

Twenty-Eight Selected Phytoconstituents Of *Neptunia Oleracea* (Water Mimosa) As Antioxidant, Antidiabetic And Antimalarial Activities: An *In-Silico* Study

Sarveshkrishna Senthilkumaran¹, Indhumathi Rajagopal Kannan², Radhakrishnan Narayanaswamy^{1*}

¹Department of Biochemistry, Saveetha Medical College and Hospital, Saveetha Institute of Medical and Technical Sciences (Deemed to be University), Saveetha University, Thandalam, Chennai, Tamilnadu-602 105, India. Email ID: ssktassml@gmail.com (SS), nrkishnan@gmail.com

²Department of Community Medicine, Saveetha Medical College and Hospital, Saveetha Institute of Medical and Technical Sciences (Deemed to be University), Saveetha University, Thandalam, Chennai, Tamilnadu-602 105, India. Email ID: rkindhumathi01@gmail.com

Running title: 28 selected phytoconstituents of Water Mimosa as antioxidant, antidiabetic and antimalarial activities: An in silico study

*Corresponding author

Dr. Radhakrishnan Narayanaswamy,
nrkishnan@gmail.com

Abstract

Neptunia oleracea (Water Mimosa) has been known to demonstrate numerous pharmacological activities. In the current investigation, we aimed to assess twenty-eight chosen phytoconstituents of *N. oleracea* (Water Mimosa) as excellent inhibitory agents of human prolidase (hPEPD), human protein tyrosine phosphatase 1B (hPTP 1B) and *Anopheles gambiae* glutathione-S-transferase delta-class (agGSTd1-6) using molecular docking method.

The twenty-eight selected constituents of *N. oleracea* (Water Mimosa) were studied on the docking behaviour of hPEPD, hPTP 1B and agGSTd1-6 by utilizing the Swiss dock approach. The docking analysis showed that quercetin-3-O-rutinoside, narcissoside and quercetin-3, 7-di-O-glucoside of *N. oleracea* (Water Mimosa) has exhibited the maximum binding energy (-9.27, -9.02 and -9.95 kcal/mol) with the hPEPD, hPTP 1B and agGSTd1-6 respectively. Thus, the current result provides a new insight regarding the twenty-eight chosen ligands of *N. oleracea* (Water Mimosa) as excellent inhibitory agents of human prolidase (hPEPD), human protein tyrosine phosphatase 1B (hPTP 1B) and *Anopheles gambiae* glutathione-S-transferase delta-class (agGSTd1-6), which thereby will aid in managing wounds, diabetes mellitus and malaria.

Keywords: *Neptunia oleracea*, Water Mimosa, Prolidase, Protein tyrosine phosphatase 1B, *Anopheles gambiae* glutathione-S-transferase, narcissoside

INTRODUCTION

Neptunia oleracea (Water Mimosa) belongs to Fabaceae (Pea) family and which is native to Africa, Asia and South America [1]. Genus *Neptunia* has been reported to comprise twenty two species (namely *Neptunia amplexicaulis*, *Neptunia dimorphantha*, *Neptunia gracilis*, *Neptunia heliophila*, *Neptunia hispida*, *Neptunia insignis*, *Neptunia javanica*, *Neptunia longipila*, *Neptunia major*, *Neptunia monosperma*, *Neptunia oleracea*, *Neptunia paucijuga*, *Neptunia plena*, *Neptunia proxima*, *Neptunia scutata*, *Neptunia tactilis*, *Neptunia valida*, *Neptunia xanthanema*) and which are distributed in the regions like Africa, Asia, Australia, Malesia, North and South America [2]. Among above said *Neptunia* species, *Neptunia oleracea* (Water Mimosa) is one the most cultivated species particularly for the purpose of human consumption and application [3]. The vernacular names for *Neptunia oleracea* (Water Mimosa) are “Pani najak” in Bengali (India), “Kanhchhnaet” in Cambodia, “Neptunie potegere” in France, “Lajjalu” in Hindi (India), “Kemon” in Indonesia, “Nidrayam” in Kannada, “Neer thottavadi” in Malayalam (India), “Kemon air” in Malaysia, “Eshing ekaithabi” in Manipuri (India), “Juqueri manso” in Portugal, “Alambusha” in Sanskrit (India), “Sundaikerai” in Tamil (India), “Niru thalavapu” in Telugu (India), “Phakkrachet” in Thailand, “Garden

puff" in United States [4, 5]. Different plant parts of *Neptunia oleracea* (Water Mimosa) are traditional used as follows: i) flowers are used to cure earache and syphilis, ii) leaves are used to cure dysentery, earache, intestinal infection and syphilis, iii) leaves, shoot and stem are used to cure uterine infections and discharge, iv) roots are used to treat dysentery, hard palate and necrosis of the nose, v) whole plant is used to cure guinea worm infection and yellow fever [5].

Neptunia oleracea (Water Mimosa) has been reported to exhibit various biological activities such as a) analgesic, b) anti-bacterial, c) anti-cancer, d) anti-inflammatory, e) antimicrobial, f) antioxidant, g) antitumor, h) antiulcer, i) antiviral, j) hepato-protective, k) glucosidase inhibition and l) 5-alpha reductase inhibition [1, 5, 6]. The previous studies prompted us to execute the current study on twenty eight selected *Neptunia oleracea* (Water Mimosa) constituents which includes quercetin-3,7-di-O-glucoside, quercetin-3-O-rutinoside, isoquercetin, quercetin-3-O-xyloside, narcissoside, quercetin-3-O-arabinoside, myricetin, quercetin, vitexin 2"-O-rhamnoside, vitexin, apigenin, salvianolic acid, hydroxytyrosol, leucenine, orientin, kaempferol 7-O-glucoside, kaempferol-3-O-arabinoside, kaempferol-3-O-rhamnoside, kaempferol, catechin, methylgallate, rutin, malic acid, syringic acid, digalloylglucose, caffeic acid, gallic acid and 3,4-O-dimethylgallic acid. These above said *Neptunia oleracea* (Water Mimosa) phytoconstituents were aimed to investigate on the docking analysis of human prolidase (hPEPD), human protein tyrosine phosphatase 1B (hPTP 1B) and *Anopheles gambiae* glutathione-S-transferase delta-class (agGSTd1-6) by utilizing the swissdock method.

METHODOLOGY

Ligand preparation

The chemical structures of twenty eight (Water Mimosa) ligands namely i) quercetin-3,7-di-O-glucoside (CID 13345441); ii) quercetin-3-O-rutinoside (CID 5280805); iii) isoquercetin (CID 5280804); iv) quercetin-3-O-xyloside (CID 5878729); v) narcissoside (CID 5481663); vi) quercetin-3-O-arabinoside (CID 12309865); vii) myricetin (CID 5281672); viii) quercetin (CID 5280343); ix) vitexin 2"-O-rhamnoside (CID 5282151); x) vitexin (CID 5280441); xi) apigenin (CID 5280443); xii) salvianolic acid (CID 5281793); xiii) hydroxytyrosol (CID 82755); xiv) leucenine (CID 440473); xv) orientin (CID 5281675); xvi) kaempferol 7-O-glucoside (CID 10095180); xvii) kaempferol-3-O-arabinoside (CID 5481882); xviii) kaempferol-3-O-rhamnoside (CID 5835713); xix) kaempferol (CID 5280863); xx) catechin (CID 9064); xxi) methylgallate (CID 7428); xxii) rutin (CID 5280805); xxiii) malic acid (CID 525); xxiv) syringic acid (CID 10742); xxv) digalloylglucose (CID 129628549); xxvi) caffeic acid (CID 689043); xxvii) gallic acid (CID 370) and xxviii) 3,4-O-dimethylgallic acid (CID 74709) were downloaded from PubChem (<https://pubchem.ncbi.nlm.nih.gov/>) compound database. The Water Mimosa ligands were prepared by utilizing ChemDraw 2D and 3D (CambridgeSoft, Cambridge, USA) software tools [7]. Thus, these prepared 3-dimensional structures were utilized for further investigations (swissdock).

Preparation of target enzymes

The 3-dimensional structure of human prolidase [hPEPD] (PDB[°] ID: 2IW2 with a resolution of 1.82 Å), human protein tyrosine phosphatase 1B [hPTP 1B] (PDB[°] ID: 2QBS with a resolution of 2.10 Å) and *Anopheles gambiae* glutathione-S-transferase delta-class [agGSTd1-6] (PDB[°] ID: 1PN9 with a resolution of 2.00 Å) was retrieved from [°]Protein Data Bank (PDB). "A" chain of these enzymes were prepared individually by deleting other chains, ligands, and even the crystallographically observed "water" (H₂O) molecules by adopting UCSF Chimera (Regents, University of California, San Francisco, USA) software tool [8].

Docking study

A docking analysis was carried out for twenty-eight chosen phytoconstituents of *Neptunia oleracea* (Water Mimosa) with three target enzymes (hPEPD, hPTP 1B and agGSTd1-6) using the Swissdock (<http://old.swissdock.ch/>) free web server [9]. Finally, PyMOL (Molecular Graphics System, Version 2.4.1, Schrödinger, New York, USA) software was used to analysis the binding site of best-docked pose for each ligand [10, 11].

RESULTS

The current Swissdock analysis showed that quercetin-3-O-rutinoside has shown the maximum binding energy [MBE] (-9.27 kcal per mol) with the human prolidase (hPEPD) enzyme. On the other hand, malic acid has exhibited the lowest binding energy (-6.33 kcal per mol) with the hPEPD enzyme (as shown in Table 1).

In the present investigation ten ligands (myricetin, quercetin, apigenin, salvianolic acid, methylgallate, malic acid, syringic acid, digalloylglucose, caffeic acid and 3,4-O-dimethylgallic acid) have shown interactions with Cys284 amino acid residue of human prolidase (hPEPD) enzyme (as shown in Table 1). Similarly, six ligands (quercetin-3-O-arabinoside, kaempferol 7-O-glucoside, kaempferol-3-O-arabinoside, kaempferol-3-O-rhamnoside, kaempferol and catechin) have shown interactions with Lys111 amino acid residue of hPEPD enzyme.

The current Swissdock analysis showed that narcissoside has exhibited the maximum binding energy [MBE] (-9.02 kcal per mol) with the human protein tyrosine phosphatase 1B (hPTP 1B) enzyme. On the other hand, methylgallate has exhibited the lowest binding energy (-6.14 kcal per mol) with the hPTP 1B second target enzyme (as shown in Table 2).

In the current investigation six ligands (quercetin-3-O-rutinoside, narcissoside, myricetin, kaempferol 7-O-glucoside, kaempferol-3-O-rhamnoside and rutin) have shown interaction with Gln262 amino acid residue of human protein tyrosine phosphatase 1B (hPTP 1B) enzyme (as shown in Table 2). Similarly, five ligands (quercetin-3-O-rutinoside, narcissoside, kaempferol 7-O-glucoside, kaempferol-3-O-rhamnoside and rutin) have shown interaction with Asp48 amino acid residue of hPTP 1B enzyme.

The docking analysis showed that quercetin-3, 7-di-O-glucoside has shown the highest binding energy [HBE] (-9.95 kcal/mol) with the *Anopheles gambiae* glutathione S-transferase delta-class (agGSTd1-6) enzyme. On the other hand, hydroxytyrosol has exhibited the lowest binding energy (-6.17 kcal per mol) with the agGSTd1-6 enzyme (as shown in Table 3).

In the present investigation eight ligands (quercetin-3-O-rutinoside, quercetin-3-O-xyloside, narcissoside, quercetin-3-O-arabinoside, vitexin, orientin, malic acid and digalloylglucose) have shown interaction with Ile52 amino acid residue of *Anopheles gambiae* glutathione S-transferase delta-class (agGSTd1-6) enzyme (as shown in Table 3). Similarly, ten ligands (quercetin-3-O-rutinoside, quercetin-3-O-arabinoside, myricetin, vitexin, apigenin, leucenine, kaempferol, catechin, syringic acid and caffeic acid) have shown interaction with Tyr113 amino acid residue of agGSTd1-6 enzyme.

Discussion

The Water Mimosa (*Neptunia oleracea*) is an edible herb which is cultivated as vegetable in Southeast Asia. And moreover *N. oleracea* (Water Mimosa) has been known to exhibit both anti-oxidant and antidiabetic activities [6]. Furthermore *N. oleracea* (Water Mimosa) has been demonstrated to inhibit enzymes (alpha glucosidase and 5-alpha reductase) activities [12, 13]. Prolidase is a hydrolase enzyme which belongs to matrix metalloproteinase (MMP) family that cleaves both 'imido-peptides and imido-tripeptides' with carboxyl (C) terminal proline (Pro) and hydroxyproline (Hyp) [14]. Prolidase activity has been reported to found in several tissues like amniotic fluid, brain, heart, kidney, liver, pancreas and stomach [15]. Prolidase enzyme activity has been reported to elevate during pathological conditions like i) bipolar disorder, ii) breast cancer, iii) endometrial cancer, iv) erectile dysfunction, v) *Helicobacter pylori* infection, vi) hypertension, vii) keloid scar formation, viii) liver disease, ix) lung cancer, x) melanoma and xi) thalassaemia [16]. Recently, serum prolidase activity has been reported to elevate due to more oxidative stress along with reduced anti-oxidant levels in prostate cancer [17]. Hence prolidase has been chosen as the first target enzyme for the present study. In the present investigation chosen *Neptunia oleracea* (Water Mimosa) ligands have shown interaction with Ser241, Asp376 and Val377 amino acid residues of human prolidase (hPEPD) enzyme. This result showed good agreement with earlier report [18].

In the current investigation one ligand (methylgallate) has shown interaction with Arg45 amino acid residue (AAR) of human protein tyrosine phosphatase 1B (hPTP 1B) enzyme. This result was on par with

previous report, where dodecanoic acid methyl ester has shown interaction with Arg45 amino acid residue of hPTP 1B enzyme [19]. Similarly, chosen *Neptunia oleracea* (Water Mimosa) ligands have shown interaction with Arg24, Tyr46, Asp48 and Gln262 amino acid residues of hPTP 1B enzyme. This result showed good agreement with earlier report [20].

In the current investigation, four ligands (isoquercetin, quercetin-3-O-xyloside, vitexin and orientin) have shown interaction with Ser9 amino acid residue (AAR) of *Anopheles gambiae* glutathione -S-transferase delta-class (agGSTd1-6) enzyme. This result showed good agreement with earlier reports [21, 22, 23, 24]. The findings of the present investigation purely based on *in silico* (molecular docking) method which gives new insight regarding the twenty eight chosen *Neptunia oleracea* (Water Mimosa) constituents and their interactions with three target enzymes namely human prolidase (hPEPD), human protein tyrosine phosphatase 1B (hPTP 1B) and *Anopheles gambiae* glutathione -S-transferase delta-class (agGSTd1-6). However, further *in vitro* biochemical assay, insecticidal potential and detoxifying enzyme inhibition studies are needed to confirm their enzyme inhibition, larvicidal and mosquito repellent activities of chosen phytoconstituents.

CONCLUSION

In the current investigation, the twenty-eight chosen *Neptunia oleracea* (Water Mimosa) phytoconstituents have shown the potential to dock with two targeted human enzymes (hPEPD and hPTP 1B) and one *An. Gambiae* GST target enzyme (agGSTd1-6) respectively. However, five ligands (quercetin, hydroxytyrosol, kaempferol-3-O-arabinoside, kaempferol-3-O-rhamnoside and 3,4-O-dimethylgallic acid) does not show any interaction with amino acid residues of *An. Gambiae* GST (agGSTd1-6) enzyme. Thus, the current results give a new information about the twenty-eight chosen ligands of *N. oleracea* (Water Mimosa) as potent inhibitory agents of hPEPD, hPTP 1B and *Anopheles gambiae* GST (agGSTd1-6), which thereby will help in managing wounds, diabetes mellitus and malaria.

Conflicts of Interest: The authors declare that they have no conflicts of interest.

Acknowledgment: The authors like to thank authorities of Saveetha Medical College and Hospital (SMCH), Saveetha Institute of Medical and Technical Sciences (SIMATS, Chennai) for providing us support to conduct the study.

REFERENCES

1. Sonia Devi O, Nabachandra singh T, Jairaj Singh L, Prasanta Singh T. Biological importance and phytochemistry of *Neptunia oleracea* Lour: A Mini review. Asian J Chem. 2021;33(10): 2276–80. doi:10.14233/ajchem.2021.23424
2. Bean AR. A revision of *Neptunia* Lour.(Leguminosae: subfamily Caesalpinioideae, Mimosoid clade) in Australia and Malesia. Austrobaileya. 2022;12:59-106. doi:10.5962/p.366317.
3. Supriya Yenkokpam, Devi Y. Eshing Ekai Thabi - An Underutilised Aquatic Vegetable of Manipur. Int J Curr Microbiol Appl Sci. 2018;7:1292-96. doi:10.20546/ijcmas.2018.710.145.
4. Singh V, Shah KN, Rana D. Medicinal importance of unexploited vegetable under North Eastern regions of India. J Med Plants Stud. 2015;3(3):33-6.
5. Sagolshemcha R, Singh R. Traditional and biological uses of *Neptunia oleracea* Lour: an overview. Int J Curr Res. 2017;9(6):51689-94.
6. Lee SY, Mediani A, Ismail IS, Maulidiani, Abas F. Antioxidants and α -glucosidase inhibitors from *Neptunia oleracea* fractions using ¹H NMR-based metabolomics approach and UHPLC-MS/MS analysis. BMC Complement Altern Med. 2019;19:1-5. doi:10.1186/s12906-018-2413-4.
7. Kumaraswamy S, Arumugam G, Pandurangan AK, Prabhakaran VS, Narayanaswamy R: Molecular docking analysis of organic acids (OA) from honey as modulators of human ferritin, transferrin, and hepcidin. J Microbiol Biotech Food Sci. 2023;12: e5743. doi:10.55251/jmbfs.5743.
8. Vishnupriya N, Mani Sankar M, Vimal S, Radhakrishnan N. STITCH and molecular docking analysis of selected Wood apple (*Limonia acidissima*) constituents as anti-dandruff and anti-acne agents. J Pharm Bioall Sci. 2024;16:S1167-72. doi:10.4103/jpbs.jpbs_508_23.
9. Grosdidier A, Zoete V, Michielin O. SwissDock, a protein-small molecule docking web service based on EADock DSS. Nucleic Acids Res. 2011;39(2):W270-77.

10. Mohan S, Prabhakaran VS, Narayanaswamy R. 2022. *In silico* analysis of *Cissus rotundifolia* constituents as human neutrophil elastase (HNE), matrix metalloproteinases (MMP 2 and MMP 9), and tyrosinase inhibitors. *Appl Biochem Biotechnol*. 2022;194(1):232-45. doi:10.1007/s12010-021-03758-8.
11. Narayanaswamy R, Prabhakaran VS, Al-Ansari MM, Al-Humaid LA, Tiwari P. An *in silico* analysis of synthetic and natural compounds as inhibitors of nitrous oxide reductase (N2OR) and nitrite reductase (NIR). *Toxics* 2023;11(8):660. doi: 10.3390/toxics11080660.
12. Kumar N, Chaiyasut C. Health promotion potential of vegetables cultivated in northern Thailand: a preliminary screening of tannin and flavonoid contents, 5 α -reductase inhibition, astringent activity, and antioxidant activities. *J Evid Based Complementary Altern Med*. 2017;22(4):573-9. doi: 10.1177/2156587216686689.
13. Lee SY, Abas F, Khatib A, Ismail IS, Shaari K, Zawawi N. Metabolite profiling of *Neptunia oleracea* and correlation with antioxidant and α -glucosidase inhibitory activities using ¹H NMR-based metabolomics. *Phytochem Lett*. 2016; 1;16:23-33. doi: 10.1016/j.phytol.2016.02.014.
14. Isbilen E, Kulaksizoglu S, Kirmiziloglu M, Karuserci Komurcu O, Tabur S. Role of prolidase activity and oxidative stress biomarkers in unexplained infertility. *Int J Gynecol Obstet*. 2022;156(3):430-5. doi: 10.1002/ijgo.13899.
15. Ekinci A, Kamasak K. Evaluation of serum prolidase enzyme activity and oxidative stress in patients with tinnitus. *Braz J Otorhinolaryngol*. 2020;86(4):405-10. doi: 10.1016/j.bjorl.2019.01.009.
16. Kitchener RL, Grunden AM. Prolidase function in proline metabolism and its medical and biotechnological applications. *J Appl Microbiol*. 2012;113(2):233-47. doi: 10.1111/j.1365-2672.2012.05310.x.
17. Kaba M, Pirincci N, Demir H, Verep S. Serum prolidase activity, oxidative stress, and antioxidant enzyme levels in patients with prostate cancer. *Urol Oncol*. 2024;42(4):116.e9-116.e15. doi: 10.1016/j.urolonc.2024.01.007.
18. Wilk P, Uehlein M, Kalms J, Dobbek H, Mueller U, Weiss MS. Substrate specificity and reaction mechanism of human prolidase. *FEBS J*. 2017;284(17):2870-85. doi: 10.1111/febs.14158.
19. Srinivasan K, Altemimi AB, Narayanaswamy R, Vasantha Srinivasan P, Najm MA, Mahna N. GC-MS, alpha-amylase, and alpha-glucosidase inhibition and molecular docking analysis of selected phytoconstituents of small wild date palm fruit (*Phoenix pusilla*). *Food Sci Nutr*. 2023;11(9):5304-17. doi: 10.1002/fsn3.3489.
20. Patel AD, Pasha TY, Lunagariya P, Shah U, Bhambharoliya T, Tripathi RK. A library of thiazolidin-4-one derivatives as protein tyrosine phosphatase 1B (PTP1B) inhibitors: an attempt to discover novel antidiabetic agents. *Chem Med Chem*. 2020 ;15(13):1229-42. doi: 10.1002/cmdc.202000055.
21. Chen L, Hall PR, Zhou XE, Ranson H, Hemingway J, Meehan EJ. Structure of an insect δ -class glutathione S-transferase from a DDT-resistant strain of the malaria vector *Anopheles gambiae*. *Acta Crystallogr D: Biol Crystallogr*. 2003;59(12):2211-17. doi: 10.1107/s0907444903018493.
22. Koirala BKS, Moural T, Zhu F. Functional and Structural Diversity of Insect Glutathione S-transferases in Xenobiotic Adaptation. *Int J Biol Sci*. 2022;18(15):5713. doi: 10.7150/ijbs.77141.
23. Awad M, Alfuhaid NA, Amer A, Hassan NN, Moustafa MA. Towards Sustainable Pest Management: Toxicity, Biochemical Effects, and Molecular Docking Analysis of *Ocimum basilicum* (Lamiaceae) Essential Oil on *Agrotis ipsilon* and *Spodoptera littoralis* (Lepidoptera: Noctuidae). *Neotrop Entomol*. 2024;53(3):669-81. doi: 10.1007/s13744-024-01137-6.
24. Vasantha-Srinivasan P, Shanmuga-Priya S, Han YS, Radhakrishnan N, Karthi S, Elsadek MF, Mustafa AEZM, Senthil-Nathan S. Phyto-chemical screening, insecticidal potential and detoxifying enzyme inhibition of *Ficus auriculata* (Lour.) extracts, against the mosquito vector and non-target aquatic predator. *Biocatal Agric Biotechnol*. 2023;53:102864. doi: 10.1016/j.bcab.2023.102864.

Table 1 The Swissdock binding energy (SDBE) analysis of 28 selected *Neptunia oleracea* (Water Mimosa) ligands with the human prolidase (hPEPD) enzyme using Swissdock method

S.no	Ligand name	SDBE [■] (-kcal per mol)	Interactions of (AAR) amino acids residues	BD [■] (Å°)
1	Quercetin-3,7-di-O-glucoside	9.19	Glu53	2.0 and 3.2
			Arg67	3.3
			Glu228	3.0
			His239	3.0
			Thr243	2.9 and 3.4
			Glu281	1.8
			Val377	3.3
2	Quercetin-3-O-rutinoside	9.27	Glu53	2.8

			Thr55	3.4
			Leu65	2.0
			Glu69	1.9
			Ser70	3.4
			Asn152	3.3
			Asp376	2.5 and 2.6
3	Isoquercetin	8.10	Glu54	2.2 and 3.0
4	Quercetin-3-O-xyloside	8.44	Glu54	2.1, 2.2 and 2.8
			Lys111	2.2
5	Narcissoside	9.18	Thr55	3.4
			Leu65	3.2
			Arg67	3.2
			Glu228	1.9
			Ser240	3.1 and 3.4
			Asp376	2.0
6	Quercetin-3-O-arabinoside	8.46	Glu54	2.1, 2.2 and 2.8
			Lys111	2.2
7	Myricetin	7.26	Tyr232	3.2
			Arg238	3.3
			Glu281	2.3 and 2.5
			Cys284	3.2
8	Quercetin	7.20	Thr79	2.2
			Gly236	2.3
			Arg238	3.5
			Cys284	3.3
9	Vitexin 2"-O-rhamnoside	7.11	Arg67	2.6
			Glu69	1.8
			Ser240	2.2
10	Vitexin	8.03	Leu65	2.4 and 3.0
			Arg67	3.1
			Ser70	3.2 and 3.3
			Ser241	1.9
			Glu281	1.9, 2.2 and 2.5
11	Apigenin	7.00	Gln68	3.4
			Arg238	3.3
			Cys284	3.2
12	Salvianolic acid	8.12	Thr79	3.3
			Gly236	2.8
			Arg238	3.5 and 3.5
			Glu281	2.7
			Tyr283	3.0
			Cys284	3.1
13	Hydroxytyrosol	7.17	Gln68	3.4
			Thr79	2.6

			Arg238	3.5
			Glu281	2.1
			Tyr283	3.4
14	Leucenine	6.89	Phe66	2.7
			Gln68	3.0
			Thr79	2.5
			Gly236	2.5 and 3.1
			Arg238	3.4
			Glu281	3.2
			Tyr283	3.2
15	Orientin	8.43	Leu65	2.3 and 3.1
			Arg67	3.1
			Glu228	1.8 and 1.9
			Ser240	3.5
			Ser241	2.7
			Glu281	1.9, 2.1 and 2.7
16	Kaempferol 7-O-glucoside	9.12	Glu54	2.3 and 2.9
			Gln68	3.0
			Lys111	2.1
			Arg238	3.1
17	Kaempferol-3-O-arabinoside	8.40	Glu54	2.1, 2.2 and 2.8
			Lys111	2.2
18	Kaempferol-3-O-rhamnoside	8.60	Glu54	2.4 and 2.9
			Lys111	2.3
19	Kaempferol	7.21	Glu54	1.8
			Thr79	3.3
			Lys111	1.9
20	Catechin	7.38	Glu54	2.0
			Lys111	2.0
21	Methylgallate	6.92	Gly236	2.5
			Arg238	3.2
			Glu281	2.3 and 2.4
			Thr283	3.5
			Cys284	3.2
22	Rutin	8.42	Glu53	2.3
			Arg67	3.5
			Ser241	2.7
			Glu281	2.3
23	Malic acid	6.33	Gly236	1.9 and 3.5
			Arg238	3.3
			Glu281	2.2
			Tyr283	3.0
			Cys284	3.2

24	Syringic acid	6.99	Gln68	3.0
			Thr79	2.6
			Tyr283	3.5
			Cys284	3.2
25	Digalloylglucose	8.30	Thr79	3.2
			Tyr232	3.0
			Gly236	2.1
			Arg238	3.0, 3.1 and 3.3
			Tyr283	3.2 and 3.4
			Cys284	3.2
26	Caffeic acid	7.05	Phe66	3.2
			Gln68	3.0
			Thr79	2.4 and 2.9
			Arg238	3.3
			Glu281	2.2 and 2.3
			Tyr283	3.4
			Cys284	3.5
27	Gallic acid	6.63	Gly236	2.1
			Arg238	3.3
			Glu281	2.2 and 2.3
			Tyr283	3.5
28	3,4-O-dimethylgallic acid	6.93	Thr79	2.6
			Glu281	2.0
			Tyr283	3.1
			Cys284	3.4

Note: SDBE[■] - Swissdock binding energy analysis, BD[■] - Bond distance.

Table 2 The Swissdock binding energy (SDBE) analysis of 28 chosen *Neptunia oleracea* (Water Mimosa) ligands with the human protein tyrosine phosphatase 1B (hPTP 1B) enzyme using Swissdock method

S.no	Ligand name	SDBE [■] (kcal per mol)	Interactions of (AAR) amino acids residues	BD [■] (Å)
1	Quercetin-3,7-di-O-glucoside	8.62	Lys73	3.0 and 3.6
			Gln78	2.6
			Ser80	3.2
			Gln102	1.8
2	Quercetin-3-O-rutinoside	8.88	Arg24	3.2
			Tyr46	3.3
			Asp48	2.5
			Ser118	3.4
			Gln262	3.3 and 3.4
3	Isoquercetin	7.96	Gln78	3.2 and 3.4

			His208	2.0
4	Quercetin-3-O-xyloside	7.75	Leu204	1.8
			Pro206	2.2
5	Narcissoside	9.02	Arg24	3.1 and 3.3
			Thr46	3.3
			Asp48	3.3
			Gln262	3.2 and 3.3
6	Quercetin-3-O-arabinoside	7.79	Leu204	1.8
			Pro206	2.1
7	Myricetin	7.08	Arg24	3.5
			Tyr46	3.4
			Gln262	3.4 and 3.5
8	Quercetin	6.91	Leu204	1.9
9	Vitexin 2"-O-rhamnoside	8.09	Arg47	2.9, 3.2 and 3.3
10	Vitexin	7.63	Leu204	2.1
			Pro206	1.9
11	Apigenin	6.83	No interactions	-
12	Salvianolic acid	7.74	Gln78	3.0
			Arg79	3.0 and 3.1
			Ser80	3.0
			Leu204	2.6
13	Hydroxytyrosol	6.26	Leu204	2.0
			Pro206	2.7
			Gly209	2.5
14	Leucenine	6.45	Asp29	1.9 and 3.2
			Arg254	3.2 and 3.3
15	Orientin	8.09	Leu204	2.2
			Glu207	2.3
			Gly209	2.5
16	Kaempferol 7-O-glucoside	7.99	Arg24	3.3
			Tyr46	3.4
			Asp48	2.2
			Lys120	3.1
			Gln262	3.3
17	Kaempferol-3-O-arabinoside	7.46	Gln78	3.3
			Pro206	2.0
18	Kaempferol-3-O-rhamnoside	7.15	Arg47	3.2
			Asp48	3.0
			Gln262	2.3
19	Kaempferol	6.77	Gln78	3.3
			Ser80	2.9
			Leu204	2.4
20	Catechin	7.55	Gln102	2.1
			Leu204	2.3

			Pro206	2.3
			Gly209	2.4
21	Methylgallate	6.14	Arg45	3.1 and 3.2
			Leu119	3.4
22	Rutin	8.70	Arg24	3.2 and 3.6
			Tyr46	3.4
			Arg47	3.1
			Asp48	3.1
			Gln262	3.2 and 3.3
23	Malic acid	6.48	Ser28	1.9
			Arg254	3.1 and 3.3
			Gly259	3.1
24	Syringic acid	6.57	Glu2	1.8 and 3.4
25	Digalloylglucose	7.63	Gln78	2.0 and 3.3
			Ser80	3.0
			Ser205	2.7
			His208	2.2
26	Caffeic acid	6.36	Leu204	2.1
			Gly209	2.5
27	Gallic acid	6.30	Pro89	1.9
			Glu136	3.4
			Asp137	2.0 and 3.4
28	3,4-O-dimethylgallic acid	6.56	Glu2	1.8 and 3.4

Note: SDBE[■] - Swissdock binding energy analysis, BD[■] - Bond distance.

Table 3 The Swissdock binding energy (SDBE) analysis of 28 chosen *Neptunia oleracea* (Water Mimosa) ligands with the *Anopheles gambiae* glutathione -S-transferase delta-class (agGSTd1-6) enzyme using Swissdock method

S.no	Ligand name	SDBE [■] (-kcal per mol)	Interactions of (AAR) amino acids residues	BD [■] (Å)
1	Quercetin-3,7-di-O-glucoside	9.95	Glu64	1.9
			Ser65	2.9 and 3.2
			Asp110	1.8
2	Quercetin-3-O-rutinoside	9.11	Gly8	2.0
			His38	3.4
			Ile52	2.8 and 3.2
			Tyr113	3.1
3	Isoquercetin	8.33	Ser9	3.1
			Glu64	1.9
			Gln106	3.2
4	Quercetin-3-O-xyloside	8.08	Ser9	3.3
			Ile52	1.9 and 3.2
5	Narcissoside	8.93	Gly8	2.4
			Ile52	2.4

			Gln106	3.5
6	Quercetin-3-O-arabinoside	7.81	Gly8	2.7
			Ile52	2.0
			Tyr113	3.3
7	Myricetin	7.05	Gly8	2.3
			Tyr113	3.5
8	Quercetin	7.05	No interactions	-
9	Vitexin 2"-O-rhamnoside	8.59	Glu64	3.4
10	Vitexin	8.14	Ser9	3.3 and 3.5
			Ile52	2.6
			Tyr105	2.9
			Tyr113	3.3
11	Apigenin	6.85	Tyr113	3.2
12	Salvianolic acid	8.60	Asp110	1.9
13	Hydroxytyrosol	6.17	No interactions	-
14	Leucenine	6.38	Gly8	2.2
			Tyr105	3.3
			Tyr113	3.4
15	Orientin	6.56	Ser9	3.1 and 3.4
			Gln49	3.3
			Ile52	1.9
			Pro53	2.6
			Glu64	2.2 and 3.0
			Ser65	3.0 and 3.1
			Arg66	3.4
16	Kaempferol 7-O-glucoside	9.05	Glu64	1.9, 2.0 and 2.5
			Arg66	3.2
17	Kaempferol-3-O-arabinoside	7.71	No interactions	-
18	Kaempferol-3-O-rhamnoside	8.01	No interactions	-
19	Kaempferol	7.07	Tyr113	3.2
20	Catechin	6.89	Gly8	2.3
			Tyr113	3.5
21	Methylgallate	6.18	Gly8	2.5
22	Rutin	9.06	Glu64	2.5
			Ser65	3.2 and 3.3
			Tyr105	3.2 and 3.5
23	Malic acid	6.54	Ile52	2.6
			Glu64	1.8
			Ser65	3.1 and 3.4
			Arg66	3.3, 3.3 and 3.5
24	Syringic acid	6.26	Tyr105	3.1 and 3.1
			Tyr113	3.5
25	Digalloylglucose	8.18	Ser9	3.2 and 3.5
			Ile52	3.0

			Glu64	2.4
			Ser65	3.5
			Tyr105	3.0 and 3.2
26	Caffeic acid	6.18	Tyr113	3.5
27	Gallic acid	6.47	Glu64	1.8
28	3,4-O-dimethylgallic acid	6.30	No interactions	-

Note: SDBE[■] - Swissdock binding energy analysis, BD[■]- Bond distance.