

Phytochemical Characterisation and Insecticidal Potential of *Erica arborea* L. Extracts Against *Sitophilus granarius* (L.): Towards an Eco-Friendly Pest Management Approach

Zerrouki Dehbia^{1*}, Darrab Djillali², Toumi Fawzia², Fendi Djamilia³, Ounas Malika³, Rouibah Asmaa³

¹Laboratory of Agro - Biotechnology and Nutrition in Semi-Arid Zones, Faculty of Nature and Life Sciences, University of Ibn Khaldoun, Tiaret, Algeria

²Laboratory for the Eco-Development of Spaces, Djillali Liabes University of Sidi Bel Abbes, Algeria

³University of Ibn Khaldoun, Tiaret, Algeria

*Corresponding author: dahbia.zerrouki@univ-tiaret.dz Zerrouki Dehbia);

Received: 03-01-2025; Accepted: 13-04-2025; Published: 11-10-2025

Abstract

This study assesses the phytochemical composition and insecticidal activity of a aqueous and ethanolic extract obtained from *Erica arborea* leaves collected in western Algeria. Extraction yields, total phenolic content, and insecticidal efficacy against *Sitophilus granarius* were determined.

The higher yield were obtained with thanolic maceration (22.18% in Oran; 20.18% in Saïda), whereas the lowest were recorded with Soxhlet extraction using hexane (~4–4.5%). The ethanolic extract exhibited a maximum mortality rate of 69% at $32 \mu\text{l}\cdot\text{ml}^{-1}$ after 72 hours. These findings confirm the potential of *Erica arborea*, as a natural source of biopesticidal compounds suitable for sustainable pest control.

Keywords: *Erica arborea*, plant extracts, phenolic compounds, *Sitophilus granarius*, biopesticide, Algeria.

1. INTRODUCTION

The excessive and uncontrolled use of synthetic pesticides in modern agricultural systems, has for several decades, led to serious ecotoxicological and public-health consequences. These chemical substances persist in the environment, accumulate in food chains, and disrupt natural ecological balances. Moreover, many insect pest species have developed increasing resistance to these molecules, which considerably reduces their long-term effectiveness (Mann, 1987).

In this context, the search for ecological and sustainable alternatives to chemical insecticides has become a major priority for crop protection and human health preservation. Aromatic and medicinal plants, owing to their secondary metabolites such as polyphenols, flavonoids, terpenoids, and alkaloids, represent a promising source of bioactive substances with biopesticidal potential. These natural compounds exhibit a wide range of biological effects—antioxidant, antibacterial, antifungal, and insecticidal—often with high biodegradability and low toxicity towards non-target organisms. Accordingly, the present study is part of a broader approach to valorise Algerian plant resources through the evaluation of the insecticidal potential of *Erica arborea* extracts collected from two regions of western Algeria (Oran and Saïda). Specifically, it aims to:

- Compare extraction yields using different techniques (maceration, ultrasound, Soxhlet).
- Quantify total phenolic compound in the extracts.
- Evaluate the insecticidal activity of these extracts against *S. granarius*.

This study therefore contributes to a better understanding of the biopesticidal potential of *Erica arborea* and promotes an integrated pest management approach that is environmentally friendly and adapted to local agroecological conditions.

2. MATERIALS AND METHODS

2.1. Study Species and Biological Model

Erica arborea L., commonly known as “tree heath”, occupies a special place within Mediterranean forest ecosystems. This woody species, belonging to the Ericaceae family, is widely distributed across the Mediterranean Basin, particularly in North Africa, southern Europe, and some Atlantic regions. In Algeria, it grows mainly in the mountainous areas of the North and West, where it contributes to soil stability, erosion control, and the regeneration of degraded maquis formations (Aït Youcef, 2006).

Sitophilus granarius (L.), commonly known as the granary weevil, is one of the major pests of stored grains

worldwide. It causes severe economic losses in both quantity and quality by damaging cerea stock and reducing their nutritional value. The use of ynthetic chemicals to control this pest often results in food contamination and environmental pollution, highlighting the importance of testing local plant extracts as alternative solutions.

2.2. Study Area

Sample of Erica arboreaL. Were collected from two representative regions of western Algeria: **Saïda** and **Oran**, characterised by contrasting bioclimatic conditions.

• Saïda Region:

Located at 34°50' N and 0°10' E, Saïda lies within a semi-arid bioclimatic stage with cold winters. Annual rain fall ranges between 300and450mm,with an average temperature of approximately 16 °C. The vegetation is mainly composed of maquis dominated by holm oak (*Quercus ilex*), Aleppo pine (*Pinus halepensis*), and heaths (*Erica arborea*,*E. multiflora*).

• Oran Region:

Situated at 35°42' N and -0°38' E, Oran experiences a milder Mediterranean climate with higher humidity. Average annual rain fall variesbetween450and 600 mm, and mean annual temperature is around 18 °C. The substrate is primarily calcareous, and the natural vegetation consists of typical coastal maquis formations with shrubby and herbaceous species.

2.3. Preparation of Plant Material

Leave ofE. arboreawere collected from healthy plants identified in the field during the flowering period (March–April 2024). Samples were placed in paper bags and transported to the laboratory for drying and processing.

The leaves were carefully washed with distilled water to remove impurities, then air- dried in the shade at ambient temperature (20–25 °C) for 10 days (Salah Dine&al., 2019).Once completely dry, the plant material was ground using an electric grinder to obtaina fine, homogeneous powder, which from light and moisture until extraction.

2.4. Extraction Techniques

Four extraction method swere compared to assess their efficiency:

1. Ethanolic Maceration(MAE):

Twentygramsof powderedleaveswereimmersedin200mlof80%ethanol for 72 hours with intermittent manual stirring. The filtrate was then concentrated under reduced pressure at 40 °C using a rotary evaporator.

2. Ethanolic Ultra sonic Extraction(UE):

Twentygramsof powderedmaterialweremixedwith200mlof80%ethanol and subjected to sonication (40 kHz) for 30 minutes at room temperature (Zengin &al., 2019).

3. Aqueous Ultra sonic Extraction(UA):

The same protocol was applied asi the UE method, butd is tilled water was used as solvent (Zengin&al., 2019, modified).

4. Soxhlet Hexane Extraction(SOH):

Twenty grams of powdered leaves were extract dusing200mlofhexanefor 6 hours in a Soxhlet apparatus (Zengin &al., 2019).

The obtained extract were filtered, concentrated, andair- driedat room temperature. Extraction yields were calculated according to the following formula:

$$R(\%) = \frac{M(\text{ext})}{M(\text{sch})} \times 100$$

$M(\text{sch})$

- M Text=mass of the xtract after solven te vaporation (g)
- M sample=mass of the plant material used (g)(Benyamina,2019)

2.5. Determination of Phenolic Compounds

2.5.1. Total Polyphenols

ThetotalpolyphenolcontentwasdeterminedusingtheFolin–Ciocalteumethod.A stock solution(1 mg/ml)of each extract was prepared. Then, 0.1 ml ofthis solution was mixed with 0.5 ml of Folin–Ciocalteu reagent and 1.5 ml of 7.5% Na_2CO_3 . After incubation for 30minutesat 25°C, absorbanc was measuredat765nmusingaUV-

Visspectro photo meter (UV-Vis1800,Shimadzu) (Feknous&al.,2014). Results were express eda smillig rams

of gallic acid equivalent per gram of dry extract (mg GAE/g).

2.5.2. Total Flavonoids

The quantitative estimation of total flavonoids in the leaf extracts was performed using a colorimetric method. Two millilitres of ethanolic $AlCl_3$ solution (2%) were added to 2 ml of each extract. After a 10-minute incubation period, absorbance was measured at 430 nm (Guendouze- Boucheffa & al., 2015).

The concentration of flavonoids was calculated using a calibration curve established with quercetin, and results were expressed as mg QE/g of extract (Heinrich & al., 2017).

2.5.3. Condensed Tannins

One millilitre of each extract was added to 2 ml of 1% vanillin solution and thoroughly mixed. The mixture was left to react at room temperature for 20 minutes, after which the absorbance was read at 500 nm against a blank. The concentration of condensed tannins was determined using a catechol calibration curve, and results were expressed as mg CE/g of extract (Benyamina, 2019).

2.6. Insecticidal Bioassays

2.6.1. Target Insect

Adults of *Sitophilus granarius* (L.) were obtained from a laboratory culture reared on durum wheat (*Triticum durum*). The insects used in the experiments were 7–14 days old.

2.6.2. Contact Toxicity on Filter Paper

The insecticidal activity of the plant extracts was evaluated using the filter paper contact test (Seri-Kouassi & al., 2004).

Groups of ten adult weevils were placed in Petri dishes containing wheat grain treated with different extract concentrations (8, 16, 24, and 32 μ l/ml).

Mortality observations were made after 24, 48, and 72 hours (Salah Dine et al., 2019). Untreated controls were included for each series, and all tests were conducted in triplicate.

2.6.3. Mortality Rate

Mortality was the primary criterion for assessing the effectiveness of the treatments. The observed mortality percentage in both control and treated groups was calculated using the formula described by Ould El Hadj & al. (2006):

$$\text{Observed mortality} = \frac{\text{Number of deaths}}{\text{Total number of individuals}} \times 100$$

Total number of individuals

2.6.3.1. Calculation of the Mortality Rate

The mortality rate was calculated according to Abbott's formula (1925):

$$MC(\%) = \frac{(M_2 - M_1)}{(100 - M_1)} \times 100$$

where:

- MC = corrected mortality (%)
- M₂ = mortality in the treated population (%)
- M₁ = mortality in the control population (%)

This correction ensures that the effect of natural mortality or handling stress in the control batches is excluded from the final analysis of insecticidal activity.

2.7. Statistical Analysis

Data were analysed using SPSS software (version 22.0). A one-way analysis of variance (ANOVA) was used to compare means, followed by Tukey's HSD test to determine significant differences ($p < 0.05$). Pearson's correlation coefficients were calculated to evaluate relationships between concentration, exposure time, and mortality rate, thus assessing the dose-response relationship.

3. RESULTS AND DISCUSSION

3.1. General Appearance and Colour of Extracts

The extracts obtained from *Erica arborea* leaves exhibited distinct physical characteristics depending on the extraction method and solvent used. (table N1)

Table 1. Physical appearance and colour of the four extracts obtained from *Erica borealis* leaves

Extraction Method

Appearance	Colour	Colour	Appearance
(Oran)	(Saïda)	(Oran)	(Saïda)
Bottlegreen	Lightgreen	Powdery	Powdery
Bottlegreen	Lightgreen	Pastelike	Pastelike
Brightbrown	Brightbrown	Powdery	Shiny
green	green	Mattpowder	Mattpowder

Ethanolic maceration (MAE)

Ethanolultra sound (UE) Aqueous ultrasound (UA)Hexane Soxhlet (SOH)

The variation in colour and texture between extracts reflects theinfluence of solvent polarityon the chemical profile of the extracts. Ethanol-based extracts are richer in oxidised phenolic compounds (hence darker tones), while aqueous extracts appear clearer due to hydrosoluble tannins. Hexane extracts are pale and oily, dominated by lipidic or terpenoid compounds.

These findings are consistent with previous observations reported by Yaici&al.(2019) for E. arborea and by Bessah & Benyoussef (2014) for other Mediterranean aromatic plants.

3.2. Extraction Yields

Extraction yields varied between 4.05% and 22.18%, depending on the extraction technique and the plant's geographical origin (Table 2, Figure N1 and Figure N2)

Table 2. Extraction yields of different Erica arborea leaf extracts

Mass (g)	Mass (g)	Yield (%)	Yield (%)
Oran	Saïda	Oran	Saïda
1.109	1.009	22.18	20.18
1.65	1.01	16.50	10.10
1.09	1.11	10.00	11.10
0.909	0.81	4.55	4.05

Extraction Method

Ethanolic maceration (MAE)

Ethanolic ultrasound (UE) Aqueous ultrasound (UA)Hexane Soxhlet (SOH)

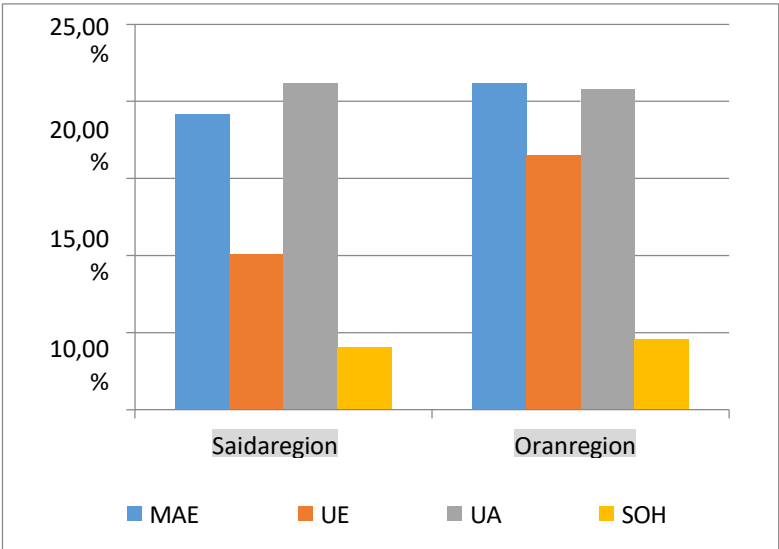


Figure 1. Extraction yields of Erica arborea leaf extracts according to geographical origin (Saïda and Oran)

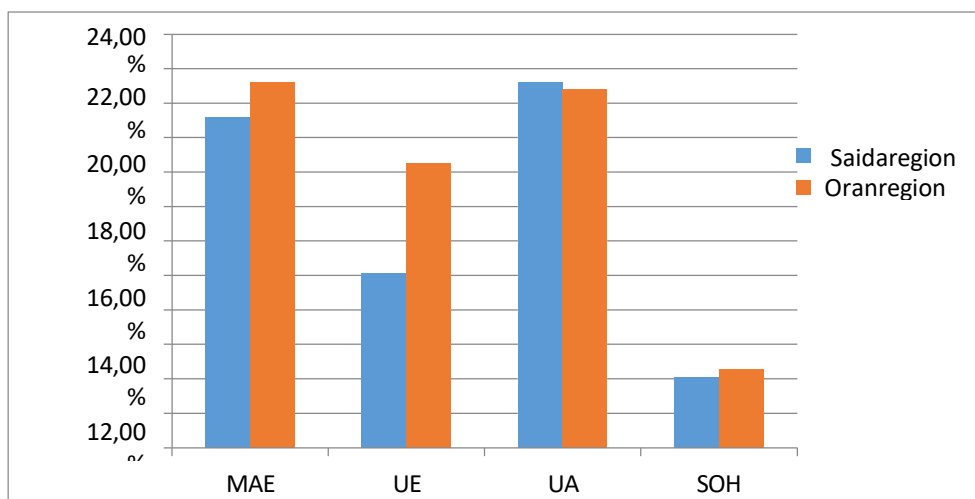


Figure2. Comparison of extract ion yields according to extract on method (MAE,UE,UA,SOH).

The **ethanolic maceration (MAE)** method produced the high yields, followed by **ethanolic ultrasonic extraction (UE)**. This can be attributed to the intermediate polarity of ethanol, which is capable of extracting both polar and slightly non-polar metabolites, such as polyphenols and flavonoids. Aqueous extraction (UA) produced intermediate yields, while Soxhlet extraction using hexane (SOH) gave the lowest results, as this non-polar solvent primarily dissolves lipids and waxes.

Comparatively, Yaici & al. (2019), working on aqueous extracts of *E. arborea* leaves from the Sétif region, reported similar though slightly lower yields (14.74%). Amroun & al. (2021), who studied ethanolic and aqueous extracts from the Tizi Ouzou region, obtained comparable values (17.17% for aqueous and 16.63% for ethanolic extractions), confirming the influence of both method and origin on extraction efficiency.

3.3. Quantitative Analysis of Phytochemicals

3.3.1. Total Phenolic Content

From the calibration curve constructed with gallic acid (Figure3),

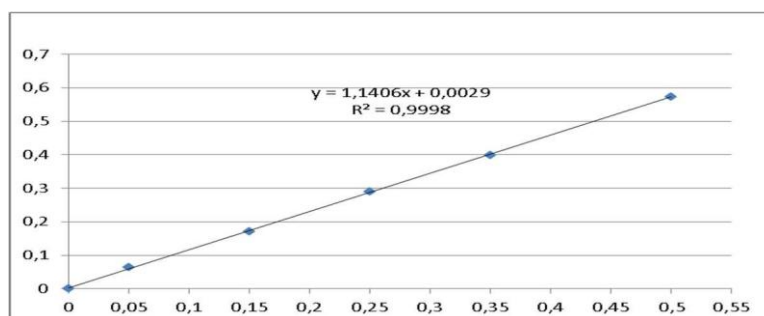


Figure 3. Calibration curve used for the determination of total phenolic compounds (gallic acid standard).

total phenolic content ranged from 60.66 ± 1.21 to 5.5 ± 0.24 mg GAE/g dry extract in Saïda samples, and from 51.7 to 13.9 mg GAE/g in Oran samples.

The highest concentration was recorded in Saïda using aqueous ultrasonic extraction (60.66 mg GAE/g), while the lowest was obtained using hexane Soxhlet extraction (5.5 mg GAE/g).

Among all methods, **ethanolic maceration** and **aqueous ultrasound** produced the richest extracts in both regions (52.95 ± 2.32 mg GAE/g in Saïda and 51.7 ± 1.95 mg GAE/g in Oran). (Figure N4 and Figure N5)

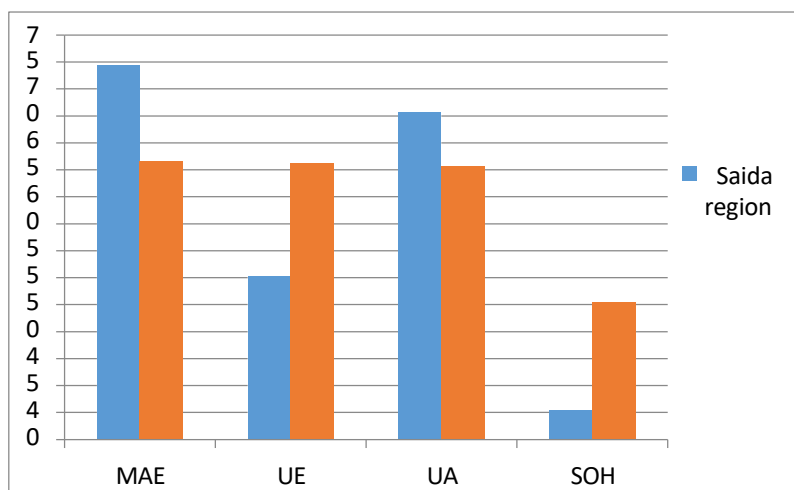


Figure4. Comparison of total polyphenol concentrations in different *Erica arborea* leaf extracts.

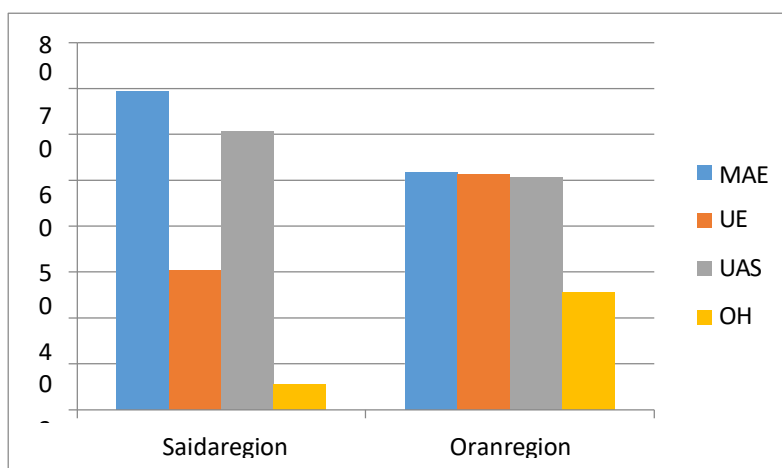


Figure5. Comparison of total phenolic contents between the two study sites (Saida and Oran).

These differences indicate that extraction efficiency strongly depends on solvent polarity and technique. When compared with other Algerian studies, our results are slightly lower than those reported by Yaici & al. (2019) in Sétif (74 mg GAE/g), Guendouze & al. (2015) in Béjaïa (70.8 mg GAE/g), Amroun & al. (2021) in Tizi Ouzou (139 mg GAE/g), and Belarbi & al. (2018) in Tlemcen (168.23 mg GAE/g). Globally, however, these values remain within the range observed for Mediterranean *E. arborea* populations. For example, reported 75.83 mg GAE/g in Turkish samples by Ay & al. (2007) and 78.49 mg GAE/g in Turkish sample Amezouar & al. (2013).

3.3.2. Total Flavonoids

Using a calibration curve established with quercitrin (Figure 6), total flavonoid content varied between 17.86 mg QE/g and 10.9 mg QE/g in Saida, and from 9.62 mg QE/g to 24.81 mg QE/g in Oran. The results are presented in figure 7.

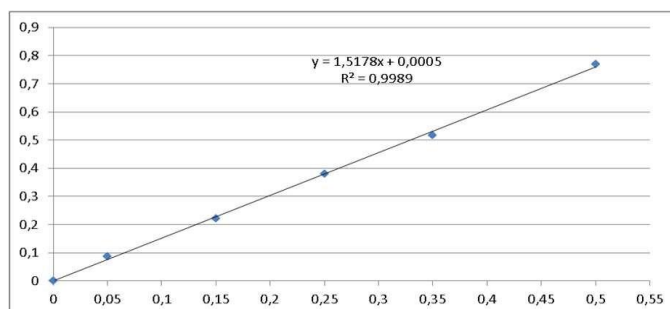


Figure6. Calibration curve used for the quantification of total flavonoids (quercitrin standard).

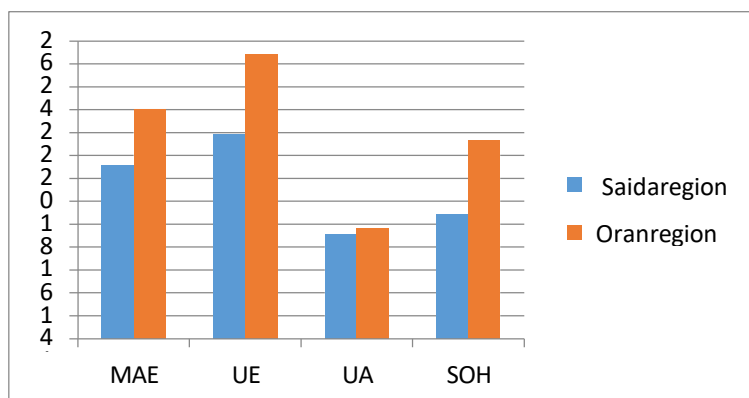


Figure7. Comparison of total flavonoid concentrations in the different *Erica* boreale extracts. The **ethanolic ultrasonic extraction (UE)** method yielded the richest flavonoid contents for both regions, followed by maceration. In general, the Oran extract contained slightly higher flavonoid levels than those from Saïda, possibly reflecting local climatic and soil influences.

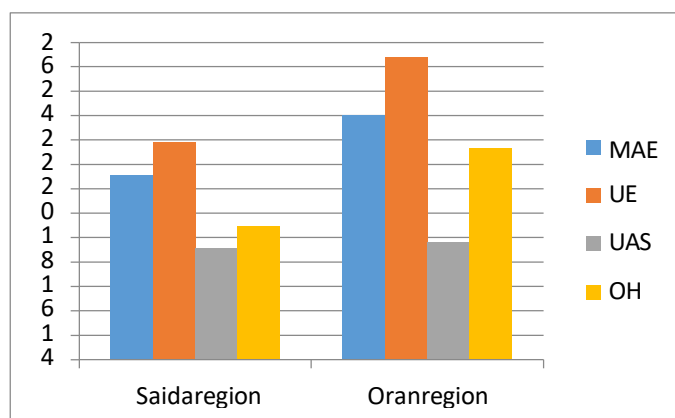


Figure8. Comparison of total flavonoid contents between Saïda and Oran samples.

3.3.3. Condensed Tannins

Based on the catechin calibration curve (Figure9),

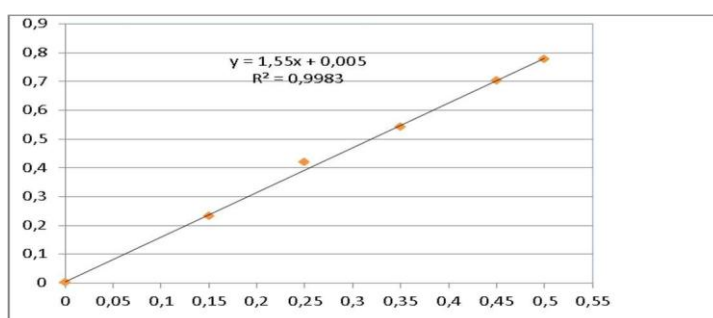


Figure9. Calibration curve used for the determination of condensed tannins (catechin standard).

The tannin content was relatively low across all extraction methods and study sites, ranging from **5.17 to 7.96 mg CE/g extract**. This variation may be related to environmental conditions, collection period, genetic factors, and extraction parameters. (Figure 10 and 11).

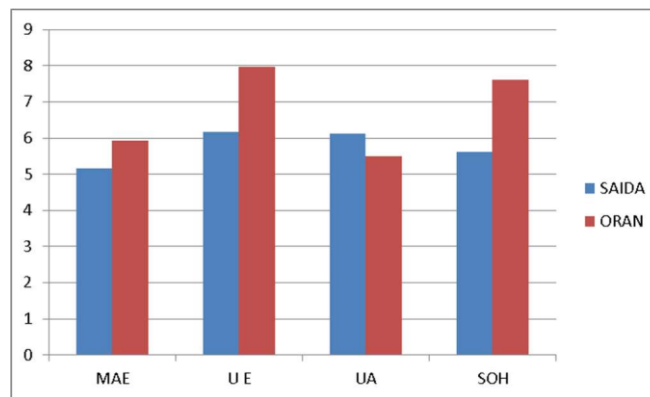


Figure10. Comparison of condensed tannin concentration sinthe different E. arborea leaf extracts.

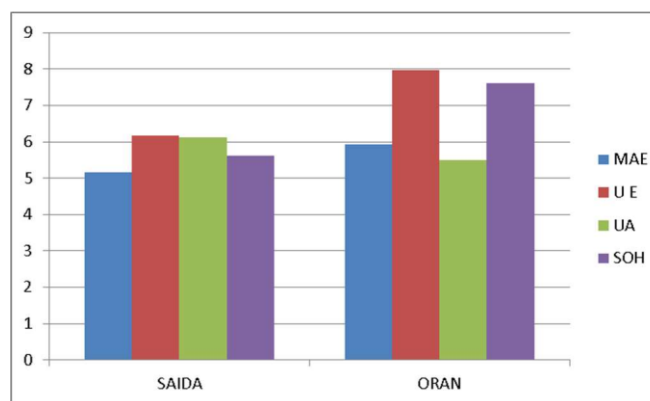


Figure11. ComparisonofcondensedtannincontentsbetweenSaïdaandOranregions.

According to Richter (1993), the content and chemical composition of secondary meta bolite scanvary widely due to both on trinsic (genetic) and extrinsic (environmental and methodological) factors.

3.4. Insecticidal Activity of *Erica*ar borea

3.4.1. Mortality Rate and Dose–Response Relationship

Because some mortality occurredin control groups, Abbott’s (1925) formula was used to correct mortality data. The mortality rates of *S. granariu* adults exposed to different concentrations of arborea extracts are shown in Figure 12. A clear variation was observed among treatments depending on extract type, concentration, and exposure duration. The **ethanolic maceration extract** produced the **highest corrected mortality (69%)**at32µl/mlafter72hours.



Figure12.PhotographicrepresentationshowingtheeffectsofdifferentconcentrationsofE. arborea leaf extracts on *Sitophilus granarius* adults.

As shown in Figures 13 and 14, mortality increased linearly with both concentration and exposure time, indicating **dose- dependent toxic effect** of E.arborea extracts on *S. granarius*.

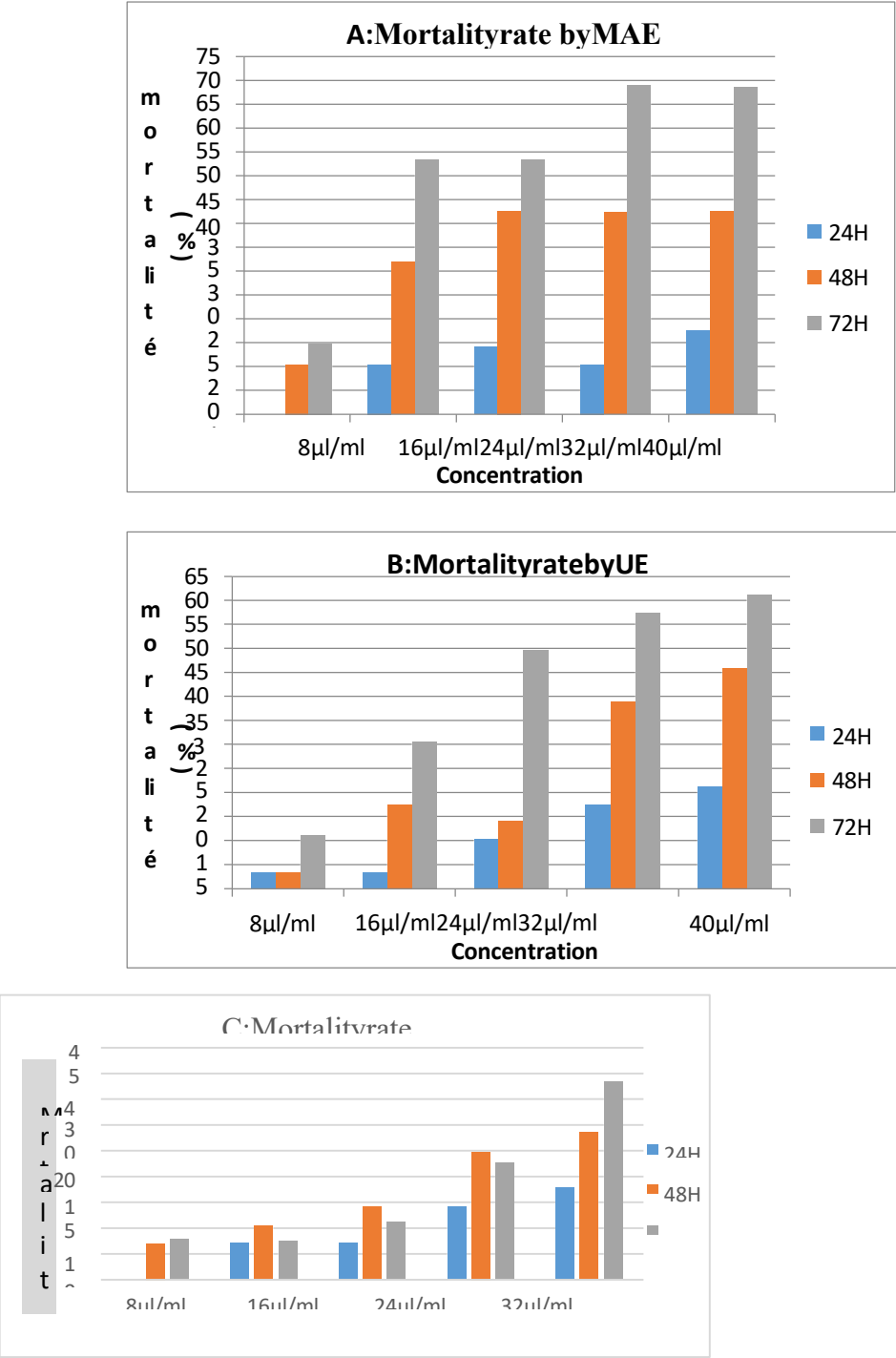


Figure13. (A,B,C): Mortality rates of *Sitophilus granarius* adults exposed to various *E. arborea* leaf extracts.

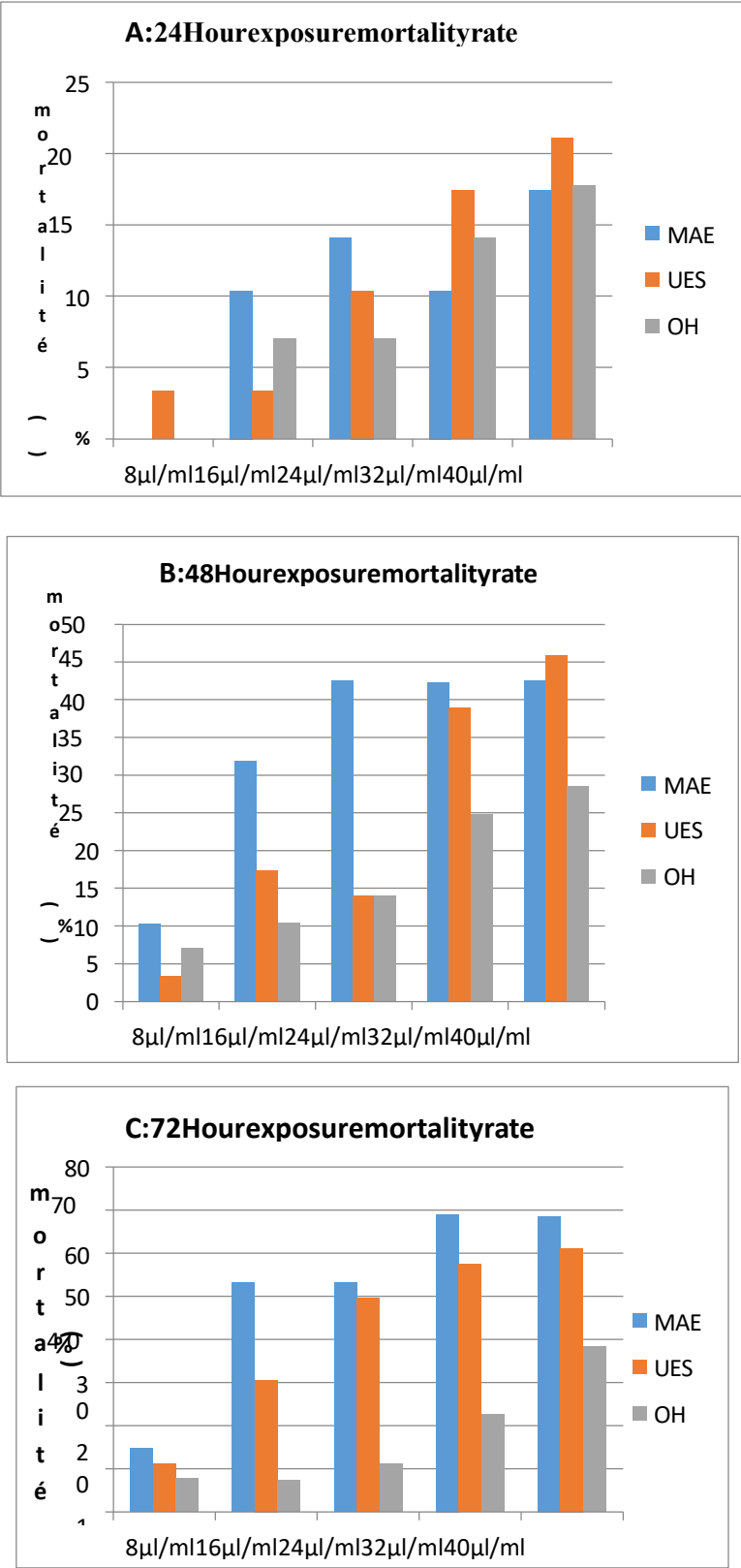


Figure14. (A,B,C): Time-course variation in the mortality rate of *Sitophilus granaries* after exposure to *E. arborea* leaf extracts.

4. GENERAL DISCUSSION

The overall result obtained in this study highlight both the **chemical richness** and the **biological potential** of *Erica arborea* extracts collected from the Saïda and Oran regions in western Algeria. Although often regarded as secondary species in Mediterranean agroecosystems, *E. arborea* demonstrates here a **significant ecological and biotechnological value**.

The **differences in extraction yield** observed among the various methods reflect the diversity of chemical compounds present in the plant and their variable affinities for the solvents used. Ethanolic extracts proved to be the most effective, both in terms of yield and total phenolic content, confirming that **polar solvents** promote the extraction of phenolic and flavonoid metabolites. These results are consistent with the findings of Guendouze et al. (2015) and Yaici et al. (2019), who reported a strong correlation between solvent polarity and the concentration of phenolic compounds in Ericaceae species. From a biological perspective, the **insecticidal activity** observed against *Sitophilus granarius* was significant and dependent on both concentration and exposure duration. The maximum mortality rate (69% after 72 hours at 32 µl/ml) demonstrates the **progressive toxicity** of *E. arborea* ethanolic extracts. This effect is likely due to the **combined action** of polyphenols, tannins, and flavonoids, which are known to damage cellular membranes, inhibit digestive enzymes, and disrupt the energy metabolism of insects. Similar mechanisms have been reported by Bessah and Benyoussef (2014) in *Rosmarinus officinalis* and *Thymus vulgaris*, two Mediterranean aromatic species also rich in phenolic metabolites.

The **geographical variations** observed between the Saïda and Oran samples could be linked to **local ecological factors** such as soil type, humidity, light intensity, and temperature, all of which influence the biosynthesis of secondary metabolites. Such **eco-chemical variations** are commonly observed in aromatic and medicinal plants, where environmental conditions play a key role in both the quality and quantity of bioactive compounds produced (Aït Youcef, 2006).

From an applied perspective, the results of this study open the way for the **valorisation of *Erica arborea*** as a potential source of **natural bio pesticides**. The use of plant-based extracts could help reduce dependence on synthetic agrochemicals, protect the environment, and contribute to the preservation of local biodiversity.

Furthermore, this approach is fully consistent with the objectives of **sustainable development and agroecology**, promoting the rational use of native plant resources.

It should also be noted that the **chemical composition of *E. arborea* extracts** may vary depending on the season, the plant part used, and the extraction conditions. Further chromatographic analyses (e.g. HPLC, GC-MS) are required to **identify and quantify** the active molecules responsible for the insecticidal effect. Future research should also assess the **toxicity of these extracts on non-target organisms** to ensure their environmental safety before any practical application.

5. CONCLUSION

The study conducted on *Erica arborea* L. demonstrated its **high phytochemical richness** and **promising insecticidal potential** against *Sitophilus granarius*, a major pest of stored cereals.

Extraction yields and total phenolic contents varied depending on both the extraction method and the geographical origin of the samples, with **ethanolic and aqueous extracts obtained by maceration and ultrasonic techniques** showing the highest concentrations of bioactive compounds. These findings confirm that **solvent polarity** plays a key role in the extraction efficiency of secondary metabolites. The bioassays revealed a **significant dose-response relationship**, with a **maximum mortality rate of 69%** observed at 32 µl/ml after 72 hours of exposure. This insecticidal efficacy, mainly attributed to the presence of **polyphenols and flavonoids**, highlights the protective role of these natural compounds against grain pests.

The variability observed between samples from Saïda and Oran underlines the **influence of ecological and bioclimatic conditions** on the chemical composition and biological activity of plant extracts. Therefore, *Erica arborea* presents a **valuable local plant resource** that could be exploited for the development of **eco-friendly biopesticides** adapted to the agroecological systems of north-western Algeria. In conclusion, *Erica arborea* can be considered a **species with high biotechnological potential**, capable of contributing to the transition towards a **greener, safer, and more sustainable agriculture**.

REFERENCES

- Abbott, W.S. (1925). A method of computing the effectiveness of an insecticide. *Journal of Economic Entomology*, **18**, 265–267.
- Aït Youcef, M. (2006). *Medicinal Plants of Kabylia*. Ibis Press, Paris.
- Amroun, D., Hamoudi, M., Khennouf, S., Boutefnouchet, S., Harzallah, D., Amrane, M. & Dahamna, S. (2021). In-vivo anti-inflammatory activity and safety assessment of the aqueous extract of Algerian *Erica arborea* L. (Ericaceae) aerial parts. *Journal of Ethnopharmacology*, **271**, 113881.
- Belarbi, K. (2018). Biological activities of extracts from a medicinal plant “*Teucrium pseudoscorodonia*” collected from the Tlemcen region. PhD Thesis, University of Tlemcen–Abou Bekr Belkaid, Algeria.
- Benyamina, A. (2019). Ethnopharmacological study and characterisation of phenolic compounds of aerial organs of *Tetralina articulata* (Vahl) Masters from western Algeria. PhD Thesis.

12. Bessah, F.&Benyoussef, L. (2014). Antioxidant potential and insecticidal activity of extracts from Mediterranean aromatic plants. *Algerian Journal of Phytotherapy*, 2(1), 45–56.
13. Feknous, S. Saidi, F.&Mohamed Said, R. (2014). Extraction, characterisation and identification of some active secondary metabolites from lemon balm (*Melissa officinalis* L.). *Nature and Technology Journal*, 6(2), 7–13.
14. Guendouze, A Boudjelal, A.&Djeddi, S. (2015). Phytochemical study and biological activity of *Erica arborea* from northern Algeria. *Phytotherapy Journal*, 13(4), 201–208.
15. Heinrich, M., Barnes, J., Prieto-Garcia, J., Gibbons, S.& Williamson, E.M. (2017). *Fundamentals of Pharmacognosy and Phytotherapy*. Elsevier Health Sciences, London.
16. Mann, J. (1987). *Secondary Metabolism*. Clarendon Press, Oxford.
17. Ould El Hadj, M.D., Tankari Dan-Badjo, A., Halouane, F.&Doumandji, S. (2006). Comparative toxicity of extract from three acrid ifuge plant on fifth-instar larvae and adults of *Schistocerca gregaria* (Forskål, 1775) (Orthoptera- Cyrtacanthacridinae). *Sécheresse*, 17(3), 407–414.
18. Salah Dine, R.M., Abdelrahamene, L., Mohamed, T.&Tahar, M. (2019). Contact toxicity and repellency evaluation of *Rosmarinus eriocalyx* leaves essential oil (Lamiaceae) against adults of *Sitophilus granarius* (Coleoptera) and its anti-free radical DPP H activity. *South Asian Journal of Experimental Biology*, 9(4), 148–156.
19. Seri-Kouassi, B.P., Kanko, C., Aboua, L.R.N., Bekon, K.A., Glitho, A.I., Koukoua,
20. G.&N' Guessan, Y.T. (2004). Effects of essential oils from two Ivorian aromatic plants on *Callosobruchus maculatus* F. infesting cowpeas. *Comptes Rendus Chimie*, 7(10–11), 1043–1046.
21. Yaici, K., Belyagoubi, L. & Boumendjel, M. (2019). Evaluation of phenolic content and antioxidant activity in *Erica arborea* L. extracts. *Phytotherapy*, 17, 82–91.
22. Zengin, G., Cvetanović, A., Gašić, U., Stupar, A., Bulut, G., Senkardes, I., Dogan, A., Sebaluck-Sandoram, R., Rengasamy, K.R.R., Sinan, K.I.&Mahomoodally, M.F. (2019). Chemical composition and bio-functional perspectives of *Erica arborea* L. extracts obtained by different extraction techniques: Innovative insights. *Industrial Crops and Products*, 142, 111843.