ISSN: 2229-7359 Vol. 11 No. 25s,2025

https://theaspd.com/index.php

Accuracy Of Mixed Antibody Detection Of Canine Parvovirus And Canine Distemper Infection In Dogs

Amira. S. H. Hassenin^{1*}, Aneela Durrani²

- ¹ Department of Animal Medicine, Faculty of Veterinary Medicine, Zagazig University, Egypt. Email: amm_microhass@yahoo.com
- ² Department of Veterinary Clinical Sciences, College of Veterinary Medicine, Lahore, Pakistan. Email: amirasaad@zu.edu.eg

Abstract: Wide spread infections in canine are caused by parvovirus (CPV) and distemper virus (CDV) in most countries. Their detection for specific isolates infection and mixed one is very common in most countries not using vaccination against these virus Mixed antibody titer detection is very high and significant in most cases. The study goal was to confirm accuracy of mixed infection with high level in spite of using vaccination to control infection. SDS Page test with ELISA compared using Passive detection of antibody neutralization test and CDV multiclonal epitopes analysis using sera collected from dog and commercial developed monoclonal antibodies in lab admitted. Monoclonal Antibody detection with following vaccination is significantly high than c one virus infection. Compared to passive virus detection.

Keywords: Canine, Distemper. F protein, Mixed, Parvo.

1. INTRODUCTION

Canine parvovirus (CPV) mostly shown clinical gastro enteritis symptoms virus infection—and with different range of clinical signs associated canine distemper virus (CDV) from respiratory, gestor enteritis and neurological disordered of wide canine species with different isolates strain. (Beineke et al., 2009; Goddard and Leisewitz, 2010; Pesavento, 2011; Greene and Decaro, 2012. Big threats from viral infection and wide spread symptomatically in multi-canine species (Beineke et al., 2009; Goddard and Leisewitz, 2010; Steneroden, 2011. Immuno-response and antibody protection against CPV and CDV remarkable and increasing titers in serum following vaccination for solid immunity in susceptible dogs (Rima et al., 1991). The titer of antibodies in serum should be measured for detecting the need for vaccination in regular and routine time (Tizard and Ni, 1998for extra precaution and protection in domesticated and sheltered canines (Newbury et al., 2009; Lecher et al., 2010).

Diagnosis CPV and CDV mainly related to history and clinical signs on different animal age but mainly confirmed in a lab by gene expression of viral detection using RT-PCR (Yuan Zhang et al 2020). level of iral antibody protection related to stages of infection, vaccination and mediated immune system expression of antigen associated cells in animal serum (Jin Oh, 2006). Several serological test for antibody detection specially confirmed monoclonal antibody using western blotting technique and SDS-page with accuracy sensitivity and specifity of serum neutralization tst and Elisa (Crawford, 2010).

The Most common Sero chemical test for IgM, IgG titers in serum for CPV were recently detected using western blott and SDPage for accuracy depending on molecular weight of protein for antibody (Jin -So etal 2006). Other serological test for antibody Elisa compared with indirect fluorescent antibody assays (IFA), Serum neutralization test and Passive hemagglutination inhibition (HI). Level detection of Ig in samples varies from HI ≥1:160 (Carmichael et al., 1983; Twark and Dodds, 2000). CDV titers are measured using ELISA, IFA, and serum neutralization (SN) tests (Tizard and Ni, 1998(add recent reference). Serological test for early antibody titers below expected range can be detected using SN (Gillespie, 1966; Appel, 1969, but following vaccination variability of titer mainly using IFA for a adequate immunity to (Twark and Dodds, 2000, Abosuena 2020).

Easy rapid assay for antibody titer detection and confirmation related to various stages of many viral infection mainly confirmed by Elisa in serum and fecal samples (Carmichael, 2005). Ways of mediated system to confirm viral infection for results expression may be challenged by different test used, cultural laboratory cell related to different manufacture of Vero and mice or invitro of rabbit cells for titration serum variability of antigen and antibody correlation of visibility

ISSN: 2229-7359 Vol. 11 No. 25s,2025

https://theaspd.com/index.php

2. MATERIALS AND METHODS

2.1 Viral cell

Viral cell were obtained and commercially purchased were modified myeloma cell Eagles medium (DMEM, Sanco techno, China) invented with 20% bovine serum (wells science, China) at 37 °C in 5% CO2. Canine distemper virus (CDV) strain Underreport (ATCC account: Ch12345767.2) was propagated in Vero cells using a viral stock with 50% tissue culture infective dose (TCID50) value of 105/mL.

2.2 Propagation and preparation commercial antibody

Selection of positive Mono clonal antibody on 8-week cell culture grown viral cell prorogated up to 250 hemagglutinating units (HAU) with various dilution 1:160 to 1:640 for IgG of CPV (Sago manufacturer, USA). Detection specific monoclonal antibody for CDV for several elution of F protein detection depending on epitopes recognition according to manufacturer instruction, (Scienc tecno-Switzerland)

2.3 Serum samples

Clinically infected dog and asymptotically serum samples was screened using Rapid assay kit for Passive hemagglutination test for viral antibody detection. samples kept in freezer below zero and at _20°C for further processing

2.4 Western blotting technique

CPV, CDV protein detection Western blot assay. Using SDS-PAGE with 10-15% with poly-vinylidene sodium membranes and poly acrylamide for Purification of serum supernatant and blocked with 8-12% non-fatty milk layers to block and clear all surfaces up to 2-6 h. Serial surface washes by Tween PBS 0.01-0.06% variability time. Incubation cell surface membrane of rabbit – poly clonal of Vero cell and mice- monoclonal with serial dilution from 1:160 up to 1:1000 at regular at 4 °C, concurrently. using Tween PBS for clarity and washes of membrane of anti-mouse IgG antibody and anti-rabbit IgG antibody serial dilution (1:1000 up to 1:3000 for titration and degradation of membrane conjugated with antibody variability depending on manufacturer (Boshide, Wuhan, China). Kit Assay of ELISA was used according to the manufacturer's instructions, reporting results for accuracy antibody titer for serum samples and invitro neutralization diagnosis of monoclonal and polyclonal antibody related to different viral infection.

2.5 Statistical methods

Expermintal analysis of Data using spread sheet and 2011 SPSS version of ANOVA analysis 11 windows (SPSS Inc., MN, USA), Qualities and aQuantive approach of experimental design of data analysis.

3. RESULTS

Vero cell in laboratory showed viral protein in different stages of infection with molecular weight variable with each virus. Several studies on Canine parvo virus protein subunit for immunity and vaccination. purification of protein in our study using SDS - page with precipitation at different molecular weight indicate accuracy.

Gene expression using E. coli for Mult surfaces protein in wide scale of laboratory variation compared with specific protein either poly clonal and multiclonal in different surfaces of scales.

Utilization of Vero cell in conjugated protein of virus using mice antibody and rabbit conjugated cell surfaced can excreted rapid easy for viral poly clonal protein detection. Accuracy of check point with protein in verocell detect high sensitivity and specificity of cpv viral infection. methodology method of antibody titer detection in many ways much more accurate in cdv infection with different age of animal with concern of wide spread related diseases. correlated to health measure and post vaccination concern (Lechner et al., 2010). Analysis of molecular weight of CPV protein in SDS- page showed visibility and more accuracy of polyclonal anibody conjugated protein (fig 1)

ISSN: 2229-7359 Vol. 11 No. 25s,2025

https://theaspd.com/index.php

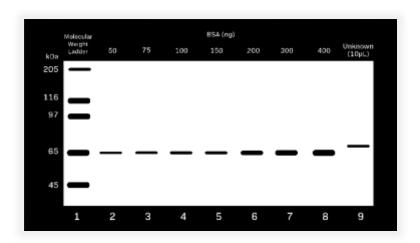


Fig. 1. SDS-page analysis of CPV Protein with different molecular weight

CDV subunit protein of multi clonal antibody showed some accuracy with titration of serial dilution of verocell in antigen visibility of protection in immune mediated system .

Results of CDV in SDS-PAGE a detected of inclusion body proteins indicated the presence of CDV-F protein with a molecular weight of variability 25-55 kDa, as expected (Fig. 2)monoclonal f protein visibility of SDS-PAGE correlated of high accurate detection using Westernblot with epitopes correlated to estimation of anti-CDV antibody of serum analysis showed solid immunity for animals (Fig 1b). Therefore, the purified CDV-F protein was suitable for use as an immunizing antigen to generate mAbsin BALB/c mice.

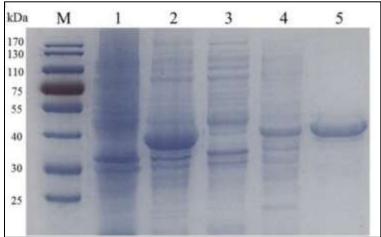


Fig. 2. Analysis of SDS-page of CDV mAbs (monoclonal antibody) molecular weight protein In our previous studies using ELISA showed that sensitivity and specificity of high range for antibody titer sufficient immunity in concurrent protection of animal post infection and after vaccination. (Hassenin et al 2023)

Comparative data analysis for results of experimental diagnosis were compared with variety of different estimation of molecular weight of protein. Western blott estimation for monoclonal and multiclonal protein of CPV and CDV related to period of immunization and infection of different stages and animal ages. sensitivity and specificity of ELISA accuracy related to protein of viral antibody in many titers is very significant to mono clonconal and F protein. Manufacturer of Vero cell, antigen antibody of cell membrane used in lab from different experimental animal surface membrane rabbit and mice conjugate sera play role in accuracy.

4. CONCLUSIONS

F-protein of CPV infection is generated in multi virus infection and cab easily detected by Western blot analysis for variability of mediated immunity post infection and following vaccination at 70kpa.CDV infection with detection of monoclonal antibody F protein from 40-75 kpa with three detection of mAbs of

ISSN: 2229-7359 Vol. 11 No. 25s,2025

https://theaspd.com/index.php

variability of 1A8, 1A12 and 13D 7 with accurate sensitivity and high specificity of Elisa in standard antibody titration with serial dilution best diagnosis of cdv post natural infection and immunization.

Conflict of Interest Statement: SDS Page & western blot kits used in this study confirmed by experimental laboratory diagnostic veterinary medicine. There is no conflict of interest of authors. All shared laboratory diagnosis, writing, publication.

Acknowledgement

This study was supported and helped staff and Scientists at Veterinary Medicine, and the Animal Health Laboratory. Agriculture association.

REFERENCES

- 1. Appel, M.J., "Pathogenesis of canine distemper". American Journal of Veterinary Research vol30, pp1167-1182, Jul 1969.
- 2. Appel, M.J., Robson, D.S., 1973. "A microneutralization test for canine distemper virus." American Journal of Veterinary Research vol 34(11) 1459-1463, Nov1973.
- 3. Beineke, A., Puff, C., Seehusen, F., Baumgartner, W., "Pathogenesis and immunopathology of systemic and nervous canine distemper". Veterinary Immunology and Immunopathology vol 15(127), pp1-18, Jan 2009.
- 4. Carmichael, L.E., Joubert, J.C., Pollock, R.V.H." Hemagglutination by canine parvovirus-2: Serologic studies and diagnostic applications". American Journal of Veterinary Research vol 41(5), 784-791, May 1980.
- Carmichael, L.E., Joubert, J.C., Pollock, R.V." A modified live canine parvovirus vaccine. II. Immune response". Cornell Veterinarian vol 73(1) 13-29. Jan1983.
- 6. Carmichael, L.E. "An annotated historical account of canine parvovirus". Journal of Veterinary Medicine, B. Infectious Diseases and Veterinary Public Health. Vol 52(7-8) 303-311, Sept-Oct2005. DOI: 10.1111/j.1439-0450.2005.00868.x.
- Crawford, C. "Canine and feline parvovirus in animal shelters". In: Proceedings of the Western Veterinary Conference, Las Vegas, NV, USA. 2010.
- 8. Gillespie, J.H. "The significance of passive immunity and the biological tests used in the study of distemper." Journal of the American Veterinary Medical Association vol 1,149{(5) 623-632, Sept 1966.
- Goddard A., Leisewitz A.L" Canine parvovirus". Veterinary Clinics of North America Small Animal Practice vol 40(6), 1041-1053. Nov 2010 Doi: 10.1016/j.cvsm.2010.07.007.
- 10. Greene, C.E., Decaro, N., 2012. Canine viral enteritis. In: Greene, C.E. (Ed.). Infectious Diseases of the Dog and Cat, Fourth Edn. Elsevier Saunders, St. Louis, MO, USA, pp. 67-80.
- 11. Pesavento, P., 2011. Canine diarrhea/2C update. In: Proceedings of the Midwest Veterinary Conference, Columbus, OH, USA.
- 12. Krakowka, S., Olsen, R., Confer, A., Koestner, A., McCullough, B." Serologic response to canine distemper viral antigens in gnotobiotic dogs infected with canine distemper virus". Journal of Infectious Diseases vol132(4), 384-392. Oct1975. doi: 10.1093/infdis/132.4.384.
- 13. Lechner, E.S., Crawford, P.C., Levy, J.K., Edinboro, C.H., Dubovi, E.J., Caligiuri, R., Prevalence of protective antibody titers for canine distemper virus and canine parvovirus in dogs entering a Florida animal shelter. Journal of the American Veterinary Medical Association vol 236(12), 1317-1321. Jun 2010 doi: 10.2460/javma.236.12.1317.
- 14. Newbury, S., Larson, L.J., Schultz, R.D., "Canine distemper virus. In: Miller, L., Hurley, K. (Eds). Infectious Disease Management in Animal Shelters. Wiley-Blackwell, Ames, IA, USA, pp. 161-172, May 2009.
- 15. Noon, K.F., Rogul, M., Binn, L.N., Keefe, T.J., Marchwicki, R.H., Appel, M.J." Enzyme-linked immunosorbent assay for evaluation of antibody to canine distemper virus". American Journal of Veterinary Research vol 41(4), 605-609. April 1980.
- Rima, B.K., Duffy, N., Mitchell, W.J., Summers, B.A., Appel, M.J." Correlation between humoral immune responses and presence of virus in the CNS in dogs experimentally infected with canine distemper virus". Archives of Virology vol 121, 1-8, March 1991.
- 17. Steneroden, K.K., Hill, A.E., Salman, M.D. A needs-assessment and demographic survey of infection-control and disease awareness in western US animal shelters. Preventive Veterinary Medicine vol 98(1), 52-57, Jan 2011. doi: 10.1016/j.prevetmed.2010.11.001.
- 18. Tizard, I., Ni, Y. "Use of serologic testing to assess immune status of companion animals". Journal of the American Veterinary Medical Association vol 213, (1) 54-60, Jul 1998.
- 19. Twark, L., Dodds, W.J. "Clinical use of serum parvovirus and distemper virus antibody titers for determining revaccination strategies in healthy dogs. Journal of the American Veterinary Medical Association vol 217(7), 1021-1024, Oct 2000. doi: 0.2460/javma.2000.217.1021.
- 20. Winters, K.A., Mathes, L.E., Krakowka, S., Olsen, R.G. "Immunoglobulin class response to canine distemper virus in gnotobiotic dogs". Veterinary Immunology and Immunopathology 5(2), 209-215, Dec 1983. doi: 10.1016/0165-2427(83)90022-3.