

Molecular Detection Of Klebsiella Pneumonia Isolated From Ruminants Secretion In AL Shatrah City, South Iraq

¹Arjewan Mohsen Agmi , ²Hayder Kh.Shanan , ³Rabab Jabbar Sekhi

¹College of Veterinary Medicine, Al-Shatrah University, Email: orjuwan@vet.shu.edu.iq

²College of Veterinary Medicine, Al-Shatrah University, Email: haydaralmalikyt@shu.edu.iq ³College of Veterinary Medicine, Al-Shatrah University, Email: rababsukhi@shu.edu.iq

Abstract

In this current study showed that detection of Klebsiella species from variation infected cattle discharge. 120 samples and swabs were collected divided in to (40 from nasal secretion, 40 from vaginal discharge and finally 40 from a milk samples). These sample cultured on different culture media and identified by using cultural characteristic and microscopic appearance. klebsiella pneumonia Identified using ViteK II system and confirm the last diagnosed of those organism by using polymerase chain reaction(PCR)assay (16S rRNA). Polymerase chain reaction used to detected some gene K1,K2(which is one of several genes responsible for capsule formation), and also blaSHv gene which is one of the genes related with multy drug resistance. Results explained that Klebsiella pneumoniae were isolated in rate (10%) from nasal discharge, while isolated in rate (10%) from milk samples, on the other hand, vaginal secretion not appeared positive result for Klebsiella pneumonia. the result involved that K1,K2 genes were not detected in all samples of K. pneumonia. Also, completely isolates of K. pneumoniae harbored blaSHv gene (100%), Also the result included that Klebsiella pneumoniae can be consider as major pathogen responsible for different medical case affected bovine in Al-Shatrah district in Thi-Qar province.

Key words: *Klebsiella pneumoniae , vaginal secretion ,nasal discharge.*

INTRODUCTION

Klebsiella species are opportunistic infections that colonize mucosal surfaces without producing disease, yet they have the potential to move from the mucosa to other tissues and endanger life. (Paczosa and Mecsas, 2016). Gram-negative, bacilliform, non-sporulation, non-ciliated bacteria with a thick cell wall, Klebsiella species are the cause of the mucous appearance of colony in a solid-state cultures grown in vitro as well as the invasion of bacteria in vivo. Klebsiella pneumonia is thought to be the most clinically significant of the several Klebsiella species and subspecies in animals as well as humans. Klebsiella species can be found in soil, wastewater, plants, surface water, and other habitats. Klebsiella pneumonia is pathogen capable of causing a different infections. Classically, Klebsiella pneumonia is known the main causes of pneumonia, Urinary tract infections and bacteremia (Choby, et al.,2020). Bacterial pneumonia causes very economic loss to the beef industry and that increase in stress and viral infection, taking antibiotics to at-risk calves is often to prevent the disease. The mass of administration of antibiotic lead to negative result in meat production and that perhaps lead to increase of resistant pathogen (Vulikh et al., 2019).

The most pathogenic bacteria which affected bovine is Gram-negative bacteria which is an significant cause of bovine mastitis all over the world (Schukken, et al., 2013).The Sub clinical mastitis in cattle is a main and silent problem causes greater economic losses to the farmers. One of the primary causes of decreased milk quality and output is mastitis, which is defined as inflammation of the mammary glands' parenchyma and is characterized by pathologic alterations to glandular tissues as well as physical, chemical, and typically bacteriological abnormalities in milk (Kumari et al.,2018).

MATERIALS AND METHODS

Sample collection:

One hundred and twenty swabs and samples was collecting from the cattle, from Thi-Qar province (Shatra dispesary, farms), duing period From the November of the 2024 to January the of 2025, the information including age and location were fixed. milk samples putted in sterile container then in ice box and swabs (nasal and vaginal) putted in transport media then in ice box then the samples and swabs transport to microbiology laboratory in veterinary collage with period less than 24 hours .

Maintenance of bacterial isolates

The following is how the bacterial isolates were maintained:

Short term storage

For several months, the pure isolates of bacteria were kept in screw-capped universal tubes filled with brain-heart agar and cultured for twenty-four hours at 37°C. The slants were then securely wrapped with Parafilm and stored for three months at 4°C (Brown & smith, 2014).

Long term storage

The next day, a loop of pure bacterial culture was used to inoculate a brain heart infusion broth, and after 18 hours of incubation at 37°C , The broth was prepared for incubation by adding glycerol at an ultimate concentration of 20% and keeping it at -20°C for two to eight months (Goldman & Green,2015).

Isolation and identification of bacteria

The samples and swabs were collected from cattle from different places from AL Shatrra province .Then, the samples and swab transport and incubation in brain heart infusion on incubator in 37°C for 24 hours then growth on MacConky agar and put on incubation in 37°C for (24 to 48) hours in aerobic environment. Gram stain method done according to (leboffe and Pierce, 2021). Identification by using automated methods [Vitek II] system Gram-negative (GN) and Gram-positive (GP) identity cards were produced (Maina & Kagotho, 2014). Molecular studies Extraction of bacterial DNA All Klebsiella pneumoniae isolates had their genomic DNA extracted using the Presto™ Mini gDNA Microbial Kit in accordance with the manufacturer's instructions.

PCR detection of genes

The genes have been amplified using the primers specified in Table (1). A total of 12.5 µl of Mastering Mix, 1 µl of the specific reverse and forward primers for each gene, 3 µl of the DNA template as well as and water that lacks nuclease complete the last reaction in a tube volume of 20 µl.

Primers

Table (3-5) contains a list of all primers used in the present study.

Primer sequences utilized in gene amplification are shown in Table (3-5).

Gene	Primer Sequences (5´-3´)	Product size	Reference

A:	16SrRNA	F*: AGAGTTTGATCCTGGCTCAG R*: GGTTACCTTGTTACGACTT	1500bp
	K1	F: 5'-AGATAGAGGTGTATTGTCGC R: GAGCTCTATATGTTGGATGC	352bp
	K2		
	blaSHV	F: TCATACTTGACAGAGGGAGTAG R: ACGATCGTTACAGTGACAAG	321 bp
		F: GGCCGCGTAGGCATGATAGA R: CCCGGCGATTTGCTGATTTC	714bp

Adanin, T: Thymin, C: Cytocin, G: Guanin

Five microliters of DNA were combined with dye for loading and put into wells to electrophorese the separated DNA samples. After that, the wells were exposed to an electric field for 45–60 minutes at 70V. Tables listed the thermocycling programs for the K1, K2, blaSHV, and 16SrRNA genes, respectively.

Statistical Analysis

Descriptive statistics were used for the statistical analysis, and percentages (%) were used to represent the nominal variables (source of isolation). Five age groups were created from the age variable, which was regarded as an ordinal variable. To investigate all of the relationships and quality of fit within the variables, the chi-square along with Fisher's accurate tests were employed. The correlation between numerical and interval variables was assessed using the Eta test. Values below 0.05 were deemed statistically significant, taking into account the Alpha significance threshold.

RESULTS AND DISCUSSION

Regarding the distribution of mastitis cases among the age groups, this study found that mastitis was significantly associated ($P < 0.05$) with the smaller age groups (1-2 years) table (1). An excellent statistical correlation (Eta value =1) was also found between mastitis and age, Figure(1). Mastitis was recorded at a higher percentage (47.5%) in the age group (1-2 years), followed by the age group 3 years with a percentage of 20%, and the age group 4 years was recorded at 15%. In comparison, the age groups 5 and 6 years were recorded at 12.5% and 5%, respectively.

Table (1): Isolation rate of different cow samples

Age	Positive NO.	Percent	P-value (χ^2)
1-2 years	19	47,5%%	000 (21.250)
3 years	8	20%	
4 years	6	15%	
5 years	5	12,5%	
6 years	2	5%	
Total	40	100%	

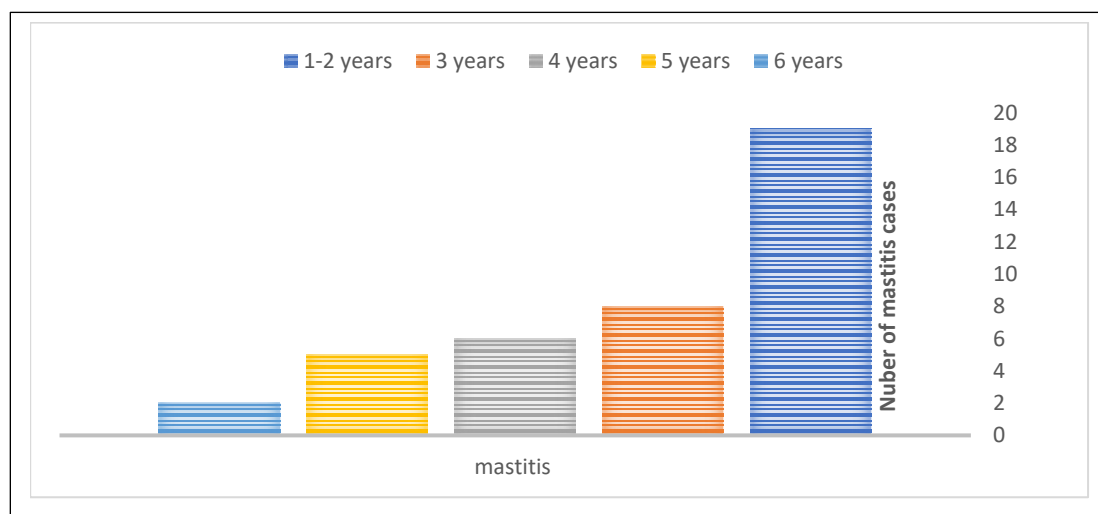


Figure (1): the correlation between the age groups and the prevalence of mastitis (Eta value=1) indicative of excellent correlation

Colony morphology

After 24-48 hours of incubation on agar from MacConkey at 37 °C, the first bacteria from the huge colony were identified, pink, regular edged, spherical colonies with a mucoid texture were produced by *Klebsiella* on MacConkey agar when the bacteria fermented the lactose.



Figure (2) *Klebsiella pneumoniae* on MacConkey agar

Table (2) represents the results of the distribution of vaginal infection among the age groups. The mean age was 1.95 ± 1.2 years, ranging from >1 year to <4 years. This study found significant differences ($P < 0.05$) in the

distribution of vaginal infection regarding the age groups. The highest infection rate was found in the age group >1 year (50%), followed by 25% in the age group 1-2 years old. However, the vaginal infection in the age groups >2-3 and >3-4 was equal (10%) for each. The lowest infection rate was found in the age group >4 years. Similarly, an excellent correlation was found between the age groups and the vaginal infection (Eta value = 98%), figure (3)

Table (2): the distribution of vaginal infection among different cow age groups

Age	Positive No.	Percent	P-value (X ²)
<1	20	50%	000 (27.00)
1-2s	10	25%	
>2-3 years	4	10%	
>3-4 years	4	10%	
>4	2	5%	
Total	40	100%	

X² = Chi-square test, degree of freedom=4

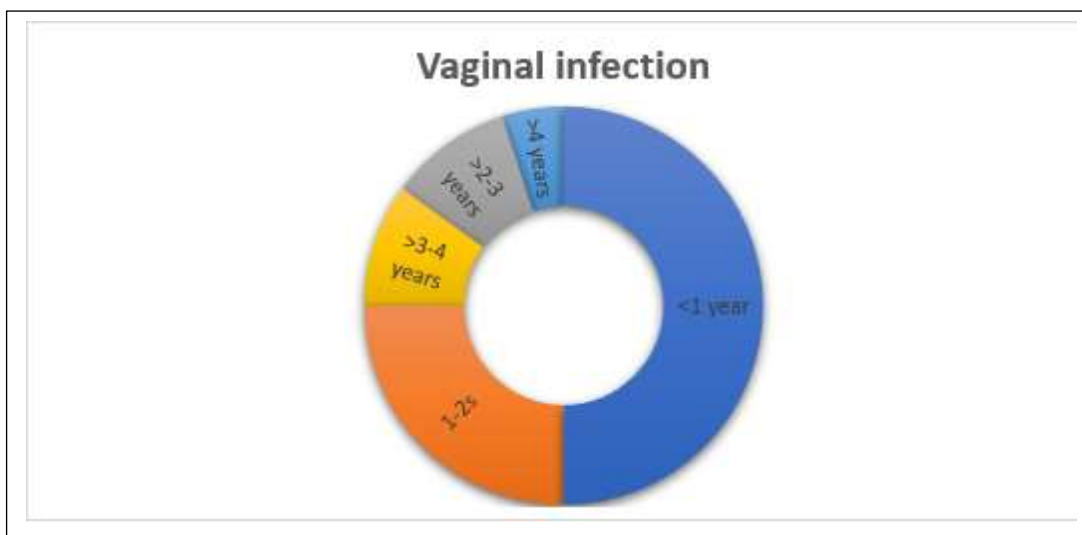
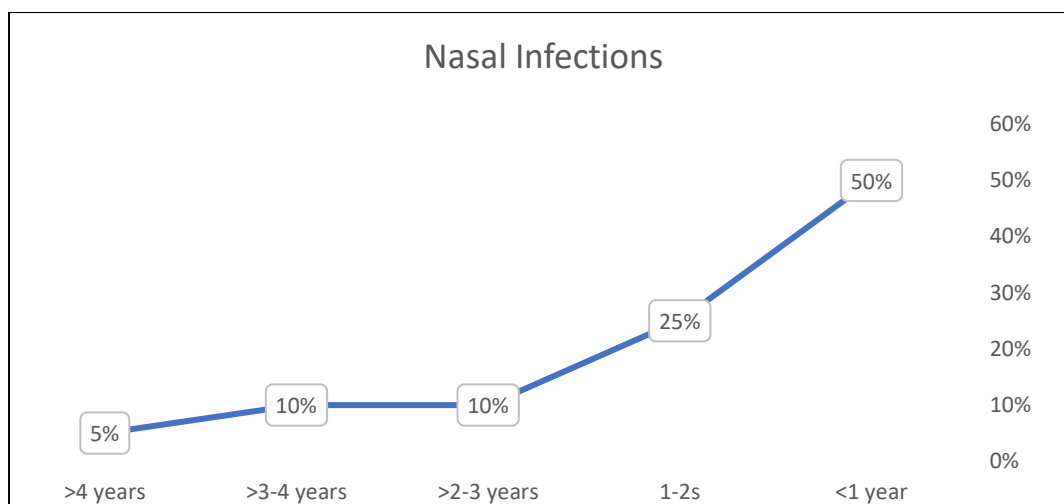


Figure (3): the correlation based on Eta test between age groups and the vaginal infection

Regarding to the nasal infections, this study found that the distribution of the nasal infection among the different age groups was statistically significant ($P < 0.05$). The highest nasal infection rate (50%) was found in the smaller age group (<1 year old), followed by the 25%, which accounted for the age group (1-2 years old). On the other hand, the age groups >2-3 years and >3-4 years recorded 10% for each, and finally the lowest infection rate was found in the older age group (>4 years). Statistically, there is an excellent correlation (Eta value = 96%) between the age groups and the nasal infection rate. Figure (4)

Table (3): the distribution of nasal infection among different cow age groups

Age	Positive No.	Percent	P-value (X2)
<1	20	50%	000 (27.00)
1-2year	10	25%	
>2-3 years	4	10%	
>3-4 years	4	10%	
>4	2	5%	
Total	40	100%	



Figure(4): the distribution of nasal infection among the different age groups, an excellent correlation (eta value = 96%)

This study found no significant differences ($p>0.05$) between the positivity of VitekII and PCR technique in identifying *K. pneumonia*, as both techniques showed only 4/54(7.40%) were *K. pneumonia*. Table (4).

Table (4): the positivity of VitekII and PCR in identification of *K. pneumonia* from different cow samples

Screening Techniques	Positive	Negative
VitekII	4	54
PCR	4	54
P-value	0.642	

Fisher exact test

Vitek II system for identification of *Klebsiella pneumonia*:

Klebsiella pneumonia identification by vitek II system rapidly in this approach with environment free of kind of pollution that may have prevented the presence of the pathogen from being identified in any other way. The result from this system's testing corroborated those from the morphological and PCR test.

Table(5):This table showed the vitek result of present of klebsiella

samples	Total number	Positive+	Percent	Negative-p
Milk	20	2	20%	18
Nasal secretion	20	2	20%	18
Vagina secretion	18	0	0%	18

RESULTS OF PCR TECHNIQUE

The results of PCR technique showed the genomic DNA of all *Klebsiella pneumoniae* samples showed in Fig (5).

The present results recorded that (4/4;100%) of *K. pneumoniae* isolates giving positive results for 16SrRNA gene. The bands of this gene which determined the size of 16SrRNA gene, nearly 1500bp, as shown in Fig. (6). While, K1,K2 genes were not detected in all samples of *K. pneumoniae*. Also, completely isolates of *K. pneumoniae* harbored blaSHv gene (100%), as in fig (7)

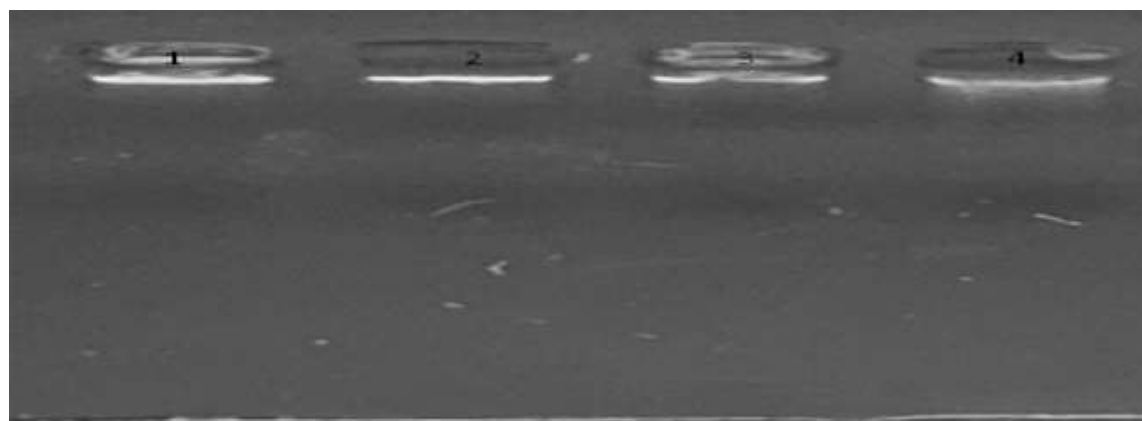


Fig (5): genomic DNA of *K. pneumoniae*

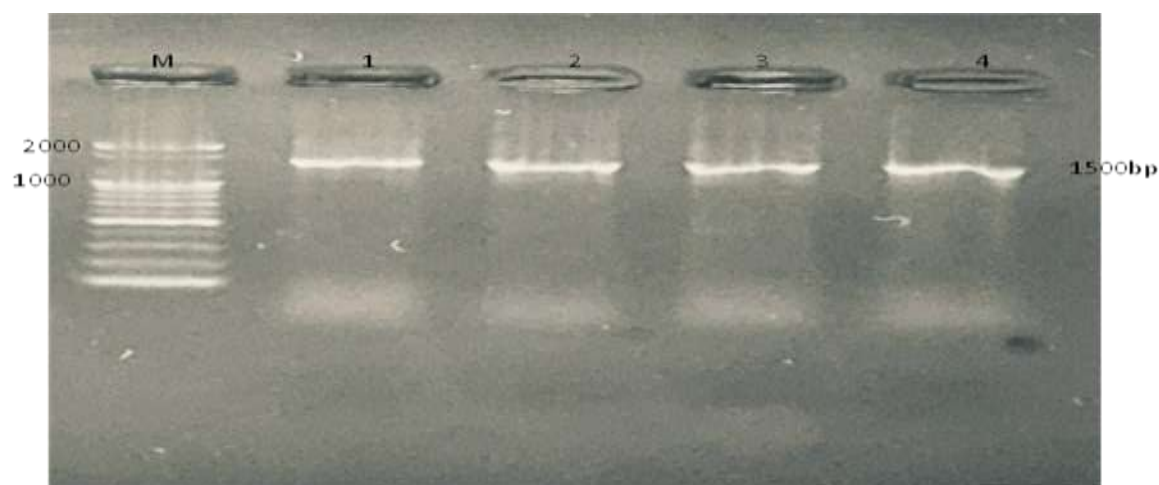


Fig. (6): Agarose gel electrophoresis of 16SrRNA gene amplification, where M: ladder, 1-4: positive results.

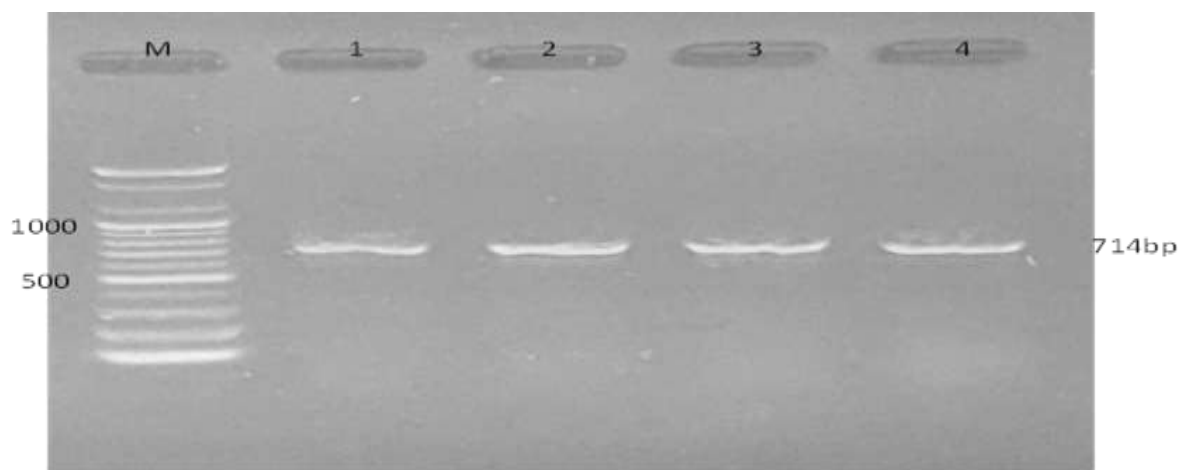


Fig. (7): Agarose gel electrophoresis of blaSHV gene amplification, where M: ladder, 1-4: positive results.

Mutation result show most genetic variation in 16s rRNA

Sample Variant Position in the PCR fragment

Sample	Variant	Position in the PCR fragment
S2,S3	C7T	7
S1,S4	C87T	87
S2,S3	T103A	103
S2,S3	A122T	122
S2,S3	A130G	130
S2,S3	A154T	154
S2,S3	C338T	338
S2,S3	C359G	359
S1,S4	Gins360	360
S2,S3	Ains360	360
S1,S4	G361A	361
S2,S3	G365A	365
S2,S3	A373C	373
S2,S3	C375T	375
S1,S2,S3,S4	C377T	377
S1,S2,S3,S4	Gins378	378
S2,S3	T379C	379
S2,S3	T380C	380
S2,S3	G381C	381
S2,S3	C494T	494
S2,S3	C498T	498
S2,S3	G504A	504
S2,S3	T541C	541

S2,S3	C544T	544
S2,S3	G553A	553
S2,S3	A557T	557
S2,S3	T564C	564
S2,S3	A649G	649
S2,S3	A719C	719
S2,S3	T730C	730
S2,S3	A778G	778
S2,S3	G791A	791
S1,S2,S3,S4	T1078C	1078
S1,S2,S3,S4	G1079T	1079

The phylogenetic tree as shown in figure (3) described the most relative sequences to studied samples were found in very closed related with *K. pneumoniae* isolates. The samples (S1 and S4) were closely related in this phylogenetic tree with (KT588650.1, CP166017.1, PQ432922.1 *K. pneumoniae*), with adjacent branch; those samples were binding with other branch with ref sequence.

While, samples (S2,S3) were closely related in the current phylogenetic tree, and two samples separated from compared samples with separated branch. The results of phylogenetic tree and genetic variation detected by DNA sequencing improved the results of similarity and relation between (S2, and S3) sample.

DISCUSSION

Isolation of *Klebsiella* isolates:

One of the most significant opportunistic pathogens in animals, *Klebsiella pneumoniae* mostly causes mastitis and respiratory infections. In a recent research. The samples were examined for presence of *Klebsiella* isolates. A total of 120 clinical samples and swabs were included (40 from milk, 40 from nasal discharge and 40 from vagina discharge) about 62 of these samples and swabs are no growth and 58(48%) of bacteria culture belong to different Gram negative bacteria .

From the milk samples were examined and the results, referred that 20 (50%) which contained bacterial isolates, on the other hand, 20 (50%) of samples were negative for bacterial isolates. From positive cultures of milk samples several Gram negative bacterial isolates were identified such as *E.coli* 5 (25%), *Enterobacter cloacae* complex 3(15%), *Leclerciaadecarboxylata* 2(10%), *Klebsiella pneumoniae* 2(10%) *Acinetobacter* spp. 2(10%) *Stenotrophomonas maltophilia* 2 (10%), while other bacterial isolates were *Pseudomonas alcaligenes*, *Providencia rettgeri*, *Pseudomonas stutzeri*, and *Pseudomonas aeruginosa* 1(5%) respectively .

From nasal swabs the total numbers were 40 samples. no growth was 20 the other negative bacteria 20(50%) include *E.coli* 13(65%), *Burkholderia cepacia* 1(5%), *Coronobacter sakazakii* 1(5%), *Achromobacter denitrificans* 1(5%), *Pseudomonas alcaligenes* 1 (5%), *Burkholderia gladioli* 1(5%) and *Klebsiella pneumoniae* 2(10%).

From 40 vaginal swabs isolated the present study involved no growth were in 22 swabs, while the positive numbers were 18(45%) included *E.coli* 13(72,5%), *Escherichia hermannii* 1(5,5%), *Achromobacter lowffii* 1(5,5%), *Aeromonas hydrophila* 1(5,5%), *Pseudomonas aeruginosa* 1(5,5%) and *Enterobacter cloacae* complex 1(5,5%).

Klebsiella pneumoniae were isolated from milk samples in rate 2/20 (10%). On the other hand, it was isolated from nasal discharge in rate 2/20 (10%). Nevertheless, *K. pneumoniae* could not be isolated from all vaginal secretion 0/18 (0%).

This result concurred nearly similar to Ramadan, M. M. (2023) which was examined samples and swaps from cattle and showed that *Klebsiella pneumoniae* number was in nasal swaps 24/233 (10.3%) and 5/60 (8.33%) milk and no recorded of *Klebsiella pneumoniae* from vagina secretion. The current study which agreement to other study done by Chiaverini and his group (2022), who examined samples from wild animals and the *Klebsiella pneumoniae* isolate recorded 13/130 in rate 13%. In addition to that, he detected the same cultural characters of *Klebsiella* species and recorded that the isolated bacteria appeared as pink, large, mucoid colonies on MacConkey agar for 24 h at 37 °C.

Higher isolation rate of *Klebsiella* species from different sources most of them were from pig, cattle, milk and the rest of isolates were isolated from vegetables, pets, livestock and farm animals (Klaper et al., 2021)

Identification of *Klebsiella pneumoniae* by culturing and colony morphology and staining reaction

The bacteria were originally identified from the colony, which is a big, spherical, regular-edged colony with a mucous membrane texture and pink due to the acids generated by *Klebsiella* on the agar from MacConkey when the microbes fermented the lactose, after being incubated on the agar at 37 degrees Celsius for 24 h. Identification of *Klebsiella* spp. by using selective enriched media involved cultivating materials on MacConkey agar with crystal violet and bile salts which is favor to growth of Enterobacteriaceae and similar enteric Gram-negative rod while discouraging the development of Gram-positive bacteria and other very fussy Gram-negative bacteria. In this media the presence of lactose can be differentiation between lactose-fermenting bacteria from non-fermenting bacteria. Fermenting bacteria when the pH is low enough, turns colonies pink from the breakdown of neutral red indicator dye.

Klebsiella pneumoniae under a microscope were confirmed to be non-motile, Gram-negative, tiny straight rods that were either single or in pairs, as previously described by (Garritty et al., 2005) which similar to Dubey et al (2013).

Antibiotic resistance of *Klebsiella pneumoniae* isolates:

In this study the isolated strain of *Klebsiella pneumoniae* tested against 14 anti-biotic with various mechanisms of action such as (Ampicillin/ sulbactam, Piperacillin/Tazobactam, Cefotaxime, Ceftazidime, Ceftazidime/Avibactam, Ceftazidime/Tazobactam, Cefepime, Imipenem, Meropenem, Amikacin, Gentamicin, Ciprofloxacin, Tigecycline and Trimethoprim/sulfamethoxazole). The result of this study revealed that all isolated of *Klebsiella pneumoniae* exhibited sensitive to all this antibiotic in rate 100%.

The other studies showed that *Klebsiella pneumoniae*'s resistance rate to some antibiotic as the following (ampicillin 98.1%, ciprofloxacin 35.9%,

trimethoprim/sulfamethoxazole 35%, cefotaxime 34%, gentamicin 31.1%,

tetracycline 31.1%) according to Haradet et al (2016).

Moreover, Gomaa (2021) verified that all *Klebsiella pneumoniae* isolates were susceptible to the drugs a antibiotic such as trimethoprim/sulphamethoxazole and trimethoprim (54.54% each), but resistant to the antibiotic ampicillin and amoxicillin-clavulanic acid (100%) following cefepime (72.72%), imipenem (82%) followed by aztreonam (55%), and amikacin and azithromycin (45%).

Montsoetal. (2019) showed that The highest multi drug resistance rate were found among klebsiellaisoltes from cattle in rate reached to (85–100%)

Brise and Duijkeren (2005) found that from 78 isolates, resistance was common against ampicillin (99%) and cephalexin (43%), but not against tetracycline, ceftazidime, , enrofloxacin, ceftiofu, gentamicin and trimethoprim/sulfamethoxazole.

Ramadan, M. M. (2023) study showed that the biofilm formation module indicated that all strains were biofilm formed strains.

It was shown that 81.81% of the pneumonia (*K. pneumonia*) isolates were high (33.33%), moderate (22.22%), or weakly (44.44%) polymeric producers, compared to 18.18% of the total number of isolates that did not form biofilms (Gomaa, 2021). This result supports the findings of Massé et al. (2020).

The objective of the current study was to identify the evolutionary relationships by phylogenetic tree of all samples of *Klebsiella pneumoniae* isolated from cattle's milk and nasal secretions, as well as to identify genetic variants of the 16SrRNA genes by DNA sequencing.

Life-threatening illnesses are caused by *Klebsiella pneumoniae*. Nonetheless, the organisms have been linked to the onset of sickness (Villegas et al., 2006). Reports that bacteria that cause *Klebsiella pneumoniae* can spread swiftly across wide geographic areas are especially worrisome (Navon-Venezia et al., 2009).

The 16SrRNA gene sequencing used to identified the *K. pneumoniae* that isolated from various sources as samples obtained from animal, or from clinical isolates such as urine, blood, feces, sputum, which were aligned and compared to the sequence of strains that deposited in GenBank database (Budiarsoetal., 2021).

Despite their potential health risks, street meals are popular in nations that are developing due to their low prices and preference among the lower and middle classes. According to Rowbotham and Ruegg (2016) and Fuenzalida and Ruegg (2019), *Klebsiella pneumoniae* and *Klebsiella oxytoca* are commonly found in farm surroundings, cow skins and the milk, teet-end cotton swabs and clinical mastitis. Additionally, the *Klebsiella pneumoniae* species was a significant pathogen linked to the health of both humans and animals. Urinary tract infections, liver abscesses, and pneumonia have all been linked to *Klebsiella pneumoniae* (Bengoechea and Pessoa 2019; Fuenzalida and Ruegg 2019).

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CONCLUSION

Klebsiella pneumoniae isolated from nasal secretion, milk secretion, and not found in vagina discharge. *Klebsiella pneumoniae* can be identified directly by vitek II system and confirmed by PCR to detect 16S rRNA. All *Klebsiella pneumoniae* isolated opened to be sensitive to most antibiotic tested. *Klebsiella pneumoniae* isolated does not have K1, K2 and contain blaSHv gene.

REFERENCE

1. Paczosa, M. K., & Mecsas, J. (2016). *Klebsiella pneumoniae*: going on the offense with a strong defense. Microbiology and molecular biology reviews, 80(3), 629-661.

2. Choby, J. E., Howard-Anderson, J., & Weiss, D. S. (2020). Hypervirulent *Klebsiella pneumoniae*—clinical and molecular perspectives. *Journal of internal medicine*, 287(3), 283-300.
3. Vulikh, K., Bassel, L. L., Sergejewich, L., Kaufman, E. I., Hewson, J., MacInnes, J. I., ... & Caswell, J. L. (2019). Effect of tracheal antimicrobial peptide on the development of Mannheimia haemolytica pneumonia in cattle. *PLoS One*, 14(11), e0225533.
4. Schukken, Y. H., Zurawski, M. J., Rauch, B. J., Gross, B., Tikofsky, L. L., & Welcome, F. L. (2013). Noninferiority trial comparing a first-generation cephalosporin with a third-generation cephalosporin in the treatment of nonsevere clinical mastitis in dairy cows. *Journal of Dairy Science*, 96(10), 6763-6774.
5. Kumari, A., Tanwar, S., Tyagi, S., & Kumar, N. (2018). Fog computing for Healthcare 4.0 environment: Opportunities and challenges. *Computers & Electrical Engineering*, 72, 1-13.
6. Ramadan, M. M. (2023). Identification and characterization of klebsiella pneumonia isolated from farm animals and their biofilm production estimation. *Benha Veterinary Medical Journal*, 44(2), 70-73.
7. Chiaverini, A., Cornacchia, A., Centorotola, G., Tieri, E.E., Sulli, N., Del Matto, I., Iannitto, G., Petrone, D., Petrini, A., Pomilio, F. 2022. Phenotypic and Genetic Characterization of
8. *Klebsiella pneumoniae* Isolates from Wild Animals in Central Italy. *Animals*, 12(11), 1347.
9. Klaper, K., Hammerl, J.A., Rau, J., Pfeifer, Y., Werner, G. 2021. Genome-Based Analysis of *Klebsiella* spp .Isolates from Animals and Food Products in Germany. *Pathogens (Basel, Switzerland)* 10, 2013-2017..
10. Yang, Y., Peng, Y., Jiang, J., Gong, Z., Zhu, H., Wang, K., ... & Shang, S. (2021). Isolation and characterization of multidrug-resistant *Klebsiella pneumoniae* from raw cow milk in Jiangsu and Shandong provinces, China. *Transboundary and Emerging Diseases*, 68(3), 1033-1039.
11. Harada, K., Shimizu, T., Mukai, Y., Kuwajima, K., Sato, T., Usui, M., Tamura, Y., Kimura, Y., Miyamoto, T., Tsuyuki, Y. 2016. Phenotypic and molecular characterization of antimicrobial resistance in *Klebsiella* spp. isolates from companion animals in Japan: clonal dissemination of multidrug-resistant extended-spectrum β -lactamase-producing *Klebsiella pneumoniae*. *Frontiers in microbiology* 7, 1021.
12. Virginia Villegas, M., Lolans, K., Correa, A., Jose Suarez, C., Lopez, J. A., Vallejo, M., & Quinn, J. P. (2006). First detection of the plasmid-mediated class A carbapenemase KPC-2 in clinical isolates of *Klebsiella pneumoniae* from South America. *Antimicrobial agents and chemotherapy*, 50(8), 2880-2882.
13. Navon-Venezia, S., Leavitt, A., Schwaber, M. J., Rasheed, J. K., Srinivasan, A., Patel, J. B., & Carmeli, Y. (2009). First report on a hyperepidemic clone of KPC-3-producing *Klebsiella pneumoniae* in Israel genetically related to a strain causing outbreaks in the United States. *Antimicrobial agents and chemotherapy*, 53(2), 818-820.
14. Budiarto, T Y., Amarantini, Ch., and Pakpahan, S. (2021). Biochemical identification and molecular characterization of *Klebsiella pneumoniae* isolated from street foods and drinks in Yogyakarta, Indonesia using 16SrRNA gene. *Biodiversitas*, 22(12): 5452-5458.
15. Fuenzalida, M. J., & Ruegg, P. L. (2019). Longitudinal study of quarter-level somatic cell responses after naturally occurring, nonsevere clinical mastitis diagnosed as culture negative, or caused by *Escherichia coli* or *Klebsiella pneumoniae*, and randomly assigned to a no-treatment group or to receive intramammary ceftiofur. *Journal of Dairy Science*, 102(12), 11476-11482.
16. Liu Z, Lozupone C, Hamady M, Bushman FD, Knight R. Short pyrosequencing reads suffice for accurate microbial community analysis. *Nucleic Acids Res.* 2007;35(18):e120. doi:10.1093/nar/gkm54123 .
17. Nee, C. W. (2020). Bovine mastitis: risk factors, therapeutic strategies, and alternative treatments. *Animal Bioscience*, 33(11), 1699-1713.