ISSN: 2229-7359 Vol. 11 No. 16s, 2025

https://www.theaspd.com/ijes.php

ACCEPTANCE LETTER - 3/7/25PUBLISHED PAPER - 2/7/25

Plant Responses To Drought Stress Under Climate Change And Recent Biotechnological Interventions For Mitigation: Mini Review

Soham Trivedi¹, Priyanka Patil², Pradeep Kumar Singh³, Meshak Dhanashekaran Cecileya Jasmin^{4,5}, Narayanswamy Radhakrishnan⁴, Krishna Rana⁵, Sujeet Kumar Mritunjay⁵, Shailaja Mohanta⁵, Sekhar Tiwari^{5*}

Email id: drsekhar.tiwari@rediffmail.com

ABSTRACT

Climate change has intensified drought stress, posing a major threat to global crop productivity. Plants respond through various adaptive mechanisms involving root architecture remodelling, altered leaf development, and modified reproductive timing. Recent biotechnological interventions offer promising strategies to enhance drought tolerance. CRISPR/Cas9 genome editing enables precise modification of key drought-responsive genes, improving traits such as root growth and water use efficiency. Omics-assisted breeding approaches, including marker-assisted selection and genomic selection, are accelerating the development of drought-resilient cultivars. Microbiome engineering using plant growth-promoting rhizobacteria (PGPR) enhances stress tolerance by regulating hormone levels, root growth, and antioxidant activity. Additionally, nanotechnology and biostimulants improve seed performance, water uptake, and physiological resilience under drought conditions. Despite these advances, challenges remain in translating laboratory successes to field applications. Integrating modern biotechnology with traditional breeding and sustainable agricultural practices is essential for ensuring crop productivity under increasing drought stress driven by climate change.

Kev words: Biostimulants; Climate change; Drought; Genome editing; Plants.

INTRODUCTION

Climate change is a complex phenomenon involving interactions among living organisms, the atmosphere, and water bodies such as land surfaces, snow, and oceans [1]. It is statistically defined as a change in the mean state of the climate, with respect to parameters like temperature, precipitation, and atmospheric pressure, over an extended period [2]. Climate change presents one of the most pressing global challenges, affecting food security, ecosystems, water resources, and human health. Since before the industrial era, the world's average temperature has increased by about 0.74°C, and carbon dioxide levels have risen from around 280 ppm to 400 ppm as of 2013 [3, 4]. These changes are already affecting farming, especially in developing countries where agriculture is a key source of livelihood. Extreme weather due to climate change has reduced crop yields, increasing malnutrition, poverty, and health problems. In India, warming is most noticeable in the northeast, central, and peninsular regions. For instance, in Punjab, a temperature increase of 1°C, 2°C, or 3°C may reduce grain yields by 5.4%, 7.4%, and 25.1% respectively [5]. Drought—a major abiotic stress-has threatened global crop productivity over the past 50 years [6]. Rising temperatures, changing precipitation, and elevated vapor pressure deficits contribute to water scarcity, intensifying drought-induced plant stress [7]. Under drought stress, especially as intensified by climate change, plants undergo developmental and morphological adaptations to maintain productivity. This paper presents an overview of plant responses to drought stress under climate change and highlights recent biotechnological interventions to mitigate drought effects.

1. Root developmental responses to drought stress

¹ Department of Biotechnology, Parul Institute of Technology, Parul University, Vadodara, Gujarat-391760

²Department of Life Sciences, Sigma University, Vadodara, Gujarat-390019

³ Rai School of Sciences, Rai University, Ahmedabad, Gujarat-382260

⁴Department of Biochemistry, Saveetha Institute of Medical & Technical Sciences, Tamil Nadu-602105

⁵School of Sciences, P P Savani University, Dhamdod, Dist Surat, Gujarat-394125

^{*}Corresponding Author: Sekhar Tiwari, PhD

^{*}School of Sciences, P P Savani University, Dhamdod, Dist-Surat, Gujarat-394125

ISSN: 2229-7359 Vol. 11 No. 16s, 2025

https://www.theaspd.com/ijes.php

Plants deploy several strategies to survive drought stress, including drought avoidance, desiccation prevention, and rapid recovery after rewetting [8, 9]. Root systems are central to drought avoidance since they enable water uptake. Dryland plants typically have deeper root systems than hydrophilic species. Root architectural remodelling (RAR), regulated by hormones like abscisic acid (ABA) and auxin, enhances water access [10]. The DEEPER ROOTING 1 (DRO1) gene improves drought tolerance by modifying root angles [11].

2. Leaf developmental responses to drought stress

Leaf growth, dependent on cell division and expansion, is highly sensitive to drought [12, 13]. In *Arabidopsis thaliana*, drought reduces leaf area and cell number [14]. In *Ricinus communis*, larger leaves recover better after drought than smaller ones [15]. Gene expression studies showed ABA signalling and cell wall modifications play key roles in leaf response to water deficit [14]. In *Nicotiana tabacum*, drought increases leaf thickness by altering cell structure and boosting expression of cell wall genes [16]. Expansins, enzymes that loosen cell walls, help maintain leaf expansion during drought, with genetically engineered plants showing greater tolerance [17].

3. Reproductive developmental responses to drought stress

Flowering time determines a plant's reproductive success during drought. Some plants escape drought by flowering early, while others delay flowering and optimize water use [18–20]. In *Arabidopsis thaliana*, *FRIGIDA* (*FRI*) delays flowering by activating *FLOWERING LOCUS C* (*FLC*) [21]. Natural variants of *FRI* affect drought response; early-flowering plants tend to escape drought, while late-flowering one's practice avoidance [22, 23].

4. Mitigation strategies

4.1 Crispr/Cas genome editing

DRO1 is a key gene regulating drought-induced root development [11]. CRISPR/Cas9 enables precise editing of drought-responsive genes. Editing *TaRPK1* in wheat improved root branching and yield under drought [24]. In soybean, knocking out *GmHdz4*, a negative regulator of lateral root development, enhanced drought tolerance [25]. Editing ARGOS8 in maize increased drought tolerance in the field [26]. In rice and tomato, CRISPR-edited genes (ERA1, SILBD40, SIMAPK3) improved water use and stomatal function [27, 28].

4.2. Omics-assisted selection and phenotyping

4.2.1 Genomic selection and marker-assisted breeding

Quantitative trait loci (QTLs) such as Stg1–Stg4 have been incorporated into elite sorghum lines for post-flowering drought resistance [29, 30]. In rice, QTLs qDTY1.1 and qDTY3.1 enhanced root depth and yield under drought [31]. In maize, genomic selection doubled genetic gains under drought [32]. Genomic selection captures small-effect loci, enabling accurate, rapid selection [33].

4.2.2 High-throughput phenotyping and AI

UAVs and multispectral imaging allow early drought detection and cultivar screening in crops like soybean [34]. AI tools also analyse multi-omics data to identify resilience markers [35].

4.2.3. Microbiome engineering and PGPR

PGPR such as *Bacillus*, *Pseudomonas*, and *Azospirillum* improve drought resilience through enhanced root growth, hormone regulation, and antioxidant activation [36, 37, 38]. These microbes produce exopolysaccharides (EPS), ABA, and IAA, supporting water retention and stress response [39, 40]. They also stimulate antioxidant enzymes like SOD, CAT, and POD [41].

4.2.4. Nanotechnology and Biostimulants

4.2.4.1 Seed nanopriming

Nanoparticles improve seed performance by forming pores in seed coats, triggering aquaporin expression and activating antioxidant pathways [42]. SiO₂ priming in wheat and ZnO priming in soybean/marigold enhanced drought tolerance [43–45]. Recent studies showed biogenic nanoparticles (e.g., Cu, Ni, Co oxides) boost biomass in maize under drought [46].

ISSN: 2229-7359 Vol. 11 No. 16s, 2025

https://www.theaspd.com/ijes.php

4.2.4.2 Biostimulants

Seaweed extracts, humic acids, and protein hydrolysates enhance drought resilience by promoting root growth, osmoregulation, and antioxidant defenses [47–49]. Ascophyllum nodosum and Sargassum extracts improved water uptake and photosynthesis under drought [50, 51]. Humic acids enhance soil water retention and root function [52]. Microbial biostimulants like mycorrhizae strengthen plant water relations [49]. Foliar/seed treatments with biostimulants also improve water use efficiency and stress tolerance [53, 54].

CHALLENGES AND PROSPECTS

Drought is a multidimensional common stress factor that negatively impacts plant growth, development, and metabolism. Thus, drought stress is a complex phenomenon adversely affecting plants at multiple levels. Plants have evolved a variety of tolerance strategies at molecular, developmental, physiological, morphological, and biochemical levels to cope with drought stress. These drought stress responses can be direct or indirect, and include regulation of water content, stomatal movements, leaf senescence, osmolytes adjustment, antioxidant metabolism, photosynthetic activity, and phytohormonal production. Different groups of beneficial bacteria that colonize plant roots promote plant growth and development, and stress tolerance through a variety of strategies. In their role as eco-friendly bio-fertilizers, PGPR can alleviate the detrimental effects of drought on plants by enhancing growth under water deficit through different processes. PGPR can promote plant growth and alleviate drought stress using the same strategies. Drought-stressed plants inoculated with PGPR display several adaptive responses to maintain water potential in the tissues such as osmotic adjustment, production of osmoprotectants and growth regulators, as well as increased antioxidant activity. The application of PGPR can be an effective tool to induce drought tolerance and sustain productivity in drought-stressed plants. Plant biotechnology plays a crucial role in this effort by providing tools such as genetic engineering and advanced breeding techniques to develop crop varieties that better tolerate climate-related stresses. These methods have shown success in maintaining productivity under harsh environmental conditions. However, progress in developing stress-resistant crops has been slow due to scientific and technical challenges. Overcoming these obstacles is vital to produce crops that can sustain in drought conditions and unpredictable climates.

ACKNOWLEDGEMENTS

All authors are thankful to their affiliated Universities

DECLARATIONS

Conflict of interest: The authors declared no conflict of interest with respect to the authorship, and publication of this review article.

REFERENCES

- 1. Lovejoy, T. E., & Hannah, L. (2005). Climate Change and Biodiversity. New Delhi: TERI Press.
- 2. Solomon, S., Qin, D., Manning, M., et al. (2007). Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the IPCC. Cambridge University Press.
- 3. Intergovernmental Panel on Climate Change. (2007). Climate Change 2007: The Physical Science Basis. Cambridge University Press.
- 4. National Oceanic and Atmospheric Administration. (n.d.). Climate change: Atmospheric carbon dioxide. Climate.gov.
- 5. Aggarwal, P. K., Singh, A. K., Samra, J. S., Singh, G., Gogoi, A. K., Rao, G. G. S. N., & Ramakrishna, Y. S. (2009). Introduction. In P. K. Aggarwal (Ed.), Global Climate Change and Indian Agriculture: Case studies from the ICAR network project (pp. 1–4). Indian Council of Agricultural Research.
- 6. Sánchez-Bermúdez, M., Del Pozo, J. C., & Pernas, M. (2022). Effects of combined abiotic stresses related to climate change on root growth in crops. Frontiers in Plant Science, 13, 918537.
- 7. Hsiao, J., Swann, A. L., & Kim, S. H. (2019). Maize yield under a changing climate: The hidden role of vapor pressure deficit. Agricultural and Forest Meteorology, 279, 107692.
- 8. Comas, L. H., Becker, S. R., Cruz, V. M. V., Byrne, P. F., & Dierig, D. A. (2013). Root traits contributing to plant productivity under drought. *Frontiers in Plant Science*, *4*, 442.
- 9. Gowda, V. R. P., Henry, A., Yamauchi, A., Shashidhar, H. E., & Serraj, R. (2011). Root biology and genetic improvement for drought avoidance in rice. *Field Crops Research*, 122, 1–13.
- 10. Vartanian, N. (1994). Drought rhizogenesis in Arabidopsis. Plant Physiology, 104(3), 761-767.

ISSN: 2229-7359 Vol. 11 No. 16s, 2025

https://www.theaspd.com/ijes.php

- 11. Feng, X., Jia, L., Cai, Y., Guan, H., Zheng, D., Zhang, W., ... Lu, Y. (2022). ABA-inducible DEEPER ROOTING 1 improves adaptation of maize to water deficiency. *Plant Biotechnology Journal*, 20(11), 2077–2088.
- 12. Aguirrezabal, L., Bouchier-Combaud, S., Radziejwoski, A., Dauzat, M., Cookson, S. J., & Granier, C. (2006). Plasticity to soil water deficit in *Arabidopsis thaliana*: dissection of leaf development into underlying growth dynamic and cellular variables reveals invisible phenotypes. *Plant*, Cell & Environment, 29(12), 2216–2227.
- 13. Baerenfaller, K., Massonnet, C., Walsh, S., Baginsky, S., Bühlmann, P., Hennig, L., ... Gruissem, W. (2012). Systems-based analysis of *Arabidopsis* leaf growth reveals adaptation to water deficit. *Molecular Systems Biology*, 8(1), 606.
- 14. Clauw, P., Coppens, F., De Beuf, K., Dhondt, S., Van Daele, T., Maleux, K., ... Inzé, D. (2015). Leaf responses to mild drought stress in natural variants of *Arabidopsis*. *Plant Physiology*, 167(3), 800–816.
- 15. Schurr, U., Heckenberger, U., Herdel, K., Walter, A., & Feil, R. (2000). Leaf development in *Ricinus communis* during drought stress: dynamics of growth processes, of cellular structure and of sink–source transition. *Journal of Experimental Botany*, 51(350), 1515–1529.
- 16. Khan, R., Ma, X., Hussain, Q., Chen, K., Farooq, S., Asim, M., ... Shi, Y. (2023). Transcriptome and anatomical studies reveal alterations in leaf thickness under long-term drought stress in tobacco. *Journal of Plant Physiology*, 281, 153920.
- 17. Samalova, M., Cosgrove, D. J., & Lee, Y. (2025). Differential expression and localization of expansins in *Arabidopsis* shoots: implications for cell wall dynamics and drought tolerance. *Frontiers in Plant Science*, 16, 1546819.
- 18. Sherrard, M. E., & Maherali, H. (2006). The adaptive significance of drought escape in *Avena barbata*, an annual grass. *Evolution*, 60(12), 2478–2489.
- 19. Heschel, M. S., & Riginos, C. (2005). Mechanisms of selection for drought stress tolerance and avoidance in *Impatiens* capensis (Balsaminaceae). American Journal of Botany, 92(1), 37–44.
- 20. Heschel, M. S., Donohue, K., Hausmann, N., & Schmitt, J. (2002). Population differentiation and natural selection for water-use efficiency in *Impatiens capensis* (Balsaminaceae). *International Journal of Plant Sciences*, 163(6), 907–912.
- 21. Searle, I., He, Y., Turck, F., Vincent, C., Fornara, F., Kröber, S., ... Coupland, G. (2006). The transcription factor FLC confers a flowering response to vernalization by repressing meristem competence and systemic signaling in *Arabidopsis*. Genes & Development, 20(7), 898–912.
- 22. Lovell, J. T., Juenger, T. E., Michaels, S. D., Lasky, J. R., Platt, A., Richards, J. H., ... McKay, J. K. (2013). Pleiotropy of FRIGIDA enhances the potential for multivariate adaptation. *Proceedings of the Royal Society B: Biological Sciences*, 280(1763), 20131043.
- 23. Schmalenbach, I., Dinneny, J. R., & Weigel, D. (2014). Flowering time variation in *Arabidopsis thaliana* under natural drought stress conditions. *Plant Physiology*, 164(1), 243–252.
- 24. Rahim, A. A., Uzair, M., Rehman, N., Fiaz, S., Attia, K. A., Abushady, A. M., ... Khan, M. R. (2024). CRISPR/Cas9 mediated *TaRPK1* root architecture gene mutagenesis confers enhanced wheat yield. *Journal of King Saud University*—Science, 36(2), 103063.
- 25. Zhong, X., Hong, W., Shu, Y., Li, J., Liu, L., Chen, X., ... Tang, G. (2022). CRISPR/Cas9 mediated gene-editing of *GmHdz4* transcription factor enhances drought tolerance in soybean (*Glycine max* [L.] Merr.). Frontiers in Plant Science, 13, 988505.
- 26. Shi, J., Habben, J. E., Archibald, R. L., Drummond, B. J., Chamberlin, M. A., Williams, R. W., ... Weers, B. P. (2015). Overexpression of ARGOS genes modifies plant sensitivity to ethylene, leading to improved drought tolerance in both *Arabidopsis* and maize. *Plant Physiology*, 169(1), 266–282.
- 27. Santosh Kumar, V. V., Verma, R. K., Yadav, S. K., Yadav, P., Watts, A., Rao, M. V., & Chinnusamy, V. (2020). CRISPR/Cas9 targeted mutagenesis of *SlLBD40*, a lateral organ boundaries domain transcription factor, enhances drought tolerance in tomato. *Plant Science*, 301, 110683.
- 28. Kumar, A., Sandhu, N., Dixit, S., Yadav, S., Swamy, B. P. M., & Shamsudin, N. A. A. (2018). Marker-assisted selection strategy to pyramid two or more QTLs for quantitative trait-grain yield under drought. *Rice*, 11, 1–16.
- 29. Kassahun, B., Bidinger, F. R., Hash, C. T., & Kuruvinashetti, M. S. (2010). Stay-green expression in early generation sorghum (Sorghum bicolor [L.] Moench) QTL introgression lines. Euphytica, 172, 351–362.
- 30. Faye, J. M., Akata, E. A., Sine, B., Diatta, C., Cisse, N., Fonceka, D., & Morris, G. P. (2022). Quantitative and population genomics suggest a broad role of stay-green loci in the drought adaptation of sorghum. *The Plant Genome*, 15(1), e20176.
- 31. Beyene, Y., Semagn, K., Mugo, S., Tarekegne, A., Babu, R., Meisel, B., ... Crossa, J. (2015). Genetic gains in grain yield through genomic selection in eight bi-parental maize populations under drought stress. *Crop Science*, *55*(1), 154–163.
- 32. Beyene, Y. (duplicate entry, same as 31 merge accordingly).
- 33. Crossa, J., Pérez-Rodríguez, P., Cuevas, J., Montesinos-López, O., Jarquín, D., et al. (2017). Genomic selection in plant breeding: Methods, models, and perspectives. *Trends in Plant Science*, 22(11), 961–975.

ISSN: 2229-7359 Vol. 11 No. 16s, 2025

https://www.theaspd.com/ijes.php

- 34. Jones, S. E., Ayanlade, T. T., Fallen, B., Jubery, T. Z., Singh, A., Ganapathysubramanian, B., ... Singh, A. K. (2024). Multi-sensor and multi-temporal high-throughput phenotyping for monitoring and early detection of water-limiting stress in soybean. *The Plant Phenome Journal*, 7(1), e70009.
- 35. Koh, E., Sunil, R. S., Lam, H. Y. I., & Mutwil, M. (2024). Harnessing Big Data and Artificial Intelligence to Study Plant Stress. *arXiv preprint arXiv:2404.15776*.
- 36. Vurukonda, S. S. K. P., Vardharajula, S., Shrivastava, M., & SkZ, A. (2016). Enhancement of drought stress tolerance in crops by plant growth promoting rhizobacteria. *Frontiers in Microbiology*, 7, 1221.
- 37. Ahmad, H. M., Fiaz, S., Hafeez, S., Zahra, S., Shah, A. N., Gul, B., ... Wang, X. (2022). Plant growth-promoting rhizobacteria eliminate the effect of drought stress in plants: A review. *Frontiers in Plant Science*, 13, 875774.
- 38. Chieb, M., & Gachomo, E. W. (2023). The role of plant growth promoting rhizobacteria in plant drought stress responses. BMC Plant Biology, 23(1), 407.
- 39. Naseem, H., Ahsan, M., Shahid, M. A., & Khan, N. (2018). Exopolysaccharides producing rhizobacteria and their role in plant growth and drought tolerance. *Journal of Basic Microbiology*, *58*(12), 1009–1022.
- 40. Ali, A., et al. (2020). Effects of ZnO nanoparticles on soybean seed germination and seedling growth under drought conditions. *Journal of Plant Nutrition*, 43(12), 1983–1992.
- 41. Sedghi, M., et al. (2013). ZnO nanoparticle treatment enhances germination and antioxidant activity in soybean under water stress. *Environmental and Experimental Botany*, 86, 76–83.
- 42. Nile, S. H., Thiruvengadam, M., Wang, Y., Samynathan, R., Shariati, M. A., Rebezov, M., ... Kai, G. (2022). Nano-priming as emerging seed priming technology for sustainable agriculture—recent developments and future perspectives. *Journal of Nanobiotechnology*, 20(1), 254.
- 43. Rai-Kalal, P., Tomar, R. S., & Jajoo, A. (2021). Seed nanopriming by silicon oxide improves drought stress alleviation potential in wheat plants. *Functional Plant Biology*, 48, 905–915.
- 44. Heidarieh, Z., Jafari, A., Ebrahimi, H. R., Haghighi, B. J., & Miri, H. R. (2024). Seed priming alleviates the adverse effects of drought stress on sesame genotypes by improving biochemical and physiological characteristics. *South African Journal of Botany*, 167, 256–269.
- 45. Calvo, P., Nelson, L., & Kloepper, J. W. (2014). Agricultural uses of plant biostimulants. Plant and Soil, 383, 3-41.
- 46. Canellas, L. P., Olivares, F. L., Okorokova-Façanha, A. L., & Façanha, A. R. (2002). Humic acids isolated from earthworm compost enhance root elongation, lateral root emergence, and plasma membrane H⁺-ATPase activity in maize roots. *Plant Physiology*, 130(4), 1951–1957.
- 47. Colla, G., Hoagland, L., Ruzzi, M., Cardarelli, M., Bonini, P., Canaguier, R., & Rouphael, Y. (2017). Biostimulant action of protein hydrolysates: Unraveling their effects on plant physiology and microbiome. *Frontiers in Plant Science*, *8*, 2202.
- 48. Rouphael, Y., & Colla, G. (2020). Biostimulants in agriculture. Frontiers in Plant Science, 11, 40.
- 49. Rouphael, Y., Colla, G., & Lucini, L. (2017). Influence of biostimulants on yield, quality, and nutraceutical properties of vegetables. *Frontiers in Plant Science*, 8, 1694.
- 50. Khan, W., Rayirath, U. P., Subramanian, S., Jithesh, M. N., Rayorath, P., Hodges, D. M., ... Prithiviraj, B. (2009). Seaweed extracts as biostimulants of plant growth and development. *Journal of Plant Growth Regulation*, 28(4), 386–399.
- 51. Sivasankari, S., Venkatesalu, V., Anantharaj, M., & Chandrasekaran, M. (2006). Effect of seaweed extracts on the growth and biochemical constituents of *Vigna sinensis*. *Bioresource Technology*, 97(14), 1745–1751.
- 52. Pizzeghello, D., Schiavon, M., Francioso, O., Dalla Vecchia, F., Ertani, A., & Nardi, S. (2020). Bioactivity of size-fractionated and unfractionated humic substances from two forest soils and comparative effects on N and S metabolism, nutrition, and root anatomy of *Allium sativum L. Frontiers in Plant Science*, 11, 1203.
- 53. Elanchezhiyan, K., Keerthana, U., & Prabakar, K. (2018). Effect of seaweed extract on drought stress tolerance in tomato (Solanum lycopersicum L.). Journal of Applied Phycology, 30, 279–289.
- 54. Crossa, J., Pérez-Rodríguez, P., Cuevas, J., Montesinos-López, O., Jarquín, D., et al. (2017). Genomic selection in plant breeding: Methods, models, and perspectives. *Trends in Plant Science*, 22(11), 961–975.