and Singh 1971, Gupta <i>et al.</i> , 2007
<i>Everniastrum nepalense</i> (Taylor) Hale	Antiproliferative agents	against human keratinocyte line HaCaT and also inhibited the leukotriene B4 biosynthesis	Kumar and Muller 1999a, b
<i>Flavoparmelia caperata</i> (L.) Hale	Usnic acid	Antibacterial activity	Gupta <i>et al.</i> , 2007
Graphidaceae			

<i>Graphina acharii</i> (Fée) Müll. Arg., <i>G. adscribens</i> (Nyl.) Müll. Arg., <i>G. glaucorufa</i> (Vainio) Zahlbr., <i>G. glaucorufa</i> (Vainio) Zahlbr., <i>G. multistriata</i> Müll. Arg., <i>G. norlabiata</i> Patw. and Kulk., <i>G. adscribens</i> (Nyl.) Müll. Arg., <i>G. glaucorufa</i> (Vainio) Zahlbr., <i>G. multistriata</i> Müll. Arg., <i>Graphina norlabiata</i> Patw. and Kulk., <i>Graphina nylanderi</i> Patw. and Kulk., <i>Graphina perstriatula</i> (Nyl.) Zahlbr., <i>Graphina salacinilabiata</i> Patw. and Kulk., <i>Graphina simulans</i> (Leighton) Müll. Arg.,	Methanol Extracts	Inhibitory properties of tyrosinase and xanthine oxidase	Behera <i>et al.</i> , 2003, 2004, 2006
<i>Graphis assamensis</i> Nagar. and Patw.,	Similar to <i>Graphina acharii</i>	Inhibitory properties of tyrosinase and xanthine oxidase	Behera <i>et al.</i> , 2003, 2004, 2006

<i>G. exalbata</i> Nyl., <i>G. garoana</i> Nagar. and Patw., <i>G. glauconigra</i> Vainio, <i>G.</i> <i>guimaraana</i> Vainio, <i>G. hossei</i> Vainio, <i>G.</i> <i>inamoena</i> Zahlbr., <i>G.</i> <i>inquinata</i> (Knight and Miller) J.D. Hooker, <i>G.</i> <i>nakanishiana</i> Patw. and Kulk. , <i>G.</i> <i>patwardhanii</i> Kulk. , <i>G.</i> <i>persicina</i> Mey and Flotow, <i>G.</i> <i>persulcata</i> Stirton, <i>G.</i> <i>pyrrhocheiloid</i> <i>es</i> Zahlbr.,			
<i>Graphis</i> <i>scripta</i> (L.) Ach.	Methanol Extract	Inhibition of tyrosine activity, Epstein-Barr virus activation induced teleocidin B-4	Yamamoto <i>et al.</i> , 1998
<i>Graphis</i> <i>sikkimensis</i> Nagar. and Patw., <i>G.</i> <i>sorediosa</i> Nagar. and Patw.	Similar to <i>Graphina</i> <i>acharii</i>	Similar to <i>G. acharii</i>	Behera <i>et al.</i> , 2003, 2004
Physciaceae			
<i>Heterodermia</i> <i>diademata</i> (Taylor) D.D. Awasthi	Paste	On cuts and wound as plaster, Immunity Booster	Saklani and Upreti 1992
<i>Heterodermia</i> <i>microphylla</i> (Kurok.) Skorepa	Seconadry Metabolites	Antimicrobial activity	Santos <i>et al.</i> , 2004

<i>Heterodermia podocarpa</i> (Bél.) D.D. Awasthi	Methanol Extract	Exhibited anti-oxidant activity, Immunity booster	Verma <i>et al.</i> , 2008a, b
<i>Heterodermia leucomelos</i> (L.) Poelt	Ethanol Extract	Antibacterial activity against <i>Mycobacterium tuberculosis</i>	Gupta <i>et al.</i> , 2007
Parmeliaceae			
<i>Hypogymnia enteromorpha</i> (Ach.) Nyl.	Methanol Extract	Inhibition of tyrosine and superoxide	Yamamoto <i>et al.</i> , 1998
<i>Hypogymnia physodes</i> (L.) Nyl.	Methanol Extract	<i>Bacillus mycoides</i> , <i>B. subtilis</i> , <i>Staphylococcus aureus</i> , <i>S. albus</i> , <i>Sarcina lutea</i> , <i>Propionibacterium acnes</i> , <i>Enterobacter cloacae</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> and fungi <i>Aspergillus flavus</i> , <i>A. fumigatus</i> , <i>Botrytis cinerea</i> , <i>Candida albicans</i> , <i>Fusarium oxysporum</i> , <i>Mucor mucedo</i> , <i>Paecilomyces variotii</i> , <i>Penicillium purpureescens</i> , <i>P. verrucosum</i> , <i>Trichoderma harzianum</i>	Yamamoto <i>et al.</i> , 1998, Ranković <i>et al.</i> , 2007, 2008
<i>Hypogymnia tubulosa</i> (Schaer.) Havaas	3-hydroxyphosphodic acid	Antimicrobial activity against <i>Aeromonas hydrophila</i> , <i>Bacillus cereus</i> , <i>B. subtilis</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i>	Yilam <i>et al.</i> , 2005
Umbilicariaceae			
<i>Lasallia pustulata</i> (L.) Mérat	The acetone and methanol Extracts	<i>Aspergillus flavus</i> , <i>A. fumigatus</i> , <i>Botrytis cinerea</i> , <i>Candida albicans</i> , <i>Fusarium oxysporum</i> , <i>Mucor mucedo</i> , <i>Paecilomyces variotii</i> , <i>Penicillium purpureescens</i> , <i>P. verrucosum</i> , <i>Saccharomyces cerevisiae</i> , <i>Trichoderma harzianum</i>	Ranković <i>et al.</i> , 2007
Lecanoraceae			
<i>Lecanora muralis</i> (Schreb.) Rabenh. em Poelt	Usnic acid and Acetone Extract	Antibacterial activity against <i>Bacillus cereus</i> , <i>B. megaterium</i> , <i>Staphylococcus aureus</i>	Saenz <i>et al.</i> , 2006
Collemataceae			
<i>Leptogium cyanescens</i> (Rabenh.) Koerb.	Alcohol Extract	Cytotoxicity against murine leukemia cells	Perry <i>et al.</i> , 1999
Parmeliaceae			
<i>Lethariella cashmeriana</i> Krog	Sub-ingredient of	Antimicrobial activity	Wang <i>et al.</i> , 2001

	some treatments		
<i>Lobaria discolor</i> (Bory in Del.) Hue	Methanol Extract	Antibacterial and antifungal activity	Singh <i>et al.</i> , unpublished data
<i>Lobaria isidiosa</i> (Mull. Arg.) Vainio, <i>Lobaria japonica</i> (Zahlbr.) Asahina, <i>Lobaria pseudopulmonaria</i> Gyeln.	Polysaccharides	Antitumor agent	Takahashi 1974
<i>Lobaria quercizans</i> Michaux, <i>Lobaria retigera</i> (Bory) Trevisan	Stored after drying	Tonic effect on the body systems, Inc. Immunity	Smith 1923, Hu <i>et al.</i> , 1980
Mykoblastaceae			
<i>Mykoblastus sanguinarius</i> (L.) Norman	Methanol Extract	Exhibited superoxide dismutase like activity	Yamamoto <i>et al.</i> , 1998
Parmeliaceae			
<i>Myelochroa entotheiochroa</i> (Hue) Elix and Hale, <i>Myelochroa irregans</i> (Nyl.) Elix and Hale	Methanol Extract	Inhibition of Tyrosin activity	Yamamoto <i>et al.</i> , 1998
Nephromataceae			
<i>Nephroma expallidum</i> (Nyl.) Nyl.	Methanol Extract	Exhibited cancer chemopreventive and cytotoxic activity	Ingólfssdóttir <i>et al.</i> , 2000).
Parmeliaceae			
<i>Parmelia thomsonii</i> (Stirton) D.D. Awasthi	Smoke of the lichen	To relieve eye pain	Sinha and Singh 2005
<i>Parmelia saxatilis</i> (L.) Ach.	Organic Extract	Exhibited antibacterial against <i>Pseudomonas aeruginosa</i>	Ingólfssdóttir <i>et al.</i> , 1998
<i>Parmelia squarrosa</i> Hale	Methanol Extract	Antibacterial and antifungal activity	Singh <i>et al.</i> , unpublished data

<i>Parmelia sulcata</i> Taylor	Acetone, chloroform, diethyl ether, methanol and petroleum ether Extracts	<i>Aeromonas hydrophila, Bacillus cereus, B. mycoides B. subtilis, Enterobacter cloacae, Klebsiella pneumoniae, Listeria monocytogenes, Proteus vulgaris</i>	Candan <i>et al.</i> , 2007, Ranković <i>et al.</i> , 2007
<i>Parmotrema abessinicum</i> (Krempelh.) Hale	Indigenous medicine	Rathipuvvu	Watt and Breyer-Brandwijk 1962, Llano 1951
<i>Parmotrema austrosinense</i> (Zahlbr.) Hale	Alcohol Extract	Showed beta-glucosidase inhibitor activity	Lee and Kim 2000
<i>Parmotrema chinense</i> (Osbeck) Hale and Ahti	Water tonic	Used in diarrhoea, dyspepsia, spermatorrhoea, amenorrhoea, dysentery and headache	Nadkarni 1976, Abdulla <i>et al.</i> , 2007).
<i>Parmotrema dilatatum</i> (Vain.) Hale	Metabolites isolated	Antimicrobial activity	Santos <i>et al.</i> , 2004
<i>Parmotrema praesorediosum</i> (Nyl.) Hale	Hexane, dichloromethane, ethyl acetate, acetone and methanol Extracts	Beta-glucosidase inhibitor activity, <i>Bacillus cereus, Corynebacterium diphtheriae, Shigella flexnerii, Staphylococcus aureus, Vibrio cholerae</i> and <i>Candida albicans</i> .	Balaji and Hariharan 2007).
<i>Parmotrema reticulata</i> (Taylor) M. Choisy	Ethanol Extract	Antibacterial activity against, <i>Mycobacterium tuberculosis</i>	Gupta <i>et al.</i> , 2007
<i>Parmotrema saccatilobum</i> (Taylor) Hale	Methanol Extract	Inhibition of Epstein-Barr virus activation induced teleocidin B-4 and superoxide dismutase like activity	Yamamoto <i>et al.</i> , 1998
<i>Parmotrema sancti-angelii</i> (Lynge) Hale	Mixing the ash with mustard or linseed oil	To treat ring-worm	Brij Lal and Upreti 1995
<i>Parmotrema stuppeum</i> (Taylor) Hale	Methyl orsenillate, orsenillic acid, atranorin and lecanoric acid	Antioxidant activity and Immunity booster	Jayapraksha and Rao 2000
<i>Parmotrema tinctorum</i> (Nyl.) Hale	Methanol Extract, lecanoric acid	Exhibited lipid peroxidation and tyrosinase enzyme activity, anti-oxidant activity	Verma <i>et al.</i> , 2008a, b

Peltigeraceae			
<i>Peltigera canina</i> (L.) Willd.	Polysaccharides	Influence cells of the immune system	Omarsdottir <i>et al.</i> , 2005
<i>Peltigera dolichorhiza</i> (Nyl.) Nyl.	Alcohol Extract	Cytotoxicity against murine leukemia cells	Perry <i>et al.</i> , 1999
<i>Peltigera horizontalis</i> (Huds.) Raumg.	Water Extract	Antimicrobial activity and Immunity booster	Vartia 1973
<i>Peltigera leucophlebia</i> (Nyl.) Gyelin.	Tenuiorin and Methyl orsellinate	Inhibitory activity against 15-lipoxygenase from soybeans, 5-lipoxygenase from porcine leucocytes and proliferation of cultured human breast, pancreatic and colon cancer cell lines	Ingólfssdóttir <i>et al.</i> , 2002).
<i>Peltigera membranacea</i> (Ach.) Nyl. emend. Thomson	Alcohol Extract	Antimicrobial against <i>Bacillus subtilis</i> , <i>Trichophyton mentagrophytes</i> , cytotoxicity against murine leukemia cells and slow growing BS-C-1 cells	Perry <i>et al.</i> , 1999
<i>Peltigera polydactylon</i> (Neck.) Hoffm.	Crushed thallus	Cuts to stop bleeding and as an antiseptic,	Saklani and Upreti 1992
<i>Peltigera praetextata</i> (Flörke ex Sommerf.) Zopf	Acetone Extract	<i>Epidermatophyton floccosum</i> , <i>Microsporum audounii</i> , <i>M. canis</i> , <i>M. nanum</i> , <i>M. gypseum</i> , <i>Trichophyton mentagrophytes</i> , <i>T. rubrum</i> , <i>T. tonsurans</i> , <i>T. violaceum</i>	Shahi <i>et al.</i> , 2003
<i>Peltigera rufescens</i> (Weiss.) Humb., <i>Peltigera venosa</i> (L.) Hoffm.	Methanol Extract	Highest antioxidant activity	Odabasoglu <i>et al.</i> , 2005
Pertusariaceae			
<i>Pertusaria amara</i> (Ach.) Nyl.	Methanol Extract	Intermittent fever	Smith 1921
Graphidaceae			
<i>Phaeographina caesiopruinosa</i> (Fée) Müll. Arg., <i>P. noralboradian</i>	Similar to <i>Graphina acharii</i>	Similar to <i>Graphina acharii</i>	Behera <i>et al.</i> , 2003, 2004

<i>s</i> Patw. and Kulk., <i>P. indica</i> (Patw. and Nag.) Sipman and Aptroot, <i>P. angulosa</i> Muell. Arg. <i>P.</i> <i>submaculata</i> Zahlbr., <i>P. subritigrina</i> (Vainio) Zahlbr.			
Physciaceae			
<i>Physcia</i> <i>aipolia</i> (Ehrh. ex Humb.) Fürnr.	Atranorin	<i>Bacillus mycoides</i> , <i>B. subtilis</i> , <i>Staphylococcus aureus</i> , <i>Enterobacter</i> <i>cloacae</i> , <i>Escherichia coli</i> , <i>Klebsiella</i> <i>pneumoniae</i> and fungi <i>Aspergillus flavus</i> , <i>A. fumigatus</i> , <i>Botrytis cinerea</i> , <i>Candida</i> <i>albicans</i> , <i>Fusarium oxysporum</i>	Ranković <i>et al.</i> , 2008
Lobariaceae			
<i>Pseudocyphellaria aurata</i> (Ach.) Vainio	Water Extract	Antimicrobial activity	Sharnoff, internet
Parmeliaceae			
<i>Punctelia</i> <i>rudecta</i> (Ach.) Krog.	Extract	Exhibited antibacterial activity	Bustinza 1952
Ramalinaceae			
<i>Ramalina</i> <i>celastri</i> (Sprengel) Krog and Swinsc.	α-D-glucan polysaccharide	Activity against Sarcoma-180 tumor cells	Fazio <i>et al.</i> , 2007
<i>Ramalina</i> <i>conduplicans</i> Vainio	Methanol Extract	<i>Bacillus cereus</i> , <i>B. subtilis</i> , <i>Enterobacter</i> <i>aerogens</i> , <i>Escherichia coli</i> , <i>Klebsiella</i> <i>pneumoniae</i> , <i>Micrococcus luteus</i>	Singh <i>et al.</i> , unpublished data
<i>Ramalina</i> <i>farinacea</i> (L.) Ach.	Methanol and water Extract	Reduce HIV-I vector	Esimone <i>et al.</i> , 2005
<i>Ramalina</i> <i>pacifica</i> Asahina	Methanol Extract	Exhibited inhibition of tyrosine activity	Yamamoto <i>et al.</i> , 1998
<i>Ramalina</i> <i>pollinaria</i> (Westr.) Ach.	Methanol Extract	<i>Escherichia coli</i> , <i>Enterococcus faecalis</i> , <i>Proteus mirabilis</i> , <i>Staphylococcus aureus</i> , <i>Bacillus subtilis</i> and <i>B. megaterium</i>	Cansaran <i>et al.</i> , 2007
<i>Ramalina</i> <i>sinensis</i> Jatta	Methanol Extract	High antibacterial and antifungal activity	Singh <i>et al.</i> , unpublished data

<i>Ramalina subfarinacea</i> (Nyl.) Nyl.	Usnic acid and Acetone Extract	<i>Bacillus cereus</i> , <i>B. megaterium</i> , <i>Staphylococcus aureus</i> and <i>Klebsiella pneumoniae</i>	Saenz <i>et al.</i> , 2006
Roccellaceae			
<i>Roccella montagnei</i> Bel. em. D.D. Awasthi	Ethyl acetate, acetone, and methanol Extracts	<i>Staphylococcus aureus</i> , <i>Salmonella typhi</i> , <i>S. para typhi-B</i> , <i>Proteus vulgaris</i> , <i>Klebsiella pneumoniae</i> , <i>Candida albicans</i>	Balaji <i>et al.</i> , 2006
Peltigeraceae			
<i>Solorina crocea</i> (L.) Ach.	Methanol Extract	Exhibited cancer chemopreventive and cytotoxic activity	Ingólfssdóttir <i>et al.</i> 2000
Stereocaulaceae			
<i>Stereocaulon alpinum</i> Laurer	Lobaric acids and atranorin	<i>Mycobacterium aurum</i> , a nonpathogenic organism with sensitivity similar to the <i>Tuberculinum bacterium</i>	Ingólfssdóttir <i>et al.</i> 1998).
<i>Stereocaulon foliolosum</i> var. <i>foliolosum</i> Nyl.	Ethanol Extract	Antibacterial activity against virulent strain of <i>Mycobacterium tuberculosis</i> H37Rv	Gupta <i>et al.</i> , 2007
<i>Stereocaulon foliolosum</i> var. <i>strictum</i> (C. Bab.) Lamb.	Chew the raw thalli	Cure kidney stones	Sinha and Singh 2005).
<i>Stereocaulon himalayense</i> D.D. Awasthi <i>et al</i> Lamb.	Water Extract	For urinary, trouble and blisters of tongue.	Saklani and Upreti 1992
Lobariaceae			
<i>Sticta nylanderiana</i> Zahlbr.	Crushed thalli	Excessive cough	Sinha and Singh 2005
Alectoriaceae			
<i>Sulcaria sulcata</i> (Lev.) Bystrek ex Brodo and D. Hawks.	Methanol Extract	Inhibition of Epstein-Barr virus activation induced teleocidin B-4 and superoxide dismutase like activity	Yamamoto <i>et al.</i> , 1998).
Imperfect lichen			
<i>Thamnolia vermicularis</i> var. <i>subuliformis</i> (Ehrh.) W. Culb.	Heteroglycans and a beta-glucan	immunomodulating activity	Omarsdottir <i>et al.</i> , 2007
<i>Thamnolia vermicularis</i>	Methanol Extract	Teleocidin B-4 and superoxide dismutase like activity	Bustinza (1952)

var. <i>vermicularis</i> (Swartz) Schaer			
Umbilicariaceae			
<i>Umbilicaria cylindrica</i> (L.) Delise ex Duby	Alcohol Extract	Cytotoxicity against slow growing BS-C-1 cells	Perry <i>et al.</i> , 1999
Parmeliaceae			
<i>Usnea bismolliuscula</i> Zahlbr.	Methanol Extract	<i>Bacillus subtilis</i> , <i>Propionibacterium acnes</i> and <i>Staphylococcus aureus</i>	Yamamoto <i>et al.</i> , 1998
<i>Usnea ghattensis</i> G. Awasthi	Methanol Extract	activity against <i>Bacillus licheniformis</i> , <i>B. megaterium</i> , <i>B. subtilis</i> , <i>Staphylococcus aureus</i>	Behera <i>et al.</i> , 2005a, 2005b, 2006, 2009; Verma <i>et al.</i> , 2008
<i>Usnea longissima</i> Ach.	Diffractaic acid and usnic acid	showed antioxidant activity	Bayir <i>et al.</i> , 2006, Odabasoglu <i>et al.</i> , 2004, 2006
<i>Usnea montifiji</i> Mot.	Methanol Extract	<i>Bacillus subtilis</i> , <i>Propionibacterium acnes</i> and <i>Staphylococcus aureus</i>	Yamamoto <i>et al.</i> , 1998
<i>Usnea sikkimensis</i> Biswas	Methanol Extract	remedy for lung troubles, haemorrhage and asthma, while powdered lichen used to strengthen hair	Biswas 1956
<i>Usnea subflorida</i> (Zahlbr.) Mot.	Usnic acid Extracted with acetone	<i>Escherichia coli</i> , <i>Enterococcus faecalis</i> , <i>Proteus mirabilis</i> , <i>Staphylococcus aureus</i> , <i>Bacillus subtilis</i> and <i>B. megaterium</i>	Cansaran <i>et al.</i> , 2006
<i>Xanthoparmelia conspersa</i> (Ach.) Hale	Water Extract	Exhibited antibacterial activity against <i>A. faecalis</i>	Bustinza 1952).
<i>Xanthoparmelia coreana</i> (Gyeln.) Kurok.	Methanol Extract	<i>Bacillus subtilis</i> , <i>Propionibacterium acnes</i> and <i>Staphylococcus aureus</i>	Yamamoto <i>et al.</i> , 1998
<i>Xanthoparmelia pulla</i> (Ach.) O. Blanco, A. Crespo, Elix & D. Hawksw.	Methanol Extract	Against colloidal bismuth subcitrate genotoxicity	Geyikoglu <i>et al.</i> , 2007
Teloschistaceae			
<i>Xanthoria elegans</i> (Links) Th. Fr.	Methanol Extract	Exhibited cancer chemopreventive and cytotoxic activity	Ingólfssdóttir <i>et al.</i> , 2000

<i>Xanthoria fallax</i> (Hepp) Arnold	Methanol Extract	Epstein-Barr virus activation induced teleocidin B-4 and superoxide dismutase like activity	Yamamoto <i>et al.</i> , 1998
<i>Xanthoria parietina</i> (L.) Th. Fr.	Methanol Extract	Exhibited cancer chemo-preventive and cytotoxic activity	Ingólfssdóttir <i>et al.</i> , 2000

CONCLUSION AND FUTURE PERSPECTIVES

It is evident from the study that lichens offer a great source of remedies that can cure almost all the ailments of mankind. In spite of the variety of problems and challenges encountered in the drug discovery from various lichens, these lichens still play a significant role in isolation and prospects of finding new compounds from their culture extracts in the new era of medicines.

Further, researchers, government and communities all need to be aware of and care about the conservation and development activities of the lichens for future references.

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