

# Aonla (*Emblica Officinalis* Gaertn): The Comprehensively Used Medicinal Plant Of Mizoram

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## ABSTRACT

The locally available fruit Aonla possesses many usages ethnically in Mizoram. Inference can be made from this study that the size of the plants, fruits as well as its moisture content showed no variation from different places. It contains various phytochemicals such as tannins, saponin, flavanoids, steroids, terpenoids, and cardiac glycosides. The results showed that the total phenolic content and flavonoid content of different plant extracts exhibit no significant variation ranging from  $68.92 \pm 1.1$  to  $71.8 \pm 1.2$  mg GAE/g, and total flavonoid content was estimated at the range of  $25.25 \pm 3.00$  to  $35.14 \pm 1.53$  mg QUE/g dried tissue though there were slight variations. The DPPH radical scavenging activity depends on the concentration of the extracts. The scavenging activity increases with the increase in concentration with an optimum concentration at  $40 \mu\text{g/ml}$  for all the plants extracts. The antimicrobial activity revealed that the *E. officinalis* has potential property to inhibit the growth of *S. aureus* and *P. aeruginosa*. However, it was found ineffective against *E. coli*. The molecular identification was performed based on NCBI.

**Keywords:** Antioxidant, *Emblica officinalis*, DPPH, Mizoram, Phytochemicals.

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## INTRODUCTION

Aonla (*Emblica officinalis* Gaertn) belongs to the family Euphorbiaceae and is found in the subtropical and tropical areas of South-east Asian countries. Its fruit is widely consumed for its special taste. It is found to possess an ample amount of vitamin C and superoxide dismutase (Verma and Gupta, 2004) and is employed in many ethnomedicinal systems like herbal medicine of Chinese, Tibetan medicine and Ayurvedic medicine (Zhang et al., 2000). This plant is known to be antihyperlipidemic and hypoglycemic (Anila and Vijayalakshmi, 2000; Abesundara et al., 2004), and is an important component of many of the presently used hepato-protective formulations (Panda and Kar, 2003). It is also used as an antimicrobial, anticancer, and anti-inflammatory agent (Dutta et al., 1998; Jeena et al., 2001; Asmawi et al., 1993) and can remedy the - clastogenic effects induced by heavy metals (Biswas et al., 1999). *E. officinalis* was reported to have a strong antioxidant activity (Bafna and Balaraman, 2004), this could be related to the presence of flavonoids and numerous gallic acid derivatives, such as epigallocatechingallate. (Anila and Vijayalakshmi, 2002).

Aonla branchlets are not smooth, feel furry due to the small hair projections and are about 10–20 cm long. The leaves are simple, feathery, linear oblong in shape and closely arranged branchlets, light green in colour, resembling pinnate leaves. The flowers range from greenish to yellow which appears with the commencing of spring season. When ripe, the fruit is nearly spherical, pale greenish to yellow in colour, smooth and hard to the touch, with six vertical stripes or furrows, and a sour or bitter taste. (Indian Medicinal Plants, 1997).

The Euphorbiaceae are medium to large deciduous plants. *E. officinalis* is commonly known as Amla or Emblic myrobalan. The tree is found throughout tropical and sub-tropical India. It is a medium-sized deciduous tree and can reach upto 18 m in height and is native to India. It bears grayish brown bark which grows well in different agro-climatic and different soil conditions (Warrier et al., 2019; Rekha et al., 2019).

Mizoram, a state of north-east India covers an area between latitude  $21^{\circ}58' - 24^{\circ} 35' \text{ N}$  and longitude  $92^{\circ} 15' - 93^{\circ} 29' \text{ E}$  with 86.27% forest coverage (Rintluanga, 2019). The elevation varies from 500 to 2157 metres. Mizoram has a moderate and mild climate, with temperatures ranging from  $20^{\circ}\text{C}$  to  $29^{\circ}\text{C}$  ( $68^{\circ}\text{F}$

to 84°F) in the summer and 7°C to 22°C (45°F to 72°F) in the winter. The rainy season has an impact on the region, which sees strong rains during the Monsoon. The geographical condition is attributed with a mixed climatic condition ranging from moist tropical to moist sub-tropical, with an average rainfall of 254 centimeters (100 in) per annum. The average rainfall in Aizawl, the capital is approximately 215 centimeters (85 in) (Geological Survey of India, 2011; Champion and Seth, 1968).

The people of Mizoram largely rely on herbal drugs which have been passed down for generations traditionally. The crushed bark of Aonla is applied to wounds and cuts. The decoction of bark is used to treat diarrhea and dysentery (Bhardwaj and Gakhar, 2005; Rai and Lalramnghinglova, 2010; Lalramnghinglova, 2016). Its fruits were reported to possess a laxative effect. A liquid of fermented fruits are used to improve digestion, anaemia, jaundice, liver disorder, urination and also for certain heart ailments. After childbirth, fruit decoction is used to discharge retained placenta and cure skin eruptions due to food allergy. Mizoram is immensely blessed with Aonla tree (Lalzarzovi and Lalramnghinglova, 2016). Therefore, it was desired to undertake the scientific study on their ethno-medicinal properties.

## MATERIALS AND METHODS

### *Collection and physical parameters of plants*

The *E. officinalis* from different locations based on height, girth (50 cm above the ground) and canopy of 10 plants from Aizawl District were collected. The *E. officinalis* was authenticated by Prof. H. Lalramnghinglova with voucher number MZU-HAMP001241, MZU-HAMP001245, MZU-HAMP001246, MZU-HAMP001248. The weight of fruit and seed of the plants were also measured and recorded. The leaves of four most distinct plants were taken from Sihphir, Tuirial, Tanhril and Falkawn for different tests.

### *Estimation of moisture content*

The fresh leaves of *E. officinalis* were collected and the weight was measured. Then the known weight of the sample was subjected for heat treatment at 70°C for 48 hrs. The moisture content was calculated using the formula (Allen et al., 1974).

$$\% \text{ MC} = \frac{(\text{MF} - \text{MD})}{\text{MD}} \times 100$$

MD

Where, MC - moisture content, MF- fresh weight, MD - weight after drying, % MC - percent of moisture.

### *Preparation of extracts*

The fresh *E. officinalis* leaves were washed, air-dried at room temperature for 24h and grinded to powder using a grinder. 250g of the powder was extracted in 500 ml of methanol and chloroform for 24h at 25°C, in a glass conical flask using a shaker and filtered using a 0.45 µm filter paper. The residue was then extracted twice with solvents such as 100 ml methanol and chloroform as described above. Both the methanol and chloroform extracts were concentrated at 40°C, using a rotary evaporator under low pressure. The extracts were dried and then stored in air-tight containers at 4°C until further use.

### *Preliminary phytochemical screening*

The phytochemicals such as saponin, flavanoids, terpenoids, cardiac glycosides, tannins and phlobatannins were carried out using standard protocols to identify the constituents as described by (Sofowara, 1993; Trease and Evans, 1998; Harborne, 1998).

### *Determination of the total phenolic and flavonoids contents*

#### *Total Phenol contents*

The total phenols were determined by Folin-Ciocalteu reagent (McDonald and Prenzler, 2001). A dilute extract of the leaves 0.5 ml (1 mg/ml) was reacted with Folin-Ciocalteu reagent (5ml, 1:10 diluted with distilled water) and aqueous Na<sub>2</sub>CO<sub>3</sub> (4ml, 1M). The mixture was incubated for 15 mins and the phenol content was measured at 756 nm using a UV/Visible double beam spectrophotometer. The total phenolic content was expressed in terms of gallic acid equivalent (mg/gm of dry weight), which is a common reference compound.

### ***Total flavonoids contents.***

Aluminium chloride technique was used for determining flavonoids (Chang et al., 2002). 0.5 ml (1 mg/ml) leaf extract was combined with 1.5 ml methanol, 0.1 ml 1 M potassium acetate, 0.1 ml 10% aluminium chloride, and 2.8 ml distilled water for each concentration. It was then incubated for 30 minutes at room temperature. The amount of flavonoid was quantified through the optical density at 415 nm with a UV/Visible double beam spectrophotometer.

### **Antioxidants Activities**

#### ***DPPH scavenging activity***

The scavenging activity of DPPH free radical was performed (Leong and Shui, 2002) with minor modifications. 0.5 ml of each of the different concentrations (5, 10, 20, 40 and 80 µg/ml) of the *E. officinalis* extract was added to 1 ml solution of 0.1 mM DPPH. It was mixed thoroughly and incubated in the dark for 30 mins and the absorbance at 523 nm was taken using methanol for the baseline correction. The results were compared with that of the control prepared as above without sample. Radical scavenging activity was expressed as a percentage and was calculated using the following formula:

$$\% \text{ Scavenging} = \frac{(A_{\text{control}} - A_{\text{sample}})}{A_{\text{control}}} \times 100$$

Where,  $A_{\text{sample}}$  is the absorbance of the test sample and  $A_{\text{control}}$  is the absorbance of the control.

#### **Antibacterial activity (Agar diffusion method)**

Antibacterial activities of the four plant extracts were tested against three pathogenic microbial strain viz *Escherichia coli*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*. Muller – Hinton agar media (Hi-Media) was used for performing the test. After the preparation of the media, it was poured onto the petri plates and allowed to solidify. Then 4 bores (3 mm depth, 4 mm diameter) were made using a sterile borer. The three pathogenic microbial strains were spread onto the plates using sterile spreaders. The crude plant extract were loaded on the bores (100 µl). The petriplates were incubated at 37°C for 24 h. Evaluation of Antimicrobial activity was performed by measuring the diameter of the growth inhibition zones using zone scale.

#### ***Molecular identification using Matk gene***

The DNA was isolated from the leaves of four selected *E. officinalis* plant using Qiagen DNeasy Plant Mini Kit following the instruction given. The commonly used primer; matK390F (5'-CGA TCT ATT CAT TCA ATA TTT C-3') and 1326R (5'-TCT AGC ACA CGA AAG TCG AAG T-3') (Schmitz-Linneweber et al., 2002) was used for molecular identification of the plants.

#### **Statistical Analysis**

Microsoft excel 2010 was used for analysis of the data and graph pad prism for significant test. The laboratory experiments data were obtained in triplicates results, n=3. A p<0.05 value was considered significant.

## **RESULTS**

There was no significant difference among the selected plants regarding the physical factors such as height, girth, fruit weight and seed weight. (Table 1). However, the canopies were found to differ based on the location where Thingsulthlah, Falkawn, Tuirial, Tanhril, Saikhamakawn, Hualngohmun and Sihphir were lower as compared to the plants found in Muthi, Hlimen, Seling were more elevated from the ground. This could be due to the effect of geographic location. (Fig 1).

The moisture content of *E. officinalis* in different locations did not show significant variation. The moisture content was highest in the plant collected from Tanhril (Plant 3) with 52.26% (Table 2). The phytochemicals such as tannins, saponin, flavanoids, steroids, terpenoids, cardiac glycosides were present but phlobatannins and alkaloid were absent in all the plants (Table 3). Likewise, total phenolic and flavonoid content in different plant extracts exhibited no significant variation. Both the total phenol and flavonoid content were found to increase with increase in concentration. The total phenol content ranged from 68.92±1.1 to 71.8±1.2 mg GAE/g with the highest content found in Tuirial (Plant 2) with 71.8 g/g

tissue (Figure 2).

The total flavonoid content was estimated in the range from  $25.25 \pm 3.00$  to  $35.14 \pm 1.53$  mg QUE/g tissue and the highest flavonoid content was found in Falkawn (Plant 4) with 35.14 mg/g tissue. The reduction of DPPH radical was dependent on the concentration of the extracts. Increase in concentration of the sample showed rise in the scavenging activity. The optimum concentration was obtained at the concentration of 40  $\mu$ g/ml for all the extracts and decreased hereafter. The maximum scavenging activity observed were- Sihphir (Plant 1) – 89.02%, Tuirial (Plant-2) – 89.07%, Tanhril (Plant -3) – 92.07% and Falkawn (Plant - 4) -92.12% respectively (Figure 3). Hence, the *E. officinalis* collected from Falkawn (Plant 4) was found to exhibit the highest scavenging activity among the four different locations. The antimicrobial activity revealed that the *E. officinalis* extract has potential property to inhibit the growth of *S. aureus* and *P. aeruginosa* at certain concentrations. However, there was no activity against *E. coli* (Table 4).

However, the raw sequence obtained was edited and analysed using MEGA7, the sequenced were submitted to Genbank, NCBI ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)). The genetic analysis and phylogenetic tree was constructed.

Using the Maximum Likelihood approach and the General Time Reversible model, the evolutionary history was inferred. It is shown the tree with the highest log likelihood (-3318.91). The initial tree(s) for the heuristic search were automatically generated by applying the Neighbour-Joining Method and BioNJ algorithms to a matrix of pairwise distances calculated using the Maximum Composite Likelihood (MCL) approach, and then selecting the topology with the highest log likelihood value. The rate variation model allowed for some sites to be evolutionarily invariable ([+I], 34.83% sites). The number of substitutions per site is used to calculate branch lengths, and the tree is depicted to scale. There were 24 nucleotide sequences in this study. 1st+2nd+3rd+Noncoding codon locations were included. The total number of positions in the final dataset was 877. Evolutionary analyses were conducted in MEGA X ( Schmitz-Linneweber et al.,2002).

## DISCUSSION

In developed countries, many people turn to herbal therapy and spend a large sum of money since it is largely believed that it promotes healthier living. Herbal medicines are, therefore, often viewed as a more mild and moderate approach balancing to healing for individuals who use them as home remedies as compared to over-the-counter drugs. This could be the reason for the increase in sales of herbal medicines which represents a substantial proportion of the global drug market (Ekor, 2013; Leong and Shui, 2002). Therefore, it was decided to analyse the phytochemical constituents of Aonla (*E. officinalis*) plants which have been used extensively for various medications in Mizoram. The Aonla plants from different locations within Aizawl district did not show significant variation in terms of physical parameters such as height, girth, fruit weight and seed weight (Table 1). Moreover, the moisture content of *E. officinalis* Gaertn in different locations was found to be within the range of 44.65 to 52.26 % (Table 2). The phytochemical analysis revealed that various secondary metabolites including tannins, saponin, flavanoids, steroids, terpenoids and cardiac glycosides were present (Table 3). The antimicrobial activity against *S. aureus* and *P. aeruginosa* showed good activity (Table 4). The phenolics and flavonoid content were comparatively high in terms of percentage (Fig 2) with high DPPH scavenging activity (Fig 3). Plant phenols have a strong antioxidant potential and are a common dietary supplement. It has been found that the DPPH free radical's scavenging effect increases with the concentration of the sample and standard to a certain extent, and so is highly dependent on the extract concentration. (Patel et al., 2010; Chanda et al., 2010; Ullah et al., 2016). Several other plants such as *Aporosa octandra*, *Castanopsis indica*, *Dysoxylum gobara*, *Eleagnus caudata*, *Milletia pachycarpa*, *Oroxylum indicum* and *Schima wallichi* which contain high phenolic and flavonoid content have also been found to exhibit DPPH scavenging activity (Vabeiryureilai et al., 2014; Lalrinzuali et al., 2015). Phenolics have been a center of attraction as potential agents to prevent and treat many oxidative stress-related diseases (Lalrinzuali et al., 2015; Gu and Weng, 2001; Pyo et al., 2001). There was slight variation in the phenolic and flavonoid content as well as the DPPH scavenging activity

in the leaves extract of *E. officinalis* plants collected from different locations. A similar result have been observed in *Moringa olifera* collected from different places. The molecular analysis confirmed the species which has similarity 99.4 with 100% query coverage (Fig 4) (An-Na et al., 2014; Panche et al., 2016; Ndhala et al., 2014). In addition to this, a previous study conducted in relation to pharmac phylogenetic on the essential oil of *M. biflora* Benth and *C. reticulata* Blanco showed good inhibitory activity against the tested water-borne bacterial pathogens (Kumar et.al., 2012). The medication of Aonla as ethno-medicine might be due to the fact that its phytochemical content is of biological origin and have also been investigated since the advent of human history (Jagetiya et al., 2005; Lalrinzuali et al., 2016). The Aonla contained high flavonoid. It was reported that more than 8000 flavonoids have been isolated from various plants. The flavonoids have a major contribution for the attractive colours of flowers with anthocyanins (Ghasemzadeh and Ghasemzadeh, 2011; Iwashina, 2015). They are also a vital component in stimulus, defense, taste, pigmentation and in plant-microorganisms relationship (Hasan et al., 2009). The antioxidant activity depends mostly on the compounds present in the plants. It expresses the amount of antioxidant needed to neutralize the unstable free radicals in the system (Pratt and Hudson, 1990). The assessment of the ability to scavenge DPPH is a very useful method to estimate the antioxidant activity. It is violet colour and has an unpaired electron in the outermost shell which turns yellow after accepting electron from any antioxidant (Goldschmidt and Renn, 1922). The leaf extracts of *E. officinalis* was found to scavenge DPPH free radical and the activity was found to increase with increasing concentration of flavonoids. A similar result has been observed earlier (Lalrinzuali et al., 2015). *E. officinalis* can also be used as a possible food-additive or in nutraceuticals and biopharmaceutical industries. Several other reports have also revealed that the various extracts and herbal formulations of Aonla showed potential therapeutic benefits against various diseases and the results are similar to those drugs in use (Grover et al., 2015).

## CONCLUSION

The Aonla (*Emblia officinalis*) leaf holds many applications ethnically. It contains various phytochemicals like tannins, saponin, flavanoids, steroids, terpenoids and cardiac glycosides, shown high phenolic and flavonoid content and increased scavenging activity. Moreover, it can be a suitable candidate for the antimicrobial activity against *S. aureus* and *P. aeruginosa*. Therefore, it can be used as a possible food-additive or biopharmaceutical or herbal formulation industries in the future.

## Acknowledgement

The authors thanked Advanced Level State Biotech Hub, Department of Biotechnology, Mizoram University, and CSIR, New Delhi for providing necessary facilities and funding to carry out the experiments.

## REFERENCES

1. Abesundara, K.J.M., Matsui T, Matsumoto K, 2004. *A-glucosidase inhibitory activity of some Sri Lanka plant extracts, one of which, Cassia auriculata, exerts a strong anti-hyperglycemic effect in rats comparable to the therapeutic drug acarbose.* J Agri Food Chem , 52: 2541-2545.
2. Allen S.E, Grimshaw H.M, Parkinson J.A, Quarmby C, 1974. *Chemical analysis of ecological materials*, Blackwell Scientific Publications. Osney, Oxford, London.
3. Anila L, Vijayalakshmi NR, 2000. *Beneficial effects of flavonoids from Sesamum indicum, Emblica officinalis and Momordica charantia.* Phytother, 14: 1-4.
4. Anila L, Vijayalakshmi NR, 2002. *Flavonoids from Emblica officinalis and Mangifera indica— effectiveness for dyslipidemia.* J Ethnopharmacol , 79: 81-87.
5. An-Na Li, Sha Li, Zhang Y, Xu XR, Chen YM, Li HB, 2014. *Resources and Biological Activities of Natural Polyphenols.* Nutrient , 6 (12): 6020-6047.
6. Asmawi MZ, Kankaanranta H, Moilanen E, Vapaatalo H, 1993. *Anti-inflammatory activities of Emblica officinalis Gaertn. Leaf extracts.* J Pharm Pharmacol, 45: 581-584.
7. Bafna PA, Balaraman R, 2004. *Anti-ulcer and antioxidant activity of DHC-1, a herbal formulation.* J Ethnopharmacol, 90: 123-127.
8. Bhardwaj S, Gakhar SK, 2005. *Ethnomedicinal plants used by the tribals of Mizoram to cure cuts & wounds.* Ind J Trad Knowl , 4(1): 75-80.

9. Biswas S, Talukder G, Sharma A, 1999. Protection against cytotoxic effects of arsenic by dietary supplementation with crude extract of *Emblica officinalis* fruit. *Phytother Res*, 13: 513-516.
10. [Champion SH, Seth SK, 2005. A revised survey of the forest types of India. Delhi, Manager of Publications.](#)
11. [Chanda SV, Nagani KV, 2010. Antioxidant capacity of Manilkara zapota L. leaves extracts evaluated by four in vitro methods. Nature Science, 8\(10\): 260-266.](#)
12. [Chang C, Yang M, Wen H, Chern J, 2002. Estimation of total flavonoid content in propolis by two complementary colorimetric methods. J Food Drug Anal, 10: 178-182.](#)
13. [Dutta BK, Rahman I, Das TK, 1998. Antifungal activity of Indian plant extracts. Mycoses, 41: 535-536.](#)
14. [Ekor M, 2013. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. Frontiers in Pharmacology, 4: 177.](#)
15. [Geological Survey of India, Government of India, 2011. Geology and mineral resources of Manipur, Mizoram, Nagaland and Tripura: Miscellaneous publication No. 30 Part IV. 1 \(Part-2\).](#)
16. [Ghasemzadeh A, Ghasemzadeh N, 2011. Flavonoids and phenolic acids: Role and biochemical activity in plants and human, J Med Plant Res, 5: 6697-6703.](#)
17. [Goldschmidt S, Renn K, 1992. Amine oxidation IV. Diphenyl-trinitrophenylhydrazyl. ChemBer, 55: 628-643.](#)
18. [Grover HS, Deswal H, Singh Y, Bhardwaj A, 2015. Therapeutic effects of amla in medicine and dentistry: A review. J Oral Res Rev, 7: 65-8.](#)
19. [Gu LW, Weng XC, 2001. Antioxidant activity and components of \*Salvia plebeian\* R.Br. – a Chinese herb. Food Chem, 73: 299-305.](#)
20. [Harborne JB, 1998. Phytochemical methods. A guide to modern techniques of plant analysis. Springer Netherlands. \(3<sup>rd</sup> Edn\), 60.](#)
21. [Hasan SR, Hossain MM, Akter R, Jamila M, Mazumder ME, Rahman S, 2009. DPPH free radical scavenging activity of some Bangladeshi medicinal plants. J Med Plants Res, 3\(11\): 875-9.](#)
22. [Iwashina T, 2015. Contribution to flower colors of flavonoids including anthocyanins: a review. Nat Prod Comm, 10: 529-544.](#)
23. [Jagetia GC, Venkatesha VA, 2005. Effect of mangiferin on radiation induced micronucleus formation in cultured human peripheral blood lymphocytes. Environ Mol Mutagen, 46: 12-21.](#)
24. [Jeena KJ, Kuttan G, Kuttan R, 2001. Antitumour activity of \*Emblica officinalis\*. J Ethnopharmacol, 75: 65-69.](#)
25. [Kumar A, Gupta R, Mishra RK, Shukla AC, Dikshit A, 2012. Pharmaco-Phylogenetic Investigation of \*Micromeria biflora\* Benth and \*Citrus reticulata\* Blanco. Natl. Acad. Sci. Lett, 35\(4\):253-257.](#)
26. [Kumar S, Stecher G, Tamura K, 2016. MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. Mol Biol Evolution, 33: 1870-1874.](#)
27. [Lalramnghinglova H, 2016. Documentation of Medicinal Plants based on Traditional Practices in the Indo-Burma Hotspots Region of Mizoram, North East India. Emer Life Sci Res, 2\(1\): 10-45.](#)
28. [Lalrinzuali K, Vabeiryureilai M, Jagetia GC, 2015. The analysis of antioxidant activity and phenolic contents of selected medicinal plants of Mizoram. Genomics and Applied Biology, 6 \(11\): 1-12.](#)
29. [Lalrinzuali K, Vabeiryureilai M, Jagetia GC, Lalawmpuii PC, 2015. Free radical scavenging and antioxidant potential of different extracts of \*Oroxylum indicum\* in vitro. Adv. Biomed. Pharma, 2\(3\): 120-130.](#)
30. [Lalzarzovi ST, Lalramnghinglova H, 2016. Traditional use of medicinal plants found within Aizawl city in Mizoram, India. Pleione, 10\(2\): 269-277.](#)
31. [Leong LP, Shui G, 2002. An investigation of antioxidant capacity of fruits in Singapore markets. Food Chem, 76: 69-75.](#)
32. [McDonald S, Prenzler PD, Autolovich M, Robards K, 2001. Phenolic content and antioxidant activity of olive extracts. Food Chem, 73: 73-84.](#)
33. [Ndhlala AR, Mulaudzi R, Ncube B, Abdelgadir HA, Plooy CP, Staden JV, 2014. Antioxidant, Antimicrobial and Phytochemical Variations in Thirteen \*Moringa oleifera\* Lam. Cultivars. Molecules, 19: 10480-10494.](#)
34. [Panche AN, Diwan AD, Chandra SR, 2016. Flavonoids: an overview. J Nutr Sci, 5: e47.](#)
35. [Panda S, Kar A, 2003. Fruit extract of \*Emblica officinalis\* ameliorates hyperthyroidism and hepatic lipid peroxidation in mice. Pharmazie, 58: 753-761.](#)
36. [Patel VR, Patel PR, Kajal SS, 2010. Antioxidant activity of some selected medicinal plants in western region of India. Adv Biol Res, 4\(1\): 23-26.](#)
37. [Prakash B, 1997. Indian Medicinal Plants, Department of ISM and Homeopathy, Indian Security Press, Nasik.](#)
38. [Pratt DE, Hudson BJ, 1990. Natural antioxidants not exploited commercially. In Food antioxidants. Springer. Dordrecht, 171-191.](#)
39. [Pyo YH, Lee TC, Logendrac RT, 2004. Antioxidant activity and phenolic compounds of Swiss chard \(\*Beta vulgaris\* subspecies \*cycla\*\) extracts. Food Chem, 85: 19-26.](#)
40. [Rai PK, Lalramnghinglova H, 2010. Ethnomedicinal plant resources of Mizoram, India: Implication of traditional knowledge in health care system. Ethnobot Leaflets, 14: 274-305.](#)
41. [Rekha RW, Joshi G, Arunkumar AN, 2019. DNA fingerprinting in industrially important medicinal trees. Annals of Phytomedicine, 8\(1\): 19-35.](#)
42. [Rintluanga P, 2019. Mizoram: A Study in Comprehensive Geography, ISBN 978-81-7211-264-6.](#)

43. Schmitz-Linneweber C, Maier RM, Alcaraz JP, Cottet A, Herrmann RG, Mache R, 2002. The plastid chromosome of spinach (*Spinacia oleracea*): complete nucleotide sequence and gene organization. *Plant Mol Biol* , 45(3): 307-15.
44. Sofowara A, 1993. Medicinal plants and Traditional medicine in Africa. Spectrum Books Ltd. Ibadan. Nigeria, 289.
45. Tamura K, 1992. Estimation of the number of nucleotide substitutions when there are strong transition-transversion and G + C content biases. *Mol Biol Evolution* , 9: 678-687.
46. Trease GE, Evans WC, 1989. Pharmacognosy. (11<sup>th</sup> ed) Brailliar Tiridel Can. McMillian publishers.
47. Ullah F, Ayaz M, Sadiq A, Hussain A, Ahmad S, Imran M, Zeb A, 2016. Phenolic, flavonoid contents, anticholinesterase and antioxidant evaluation of *Iris germanica* var; florentina. *Nat Product Res* , 30(12) 1440-4.
48. Vabeiryureilai M, Lalrinzuali K, Rosangkima G, Jagetia GC, 2014. Qualitative phytochemical analysis and antioxidant activity of *Aporosa octandra* (Buch.-Ham. ex D. Don) Extracts. *Int J Pharmaceut Res*, 6(4): 69.
49. Verma RC, Gupta A, 2004. Effect of pre-treatments on quality of solar-dried Amla. *J Food Engineer*, 65:397-402.
50. Warriar RR, Joshi G, Arunkumar AN, 2019. DNA fingerprinting in industrially important medicinal trees. *Annals of Phytomedicine*. 8(1): 19-35.
51. Zhang YJ, Tanaka T, Iwamoto Y, Yang CR, Kouno I, 2000. Phyllaemblic acid, a novel highly oxygenated norbisabolane from the roots of *Phyllanthus Emblica*. *Tetrahedron Lett*, 41: 1781-1784.

**Table 1: Physical parameters of different *Emblica officinalis* Gaertn plant**

Location	Height (Ft.)	Girth (Inches)	Canopy (Ft.)	Fruit weight (5Fruits)(g)	Seed weight (5Seeds)(g)
Sihphir (Plant 1)	20	28	13	41.56	3.42
Tuirial (Plant 2)	32	26	10	28.56	2.57
Tanhiril (Plant 3)	16	20	08	35.23	3.96
Falkawn (Plant 4)	15	18	08	09.53	1.67

**Table 2: Moisture content of different samples of *Emblica officinalis* Gaertn leaves**

Plant	Sample weight (g)		Moisture content (g)	Moisture (%)
	Fresh	Dried		
Sihphir (Plant 1)	250	111.64	138.64	44.65
Tuirial (Plant 2)	250	120.92	129.08	51.60
Tanhiril (Plant 3)	250	119.35	130.65	52.26
Falkawn (Plant 4)	250	121.06	128.94	51.57

**Table 3: Qualitative phytochemical analysis of *Emblica officinalis* Gaertn leaves extract.**

Sample	Tannins	Phlobatannis	Saponin	Flavonoids	Steroid	Terpenoid	Glycosides	Alkaloids
Sihphir (Plant 1)	+	-	+	+	+	+	+	-
Tuirial (Plant 2)	+	-	+	+	+	+	+	-
Tanhiril (Plant 3)	+	-	+	+	+	+	+	-
Falkawn (Plant 4)	+	-	+	+	+	+	+	-

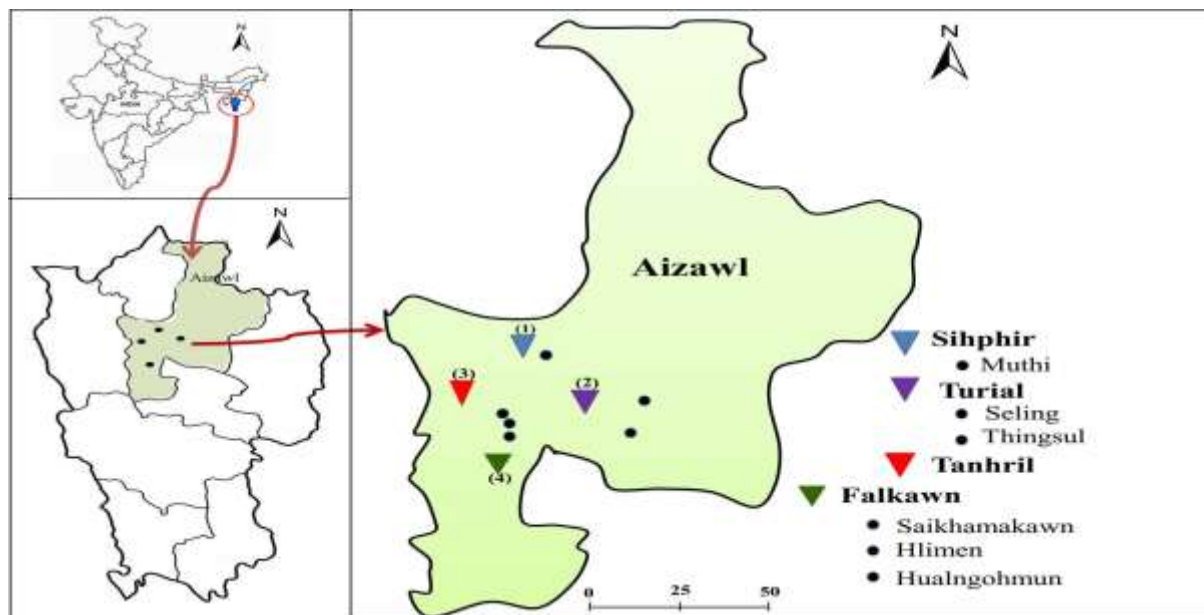
+ indicates presence of phytochemicals and

- indicates absence of phytochemicals.

**Table 4: Antimicrobial activity *Emblica officinalis* Gaertn leave extract**

Extracts	Antimicrobial activity against selected microbes		
	<i>S. aureus</i>	<i>P. aeruginosa</i>	<i>E. coli</i>
Sihphir (Plant 1)	19 mm	14 mm	Extract do not show any activity
Tuirial (Plant 2)	22 mm	16 mm	

Tanhril (Plant 3)	21 mm	13 mm
Falkawn (Plant 4)	No clear zone	No clear zone



 Sihphir  
 Turial  
 Tanhril  
 Falkawn  
 - ..

Figure 1: Black spot in the Map indicates sampling the location. There were 10 site or plants collected and catagorised into 4 different sites based on location. These four representative samples were further investigated.

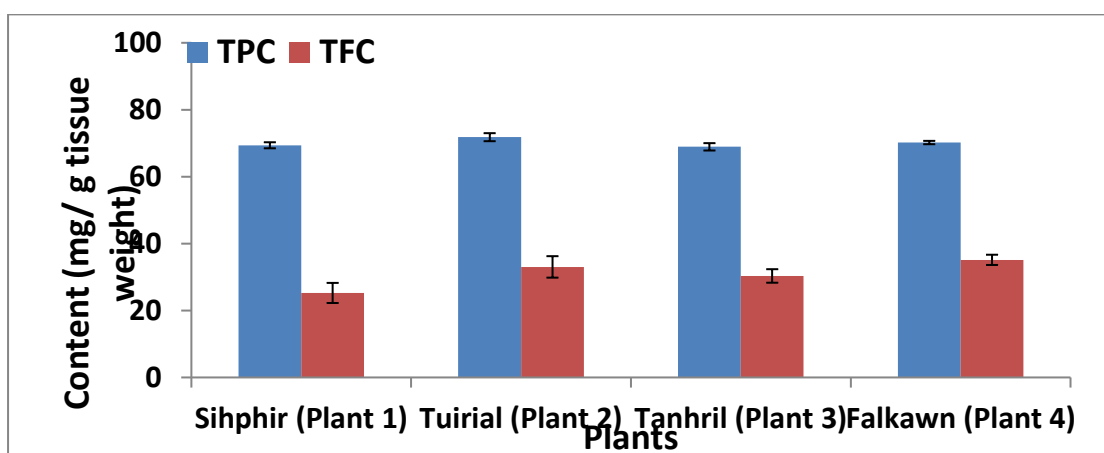


Figure 2: Total phenols and flavonoid content of *Aonla* leaves extracts. The results were expressed as Mean  $\pm$  SD, n=3.



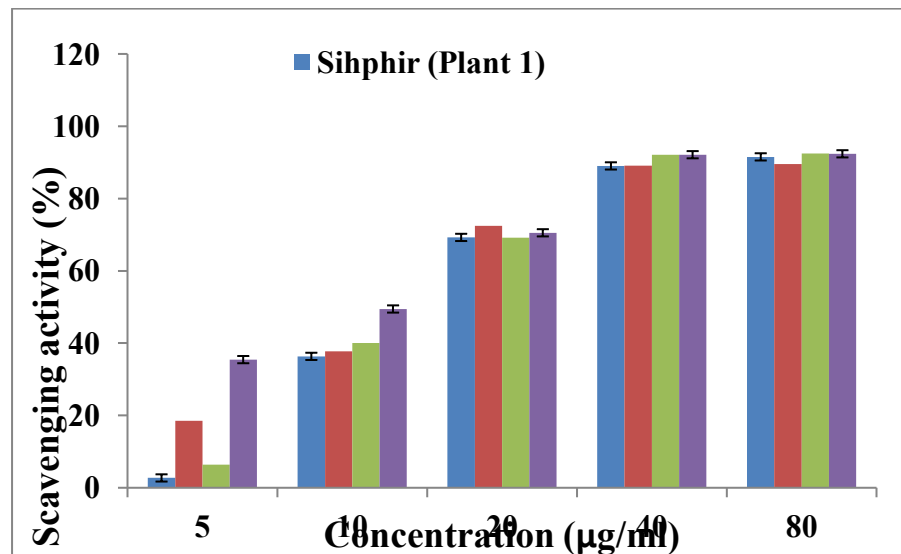


Figure 3: DPPH radical scavenging activity of *Aonla* leaves extracts. The results were expressed as Mean  $\pm$  SD, n=3.

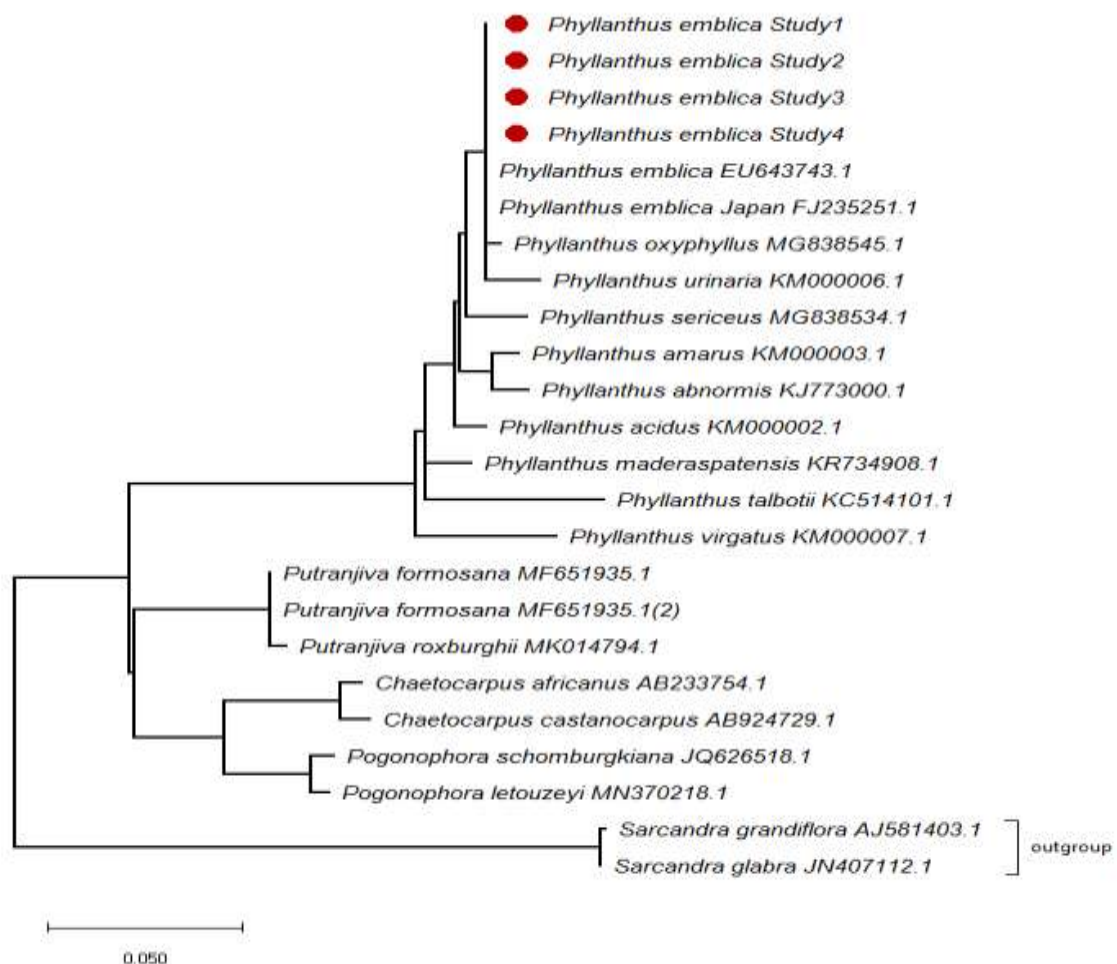


Figure 4: Molecular Phylogenetic analysis by Maximum Likelihood method